

Veteriner Hekimlikte Güncel Yaklaşımlar

Editör

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ÖNSÖZ

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Bölüm 1

BÜYÜKBAŞ HAYVAN KAYNAKLI BAKTERİYEL ZOONOZ HASTALIKLAR: MULTİDİSİPLİNER BİR DEĞERLENDİRİLME

Sema İŞİSAĞ ÜÇÜNCÜ¹

Burak GÖKÇE²

Bu derlemede büyükbaş hayvanlardan kaynaklanan ve özellikle veteriner hekimler ile teknisyenleri için ciddi sağlık sorunları oluşturabilen bakteriyel zoonozlar, son on yıla ağırlık verilerek gözden geçirilen literatur bilgileri ışığında ve iş sağlığı-güvenliği yaklaşımıyla ele alınmış, böylece farklı disiplinlerde konuya ilgi duyan araştırmacılar için genel bir başvuru kaynağı oluşturulması hedeflenmiştir.

Çok geniş bir tanımla zoonozlar, diğer hayvanlardan ve insanlardan insanlara geçerek çeşitli enfeksiyon hastalıkları oluşturan, biyolojik açıdan da kendini çoğaltabilen patojen proteinler olan prionlar, ayrıca virusler, bakteriler, funguslar, mikro ve makro parazitler olarak farklı sistematik kategorilerde incelenen organizmalardır. Bilinen insan patojenlerinin % 60'ından fazlasının, ayrıca ilk kez görülen ya da yeniden ortaya çıkan bulaşıcı hastalıkların %75'inin etmeni olarak kaydedilirler ⁽¹⁾. Görülme sıklıkları, yayılma hızları, tanımlanmış olup olmamaları, insan ve hayvan sağlığı üzerindeki etkileri ve ekonomik etkileri olarak belirlenen beş ayrı ölçütle değerlendirilirler ⁽²⁾. Dünya genelinde

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kullanım oranları da ilk grupta yine %1.2 (1 kişi), ikinci grupta %1.9 (12 kişi) olarak bildirilmektedir ⁽¹¹³⁾. Bir diğer araştırma kapsamında 120 laborant ve veteriner sağlık öğrencisine uygulanan ankette, zoonotik hastalıklar hakkındaki farkındalığın, İş Sağlığı ve Güvenliği dersini başarıyla bitirenlerde dersi almayanlara göre %17.14 daha yüksek olduğu görülmüştür ⁽⁴²⁾. Veteriner hekimlikte iş sağlığı ve güvenliğine yönelik bazı lisansüstü bitirme projeleri de mevcuttur ⁽⁴⁴⁾. Bu tür çalışmaların çoğalmasıyla veri tabanının metaanaliz yapılabilecek genişliğe ulaşması beklenmektedir.

İnsan ve toplum sağlığını korumak, iş sağlığı ve güvenliği kültürünü yerleştirip geliştirmek için, veteriner hekimlik özelinde ve konuyla ilgili diğer iş kollarında yoğun ve sürekli eğitime, ayrıca çok sıkı denetime ihtiyaç olduğu açıktır. Bu gereksinimin giderilmesinde disiplinler arası çalışmalar ve konuya farklı açılardan getirilecek yaklaşımlar halk sağlığı ve çalışanların sağlığı anlamında büyük önem taşımaktadır. Ülkemizde iş sağlığı ve güvenliği alanında bilgi birikimi arttıkça yukarıda kaydedilenlere benzer yöntemlerle yapılacak çalışmaların da çoğalması umulmaktadır. Bu konuda üniversitelere, Sağlık Bakanlığına, Veteriner Hekimler Odalarına ve Türk Veteriner Hekimleri Birliğine büyük görev düşmektedir.

KAYNAKLAR

1. Taylor LH, Latham SM, Woolhouse ME. Risk factors for human disease emergence. *Philosophical Transactions of the Royal Society of London*, 2001;356: 983–989.
2. Institute of Medicine and National Research Council, 2009. Sustaining global surveillance and response to emerging zoonotic diseases, summary. In: G. T. Keusch, M. Papaioanou, M.C. Gonzalez, K.A. Scott, and P. Tsai (eds), pp. 1–2. The National Academies Press, Washington, DC.
3. Shanko K, Kemal J, Kenea D, A review on confronting zoonoses: the role of veterinarian and physician. *Veterinar Science and Technology*, 2015;6(2): 1-7.

4. Dewey-Mattia D, Manikonda, K, Hall AJ, et al. Surveillance for foodborne disease outbreaks-United States, 2009–2015. *MMWR Surveill Summ.* 2018; 67(No. SS-10):1–11. DOI: <http://dx.doi.org/10.15585/mmwr.ss6710a1external icon>
5. CDC (Centers for Disease Control and Prevention), Surveillance for foodborne disease outbreaks, United States, 2016, Annual Report. Atlanta, Georgia: U.S. Department of Health and Human Services, CDC, 2018. https://www.cdc.gov/fdoss/pdf/2016_FoodBorneOutbreaks_508.pdf. Erişim Tarihi: 06.08.2019.
6. Christie L, Glaser C, Zoonoses diseases. Global Health Education Consortium, 2007. https://www.cugh.org/sites/default/files/63_Zoonoses_Diseases_FINAL_0.pdf (Erişim Tarihi: 25.12.2018).
7. Mills JN, Gage KL, Khan AS, Potential influence of climate change on vector-borne and zoonotic diseases: a review and proposed research plan. *Environmental Health Perspectives*, 2010;118(11): 11: 1507-1514.
8. Lal A, Hales S, French N, et al, Seasonality in human zoonotic enteric diseases: a systematic review. *PLOS ONE* 2012;7(4): e31883. <https://doi.org/10.1371/journal.pone.0031883>.
9. Karesh WB, Dobson A, Lloyd-Smith JO, et al, Ecology of zoonoses: natural and unnatural histories. *Lancet*, 2012;380: 1936–1945.
10. Kilpatrick AM & Randolph SE, Drivers, dynamics, and control of emerging vector-borne zoonotic diseases. *Lancet*, 2012;380: 1946–1955.
11. Morse SM, Mazet JAK, Woolhouse M, et al, Prediction and prevention of the next pandemic zoonosis. *Lancet*, 2012;380: 1956–1965.
12. Bidaisee S, & Macpherson, CN, Zoonoses and one health: a review of the literature. *Journal of Parasitology Researches*, 2014;874345. doi:10.1155/2014/874345.
13. Bliven KA, & Maurelli AT, Evolution of bacterial pathogens within the human host. *Microbiology Spectrum*. 2016;4(1)- DOI:10.1128/microbiolspec.VMBF-0017- 2015.
14. Erol İ, Yeni ve yeniden önem kazanan gıda kaynaklı bakteriyel zoonozların epidemiyolojisi, *Veteriner Hekim Dergisi*, 2016;87(2): 63-76.
15. Plowright RK, Parrish CR, McCallum H, et al, Pathways to zoonotic spillover. *Nature Reviews Microbiology*, 2017;15: 502–510.
16. Bird BH & Mazet JAK, Detection of emerging zoonotic pathogens: an integrated one health approach. *Annual Review of Animal Biosciences*, 2018;6:121-139. <https://doi.org/10.1146/annurev-animal-030117-014628>
17. Rabinowitz P, & Conti L, Links among human health, animal health, and ecosystem health. *Annual Review of Public Health*, 2013;34:189-204 <https://doi.org/10.1146/annurev-publhealth-031912-114426>
18. Kahn LH, Kaplan B, Steele JH, Confronting zoonoses through closer collaboration between medicine and veterinary medicine (as ‘one medicine’). *Veterinaria Italiana*, 2007;43(1): 5-19.

19. Calistri P, Iannetti S, Danzetta ML, et al, The components of 'one world - one health' approach. *Transboundary and Emerging Diseases*, 2013;60 (Supl 2):4-13.
20. Cantas L, & Suer K, Review: The important bacterial zoonoses in "one health" concept. *Frontiers in Public Health*, 2014;2: 144.
21. Baker WS, & Gray GC, A review of published reports regarding zoonotic pathogen infection in veterinarians. *American Veterinary Medicine Association*, 2009;15;234(10):1271-1278. doi: 10.2460/javma.234.10.1271.
22. Epp T, & Waldner C, Occupational health hazards in veterinary medicine: zoonoses and other biological hazards. *Canadian Veterinary Journal*, 2012;53(2): 144-150.
23. van Metre DC, & Morley PS, Zoonotic diseases-fostering awareness in critical audiences. *Journal of Extension*, 2015;53(6) <https://joe.org/joe/2015december/a7.php>
24. Sánchez A, Prats-van der Ham M, Tatay-Dualde J, et al., Zoonoses in veterinary students: a systematic review of the literature. *plos ONE* 2017;12(1)
25. Une Y, & Mori T, Tuberculosis as a zoonosis from a veterinary perspective. *Comparative Immunology, Microbiology and Infectious Diseases*, 2007;30(5-6):415-425.
26. Kumar HBC, Lokesh KM, Madhavaprasad CB, et al, occupational zoonoses in zoo and wildlife veterinarians in India: A Review. *Veterinary World*, 2013;6(9): 605-613, doi: 10.5455/vetworld.2013.605-613.
27. Ariful Islam M, Zoonoses in Bangladesh: the role of veterinarian in public health. *Bangladesh Journal of Veterinary Medicine*, 2014;12 (2): 93-98.
28. Helmy YA, El-Adawy H, Abdelwhab EM, A comprehensive review of common bacterial, parasitic and viral zoonoses at the human-animal interface in Egypt. *Pathogens*, 2017;6(3), 33; <https://doi.org/10.3390/pathogens6030033>
29. Asante J, Noreddin A, El Zowalaty ME, Systematic review of important bacterial zoonoses in Africa in the last decade in light of the 'one health' concept. *Pathogens*, 2019;8(2): 50; <https://doi.org/10.3390/pathogens8020050>
30. Çelebi B, Babür C, Kılıç S, & ark, Zoonotik enfeksiyonlardan Q ateşi, listerioz, toksoplazmoz ve kistik ekinokokkoz'un risk grubunda seroprevalansının araştırılması. *Türk Hijyen ve Deneysel Biyoloji Dergisi*, 2008;65 (2): 67-73.
31. Özbey G, Kalender H, Muz A, Sığır tüberkülozu'nun epidemiyolojisi ve teşhisi, *Fırat Üniversitesi Sağlık Bilimleri Dergisi*, 2008;22(5):307-314.
32. Aslan G, Kuyucu N, Çalıkoğlu M, & ark., *Mycobacterium bovis*'in etken olduğu tüberküloz olguları, *ANKEM Dergisi*, 2009;23(4):182-187.
33. Yıldırım T, Şimşek F, Çelebi B, & ark, Ağır sarılıkla başvuran nadir görülen bir akut q ateşi olgusu ve literatüre bakış. *Klinik Dergisi*, 2010;23(3): 124-129.
34. Aydın N, Hayvanlarda brucellosis ve epidemiyolojisi, 2011. https://www.klimik.org.tr/wp-content/uploads/2012/02/1282011123712-3A_Bruselloz_Slaydlari.pdf (Erişim Tarihi: 25.12.2018)

35. Ögütlü A, Şarbon, *Deneyisel ve Klinik Tıp Dergisi*, 2012;29:155-162
36. Yeşilbaş Z, & Kader Ç, Deri şarbonu. *Klinik Dergisi*, 2012;27(3): 114-117.
37. Kul B, Tüberkülozis, 2013. <http://www.veteriner.cc/hasta/tuberkiloz.asp> (Erişim Tarihi 22.12.2018)
38. Çaylak SD, Sözen H, Aksözek A, & ark., Muğla ilinde veteriner hekimlerde *Brucella* Seropozitifliği, *Klinik Dergisi*, 2015;28(3): 108-111.
39. Özkol HU, Karadaş S, Sünnetçioğlu M, & ark, Yatırılarak takip edilen 63 deri şarbonu olgusunun klinik ve laboratuvar bulgularının değerlendirilmesi: Türkiye'de deri şarbonunun karakteristiği, *Turkish Archives of Dermatology and Venereology, Türkderm*, 2014;48: 197-203.
40. Öztürkler Y, Veteriner hekimlerin güvenlik rehberi, *Çiftlik Dergisi*, 2015. http://www.turkvet.biz/yazi/mp_vet_guvenlik.htm (Erişim Tarihi: 26.12.2018)
41. Aslım G, & Yaşar A, Resmi veteriner hekimlerin iş sağlığı ve güvenliğinin değerlendirilmesi. 2017. <http://iohsc2017.org/upload/67bf8b4a.pdf> (Erişim Tarihi: 19.12.2018).
42. Kayabaşı R, Laborant ve veteriner sağlık önlisans öğrencilerinin zoonotik meslek hastalıkları hakkında bilinç seviyelerinin ölçülmesi, *Electronic Journal of Vocational Colleges*, Ağustos 2018;11-20.
43. Yılmaz O, & Oymak S, Zoonozlarda mücadelede veteriner halk sağlığı ve tek sağlık, *Veteriner Halk Sağlığı Yayını*, 2018, Yayın No:2018/1,81pp.
44. Çınar E, Büyükbaş hayvan kaynaklı zoonotik hastalıkların veteriner hekimlikte iş sağlığı ve güvenliği açısından incelenmesi. 2019. Tezsiz yüksek lisans bitirme tez projesi Ege Üniversitesi Fen Bilimleri Enstitüsü İş Güvenliği Anabilim Dalı.
45. McDaniel CJ, Cardwell DM, Moeller RB, et al, Humans and cattle: a review of bovine zoonoses. *Vector-Borne Zoonotic Diseases*, 2014;14:1–19. doi: 10.1089/vbz.2012.1164.
46. Rycroft AN, & Garside LH, Review: *Actinobacillus* species and their role in animal disease *The Veterinary Journal*, 2000;159: 18–36.
47. Christensen H, & Bisgaard M, Revised definition of *Actinobacillus sensu stricto* isolated from animals: A review with special emphasis on diagnosis. *Veterinary Microbiology*, 2014;99(1): 13-30.
48. Yoshimura H, Kojima A, Ishimaru M, Antimicrobial susceptibility of *Arcanobacterium pyogenes* isolated from cattle and pigs. *Zoonoses and Public Health*, 2001;47(2): 139-143
49. Yassin AF, Hupfer H, Siering C, et al, Comparative chemotaxonomic and phylogenetic studies on the genus *Arcanobacterium* Collins *et al.* 1982 emend. Lehnen *et al.* 2006: proposal for *Trueperella* gen. nov. and emended description of the genus *Arcanobacterium*. *International Journal of Systematic and Evolutionary Microbiology*, 2011;61:1265–1274.
50. Turnbull PCB, Introduction: anthrax history, disease and ecology. *Current Topics in Microbiology and Immunology*, 2002;271:1-19.

51. Turnbull PCB, & Shadomy SV, Anthrax from 500 BC to AD 2010. In: *Bacillus anthracis* and anthrax. Bergman NH, Ed. New Jersey: WileyBlackwell, 2011;1-15.
52. Doğanay M, & Demiraslan H, Human anthrax as a re-emerging disease. *Recent Patents on Anti-Infective Drug Discovery*, 2015;10(1): 10-29.
53. Goel AK, Anthrax: A disease of biowarfare and public health importance. *World Journal of Clinical Cases*, 2015;3(1):20-33. doi:10.12998/wjcc.v3.i1.20
54. Brisson D, Drecktrah D, Eggers CH, et al, Genetics of *Borrelia burgdorferi*. *The Annual Review of Genetics*, 2012;46:515-536. doi:10.1146/annurev-genet-011112-112140
55. Dumler JS, Molecular diagnosis of lyme disease: review and meta-analysis. *Molecular Diagnosis*, 2001;6(1): 1-11.
56. Lebech AM, Polymerase chain reaction in diagnosis of *Borrelia burgdorferi* infections and studies on taxonomic classification. *APMIS Suppl.* 2002;(105):1-40.
57. Marques A, Chronic lyme disease: an appraisal. *Infectious Disease Clinics of North America*, 2008;22(2):341-360. doi: 10.1016/j.idc.2007.12.011
58. Biesiada G, Czepiel J, Leśniak MR, et al, Lyme disease: review. *Archives of Medical Science*, 2012;8(6):978-982. doi:10.5114/aoms.2012.30948.
59. Köksal İ, Lyme hastalığı: tanı ve tedavi, *Türkiye Klinikleri Journal of Infectious Disease, Special Topics*. 2014;7(2):48-51.
60. Steere AC, Strle F, Wormser GP, et al, Lyme borreliosis. *Nature Reviews Disease Primers*, 2, Article number: 16090 (2016).
61. Wills MKB, Kirby AM, Lloyd VK, Detecting the lyme disease spirochete, *Borrelia burgdorferi*, in ticks using nested PCR. *The Journal of Visualized Experiments*, 2018;(132), e56471, doi:10.3791/56471
62. Zvizdić Š, Čengić D, Maja Bratić M, *Brucella melitensis* review of the human infection case. *Bosnian Journal of Basic Medical Sciences*, 2006;6(1): 15-18.
63. Hemarajata P, Baghdadi JD, Hoffman R, et al, *Burkholderia pseudomallei*: challenges for the clinical microbiology laboratory. *Journal of Clinical Microbiology*, 2016;54(12): 2866-2873; DOI: 10.1128/JCM.01636-16
64. Perumal Samy R, Stiles BG, Sethi G, et al, Melioidosis: clinical impact and public health threat in the tropics. *PLOS Neglected Tropical Diseases*, 2017;11(5):e0004738. doi:10.1371/journal.pntd.0004738
65. Silva J, Leite D, Fernandes M, et al, *Campylobacter* spp. as a foodborne pathogen: a review. *Frontiers in Microbiology*, 2011;2:200. Published 2011 Sep 27. doi:10.3389/fmicb.2011.00200
66. Wagenaar JA, van Bergen MA, Blaser MJ, et al, *Campylobacter fetus* infections in humans: exposure and disease. *Clinical Infectious Diseases*, 2014;58(11):1579-86. doi: 10.1093/cid/ciu085. Epub 2014 Feb 18.
67. Gazonne L, Legrand P, Renaud B, et al, *Campylobacter fetus* bloodstream infection: risk factors and clinical features, *European Journal of Clinical Microbiology & Infectious Diseases*, 2008;27: 185-189.

68. Nagano N, Isomine S, Kato H, et al, Human fulminant gas gangrene caused by *Clostridium chauvoei*. *Journal of Clinical Microbiology*, 2008;46(4), 1545-1547.
69. Rychener L, Albon S, Djordjevic SP, et al, *Clostridium chauvoei*, an evolutionary dead-end pathogen. *Frontiers in Microbiology*, 2017;8, 1054. doi:10.3389/fmicb.2017.01054.
70. van Roeden SE, Wever PC, Kampschreur LM et al, Chronic Q fever-related complications and mortality: data from a nationwide cohort. *Clinical Microbiology and Infection*, 2019; 25(11):1390-1398. pii: S1198-743X(18)30766-3. doi: 10.1016/j.cmi.2018.11.023.
71. Giuffrida R, Dermatoflose. In: Megid, J.; Riberio, M.G.; Paes, A.C. (Eds.). *Doenças Infecciosas em Animais de Produção e de Companhia*. Rio de Janeiro: Roca, 2016;81-86.
72. Canpolat N, Hemolytic uremic syndrome. *Türk Pediatri Arşivi*, 2015;50(2):73–82. doi:10.5152/tpa.2015.2297
73. Rasko DA, Webster DR, Sahl JW, et al, Origins of the *E. coli* strain causing an outbreak of hemolytic-uremic syndrome in Germany. *The New England Journal of Medicine*, 2011;365(8):709–717. doi:10.1056/NEJMoa1106920
74. Ko H, Maymani H, Rojas-Hernandez C, Hemolytic uremic syndrome associated with *Escherichia coli* O157:H7 infection in older adults: a case report and review of the literature. *Journal of Medical Case Reports*, 2016;10:175. doi:10.1186/s13256-016-0970-z
75. Kitchens CS, Purpura and other hematovascular disorders. In: Consultative Hemostasis and Thrombosis. (Fourth Edition). 2019. Eds. C S. Kitchens, C.M. Kessler, B.A. Konkle, M.B. Streiff, D.A. Garcia,
76. Sam C, Desai P, Laber D, et al, Pegylated bovine carboxyhaemoglobin utilisation in a thrombotic thrombocytopenic purpura patient. *Transfusion Medicine, Official Journal of the British Blood Transfusion Society*, 2017. <https://doi.org/10.1111/tme.12407>
77. Evangelista KV & Coburn J, *Leptospira* as an emerging pathogen: a review of its biology, pathogenesis and host immune responses. *Future Microbiology*, 2010;5(9):1413–1425. doi:10.2217/fmb.10.102
78. Costa F, Hagan JE, Calcagno J, et al, Global morbidity and mortality of leptospirosis: a systematic review. *PLoS Negl Trop Dis*. 2015;17;9(9):e0003898. doi: 10.1371/journal.pntd.0003898.
79. De Brito T, Silva AMGD, Abreu PAE, Pathology and pathogenesis of human leptospirosis: a commented review. *The Revista do Instituto de Medicina Tropical de São Paulo* 2018. 60:e23. doi:10.1590/s1678-9946201860023
80. Gasanov U, Hughes D, Hansbro PM, Methods for the isolation and identification of *Listeria* spp. and *Listeria monocytogenes*: a review. *FEMS Microbiology Reviews*, 2005;29(5): 851–875.
81. Scobie A, Kanagarajah S, Ross J, Harris RJ, Mortality risk factors for listeriosis-A 10 year review of non-pregnancy associated cases in England 2006–2015. *Journal of Infection*, 2019;78(3): 208-214.

82. Pai M, Behr MA, Dowdy D, et al, Tuberculosis. *Nature Reviews Disease Primers*, 2016, volume 2, Article number: 16076.
83. Chai Q, Zhang Y, Liu CH, *Mycobacterium tuberculosis*: an adaptable pathogen associated with multiple human diseases. *Frontiers in Cellular and Infection Microbiology*, 2018;8:158. doi:10.3389/fcimb.2018.00158
84. EFSA, The community summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in the European Union in 2008. *EFSA J.* 2010;8, 1496–1906.
85. Eng SK, Pusparajah P, Ab Mutalib NS, et al, Salmonella: a review on pathogenesis, epidemiology and antibiotic resistance. *Frontiers in Life Science*, 2015;8(3):284–93.
86. Jennison AV, & Verma NK, *Shigella flexneri* infection: pathogenesis and vaccine development. *FEMS Microbiology Reviews*, 2004;28(1):43–58.
87. Ballal M, Trends in antimicrobial resistance among enteric pathogens. In: Antibiotic Resistance, Chapter 4, Academic Press, 2016. pp.63–92. <https://doi.org/10.1016/C2015-0-00104-6>
88. Bhattacharya SK, & Sur D, Shigellosis. In: International Encyclopedia of Public Health (Second Edition), Ed. in-Chief S. R. Quah, Assoc.Ed.-in-Chief: W.C.. Cockerham, Elsevier, 2017.
89. Çakıcı N, Demirel-Zorba NN, Akçalı A, Gıda endüstrisi çalışanları ve stafülokokal gıda zehirlenmeleri. *Türk Hijyen ve Deneysel Biyoloji Dergisi*, 2015;72(4): 337–350.
90. Tong SYC, Davis JS, Eichenberger E, et al., *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. *Clinical Microbiology Reviews*, 2015; 28(3):603–61. doi:10.1128/CMR.00134-14
91. Turner NA, Sharma-Kuinkel BK, Maskarinec SA et al, Methicillin-resistant *Staphylococcus aureus*: an overview of basic and clinical research. *Nature Reviews Microbiology*, 2019;17:203–218.
92. Fox A, Streptococci groups A, B, D and others *Enterococcus faecalis*, In: Bacteriology – Microbiology and Immunology On-line, Chapter Twelve, Edited by R. Hunt, 2016. <http://www.microbiologybook.org/fox/streptococci.htm>
93. Stevens DL, & Bryant A, Group A streptococcus: virulence factors and pathogenic mechanisms, 2019. UpToDate, <https://www.uptodate.com/contents/group-a-streptococcus-virulence-factors-and-pathogenic-mechanisms>. Erişim Tarihi: 15.08.2019.
94. Weiser JN, Ferreira DM, Paton JC, *Streptococcus pneumoniae*: transmission, colonization and invasion. *Nature Reviews Microbiology*, 2018;16: 355–367.
95. Darwish SF, Asfour HAE, Allam HA, Incidence of *Yersinia enterocolitica* and *Yersinia pseudotuberculosis* in raw milk samples of different animal species using conventional and molecular methods. *Alexandria Journal of Veterinary Sciences*, 2015;44:174–185.

96. Tauxe RV, Clinical manifestations and diagnosis of *Yersinia* infections, 2019. UpToDate, <https://www.uptodate.com/contents/clinical-manifestations-and-diagnosis-of-yersinia-infections>. Erişim Tarihi: 15.08.2019.
97. Devriese LA, Vancanneyt M, Baele M, et al, *Staphylococcus pseudintermedius* sp. nov., a coagulase-positive species from animals. *International Journal of Systematic and Evolutionary Microbiology*, 2005;55:1569-1573.
98. Donegan S, Bellamy R, Gamble CL, Vaccines for preventing anthrax. *Cochrane Database of Systematic Reviews*, 2009, Issue 2. Art. No.: CD006403.DOI: 10.1002/14651858.CD006403.pub 2.
99. Ruiz S & Wolfe DN, Vaccination against Q fever for biodefense and public health indications. *Frontiers in Microbiology*, 2014;5:726. Published 2014 Dec 16. doi:10.3389/fmicb.2014.00726
100. Veterinary Medicine, 11th Edition, Biosecurity and infection control, Eds. P.D. Constable, K.W. Hinchcliff, S.H. Done, W. Grünberg. Elsevier, 2017. pp. 36-42.
101. Ceyhan İ, Biyogüvenlik laboratuvar seviyeleri ve biyogüvenlik kabinlerinin seçimi kullanımı ve bakımı. 4. *Ulusal Sterilizasyon Dezenfeksiyon Kongresi Bildiri Kitabı*, 2005. pp.608-633.
102. Wright JG, Jung S, Holman RC, et al, Infection control practices and zoonotic disease risks among veterinarians in the United States. *Journal of the American Veterinary Medical Association*, 2008;232(12): 1863-1872.
103. AVMA Personal protective equipment, 2019. <https://www.avma.org/KB/Resources/Reference/Pages/personal-protective-equipment.aspx>; Erişim Tarihi: 30.01.2019.
104. Lucas M, Injuries in Australian veterinarian. Ph. D. Thesis, University of Western Australia, 2012. pp. 85-115.
105. Bharwana SA, Shafaqat A, Abbas F, et al, Risks of zoonoses in veterinarians and infection control practices. *Greener Journal of Biological Sciences*, 2012;2 (1):007-012.
106. Bushra FM, O El, Effect of health education in the promotion of knowledge, attitude and practice of veterinary workers towards prevention of brucellosis infection in Al Ain area, United Arab Emirates, Eastern Region – 2012. MSc Thesis, University of Gezira, 100 pp.
107. Heinrich ERE, Risk of zoonotic pathogen exposure among veterinary professionals and students at veterinary schools and best practices to minimize this risk on individual and institutional levels. M.Sc. Thesis, Department of Clinical Sciences College of Veterinary Medicine Kansas State University, 2014, 61pp.
108. Anderson MEC, & Weese JS, Self-reported hand hygiene perceptions and barriers among companion animal veterinary clinic personnel in Ontario, Canada. *The Canadian Veterinary Journal*, 2016;57(3): 282–288.
109. Robin C, Bettiridge J, McMasteri F, Zoonotic disease risk perceptions in the British veterinary profession. *Preventive Veterinary Medicine*, 2017;136: 39-48.

110. Vinks T, Infection control in a veterinary hospital by improving adherence to hand hygiene protocols through personal feedback on plate counts and with special attention to resistant flora and their origin (microbial resistance genes). Research Project December '16 – February 2017. University of Liverpool, Leahurst campus.
111. Huertas PS, León EA, Tarabla HD, Zoonosis and veterinary waste disposal in rural practice. *Revista Argentina de Microbiología*, 2018;51(3): 251-254, <https://doi.org/10.1016/j.ram.2018.08.004>
112. Rood KA, & Pate ML, Assessment of musculoskeletal injuries associated with palpation, infection control practices, and zoonotic disease risks among Utah clinical veterinarians, *Journal of Agromedicine*, 2019;24:135-145, DOI: 10.1080/1059924X.2018.1536574
113. Kutlu M, Ergönül Ö, Sayın-Kutlu S, et al, Risk factors for occupational brucellosis among veterinary personnel in Turkey. *Preventive Veterinary Medicine*, 2014;117(1): 52-58

Bölüm 2

İNEKLERDE PERİPARTURİENT DÖNEMDE SİTOKİN DÜZEYLERİNE ETKİ EDEN FAKTÖRLER

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GİRİŞ

Periparturient dönem (Geçiş dönemi, transition period), doğumun 3 hafta öncesi ve 3 hafta sonrasını içine alan, sağlık, üretim ve süt ineklerinin karlılığı açısından kritik öneme sahip dönem olarak adlandırılmaktadır ⁽¹⁾.

Süt sentezinin başlaması ve düşük yem tüketimiyle birlikte enerji ve besin ihtiyacının artması, periparturient dönemdeki ineklerde Negatif Enerji Dengesi (NED) ve mikro besin eksikliklerinin ortaya çıkmasına sebep olur. Negatif enerji dengesi, ineklerin vücut yağının esterleşmemiş yağ asitleri (NEFA) ve sonra da Beta-Hidroksibütirik Asite (BHBA) dönüşmesine ve kanda birikmesine sebep olur. Her ne kadar bu değişiklikler yüksek süt verimli ineklerde normal bir süreç olsa da inekler bu metabolik zorluğa uyum sağlayamazsa, metabolik ve bulaşıcı hastalıklar oluşmaya başlar. Bu durum da periparturient dönem ve sonrasında üretkenlik ve üreme verimliliğinin düşmesine yol açar. Periparturient dönemdeki ineklerin biyolojisinin anlaşılmasındaki

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KAYNAKLAR

1. Drackley JK. Biology of dairy cows during the transition period: The final frontier? J Dairy Sci. 1999;82(11):2259-73.
2. Ametaj BN, Hosseini A, Odhiambo JF, et al. (2011). Application of acute phase proteins for monitoring inflammatory states in cattle. In: Veas F(Editors). Acute Phase Proteins as Early Non-Specific Biomarkers of Human and Veterinary Diseases (299-354). InTech: US.
3. Herdt TH, Ruminant adaptation to negative energy balance: Influences on the etiology of ketosis and fatty liver. Vet Clin North Am Food Anim Pract. 2000;16(2):215-30.
4. Bobe G, Young JW, Beitz DC, Invited review: Pathology, etiology, prevention, and treatment of fatty liver in dairy cows. J Dairy Sci. 2004;87(10):3105-24.
5. Sordillo LM. Factors affecting mammary gland immunity and mastitis susceptibility. Livest Prod Sci. 2005;98(1):89-99.
6. Dembic Z (2015). The cytokines of the immune system: The Role of Cytokines in Disease Related to Immune Response, New York: Elsevier.
7. Goff JP, Horst RL. Physiological changes at parturition and their relationship to metabolic disorders. J Dairy Sci. 1997;80(7):1260-8.
8. Lucy MC. Nutrient Partitioning and Reproductive Performance in Dairy Cows. Proceedings of Intermountain Nutrition Conference. Syracuse, NY, 2008: 139-50.
9. White HM. The role of TCA cycle anaplerosis in ketosis and fatty liver in periparturient dairy cows. Animals. 2015;5(3):793-802.
10. Chaouat G. Reproductive immunology: reassessing the Th1/Th2 paradigm? Immunol Lett. 2004;92(3):207-14.
11. Koch CA, Platt JL. T cell recognition and immunity in the fetus and mother. Cell Immunol. 2007;248(1):12-7.
12. Kwak-Kim JY, Gilman-Sachs A, Kim CE. T helper 1 and 2 immune responses in relationship to pregnancy, nonpregnancy, recurrent spontaneous abortions and infertility of repeated implantation failures. Chem Immunol Allergy. 2005;88:64-79.
13. Redline RW. The structural basis of Maternal fetal immune interactions in the human placenta. Curr Top Microbiol Immunol. 1997;222:25-44.
14. Redman CW, McMichael AJ, Stirrat GM, Sunderland CA, Ting A. Class I major histocompatibility complex antigens on human extravillous trophoblast. Immunology. 1984;52(3):457-68.
15. Shomer B, Toder V, Egorov I, Ehrlich R. Expression of allogeneic MHC class I antigens by transgenic mouse trophoblast does not interfere with the normal course of pregnancy. Transgenic Res. 1998;7(5):343-55.

16. Raghupathy R. Th1-type immunity is in compatible with successful pregnancy. *Immunol Today*. 1997;8(10):478-82.
17. Johansson M, Bromfield JJ, Jasper MJ, Robertson SA. Semen activates the female immune response during early pregnancy in mice. *Immunol*. 2004;112(2):290-300.
18. Makhseed M, Raghupathy R, El-Shazly S, et al. Proinflammatory maternal cytokine profile in preterm delivery. *Am J Reprod Immunol*. 2003;49(5):308-18.
19. Bennett WA, Lagoo-Deenadayalan S, Stopple JA, et al. Cytokine expression by first trimester human chorionic villi. *Am J Reprod Immunol*. 1998;40(5):309-18.
20. Raghupathy R. Pregnancy: success and failure within the Th1/Th2/Th3 paradigm. *Semin Immunol*. 2001;13(4):219-27.
21. Genc, MR, Onderdonk A, Witkin SS. Innate immune system gene polymorphisms in women with vulvovaginal infections. *Curr Infect Dis Rep*. 2004;6:462-8.
22. Elenkov IJ, Chrousos GP. Stress hormones, proinflammatory and antiinflammatory cytokines, and autoimmunity *Ann N Y Acad Sci*. 2002;966:290-303.
23. Dudley DJ. (2000). Cytokines in preterm and term parturition. In: Hill JA, editor. *Cytokines in human Reproduction* (171-99). New York: Wiley-Liss.
24. Keelan JA, Blumenstein M, Helliwell RJA, et al. Cytokines, Prostaglandins and Parturition—A Review. *Placenta*. 2003;17:33-46.
25. Kelly RW, King AE, Critchley HOD. Inflammatory mediators and endometrial function: focus on the perivascular cell. *J Reprod Immunol*. 2002;57:81-93.
26. Sabatini M, Boyce B, Aufdemorte T, Bonewald L, Mundy GR. Infusions of recombinant human interleukins 1 α and 1 β cause hypercalcemia in normal mice. *Proc Natl Acad Sci USA*. 1988;85:5235-9.
27. Tribe RM, Moriarty P, Dalrymple A, Hassoni AA, Posten L. Interleukin-1 β induces calcium transients and enhances basal and store operated calcium entry in human myometrial smooth muscle. *Biol Reprod*. 2003;68:1842-9.
28. Osman I, Young A, Ledingham MA, et al. Leukocyte density and pro-inflammatory cytokine expression in human fetal membranes, decidua, cervix and myometrium before and during labour at term. *Mol Hum Reprod*. 2003;9:41-5.
29. Rauk PN, Friebe-Hoffman U, Winebrenner LD, Chao JP. Interleukin-6 up-regulates the oxytocin receptor in cultured uterine muscle cells. *Am J Reprod Immunol*. 2001;45:148-53.
30. Keelan JA, Blumenstein M, Helliwell RJA, et al. Cytokines, Prostaglandins and Parturition—A Review. *Placenta*. 2003;17:33-46.

31. Fitzgerald KA, O'Neil LAJ, Gearing AJH. (2001). The cytokines factbook (2nd ed.) New York: Academy Press.
32. Dunlevy JR, Couchman JR. Interleukin-8 induces motile behaviors and loss of focal adhesions in primary fibroblasts. *J Cell Sci.* 1995;108:311-21.
33. Fortunato SJ, Menon R, Lombardi SJ. Role of tumor necrosis factor- α in the premature rupture of membranes and PTL pathways. *Am J Obstet Gynecol.* 2002; 187: 1159-62.
34. Pfeffer K. Biological functions of tumor necrosis factor cytokines and their receptors. *Cytokine Growth Factor Rev.* 2003;14:185-91.
35. Sagol S, Sagol O, Ozkal S, Asema U. Role of apoptosis, bcl- 2 and bax protein expression in premature rupture of fetal membranes. *Reprod Med.* 2002;47:809-15.
36. Hanna N, Hanna I, Hleb M, et al. Gestational age-dependent expression of IL-10 and its receptor in human placental tissues and isolated cytotrophoblasts. *J Immunol.* 2000;164:5721-8.
37. Xu W, Presnell SR, Parrish-Novak J, et al. A soluble class II cytokine receptor, IL-22RA2, is a naturally occurring IL-22 antagonist. *Proc Natl Acad Sci SA.* 2001;98:9511-6.
38. Gruenberg BH, Schoenemeyer A, Weiss B, et al. A novel, soluble homologue of the human IL-10 receptor with preferential expression in placenta. *Genes Immun.* 2001;2:329-34.
39. Bry K, Hallman M. Transforming growth factor-beta opposes the stimulatory effects of interleukin-1 and tumor necrosis factor on amnion cell prostaglandin E2 production: Implication for preterm labor. *Am J Obstet Gynecol.* 1992;167:222-6.
40. Dudley DJ, Edwin SS, Mitchell MD. Macrophage inflammatory protein-1-alpha regulates prostaglandin E(2) and interleukin-6 production by human gestational tissues in vitro. *J Soc Gynecol Invest.* 1996;3:12-6.
41. Simpson KL, Keelan JA, Mitchell MD. Labour-associated changes in the regulation of production of immunomodulators in human amnion by glucocorticoids, bacterial lipopolysaccharide and pro-inflammatory cytokines. *J Reprod Fertil.* 1999;116.
42. Corwin EJ, Bozoky I, Pugh LC, Johnston N. Interleukin-1 β elevation during the postpartum period. *Ann Behav Med.* 2003;25(1):41-7.
43. Vassiliadis S, Ranella A, Papadimitriou L, Makrygiannakis A, Athanassakis I. Serum levels of pro- and anti-inflammatory cytokines in non-pregnant women, during pregnancy, labour and abortion. *Mediators Inflamm.* 1998;7:69-72.
44. Bessler H, Straussberg R, Hart J, Notti I, Sirota L. Human colostrum stimulates cytokine production. *Biol Neonate.* 1996;69:376-82.

Bölüm 3

MEME SAĞLIĞI ÜZERİNE VİRÜSLERİN ETKİLERİ

Tarık ŞAFAK¹

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GİRİŞ

Oldukça maliyetli hastalıklardan biri olan mastitis süt veriminin düşmesi, tedavi masraflarının yüksek olması, enfekte sütlerin dökülmesi, sürüden hayvanın çıkarılma oranının fazla olması, nüks oranının yüksek ^(1,2) ve reproduktif performansın düşmesine neden olduğu için süt sığırcılığı işletmelerinde oldukça önemli bir yere sahiptir ⁽³⁾. Mastitisin etiyolojisine bakıldığında Tablo 1'de de gösterildiği gibi bakteri, virüs, maya gibi birçok enfeksiyöz ajan etkili olmaktadır. Bunun yanı sıra fiziksel travmalar ve hayvana bağlı faktörlerin de mastitislerin oluşmasında etkisi vardır. Mastitisin oluşmasında en önemli etkenler bakteriyel ajanlar olmasına rağmen, meme sağlığı üzerinde oldukça önemli bir yer tutan virüsler de göz ardı edilmemelidir ^(4,5).

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rın da dahil olmasıyla dolaylı yoldan mastitise neden olurlar. Sığır enterovirüs de süten izole edilen ve süt somatik hücre sayısını arttıran diğer bir etkidir ^(4,6).

SONUÇ

Mastitis, süt sığırcılığında ciddi ekonomik kayıplara neden olmaktadır. Mastitisin etiyolojisi üzerine yapılan araştırmalar bakteriyolojik yönde yoğunlaşmaktadır. Ancak, yapılan araştırmalar göstermektedir ki, bakteriyolojik kaynaklı olmayan ve nedeni tam olarak bilinmeyen mastitislerde vardır. Nedeni bilinmeyen mastitislerde virüslerin de büyük pay sahibi olduğu unutulmamalıdır. Virüsler sığırlarda meme sağlığı açısından doğrudan bir mastitis oluşturmaya da, immün sistemi baskılamak gibi diğer sistemler üzerine ciddi şekilde etki ederek ve memede lezyonlar oluşturarak primer mastitis etkeni olan bakteriyel ajanlar için mastitise zemin oluşturmak kaydıyla ciddi ekonomik kayıplara neden olmaktadır. Bu etkenlerin tedavisinin olmaması da göz önüne alınarak, etkenlere karşı mücadelede koruyucu hekimlik esastır ve gerekli koruma-kontrol programları yapılmalıdır. Ayrıca, koruma-kontrol programları kapsamında immün sistem güçlendirici uygulamalar yapılmalıdır.

KAYNAKLAR

1. Viguier C, Arora S, Gilmartin N, et al. Mastitis detection: current trends and future perspectives. *Trends in Biotechnology*. 2009; 27 (8), 486-493. Doi: 10.1016/j.tibtech.2009.05.004
2. Awale MM, Dudhatra GB, Avinash K, et al. Bovine mastitis: a threat to economy. *Open Access Scientific Reports*. 2012; 1: 295. Doi: 10.1016/j.tibtech.2009.05.004
3. Kumar N, Manimaran A, Kumaresan A, et al. Mastitis effects on reproductive performance in dairy animals: a review. *Tropical Animal Health and Production*. 2017; 49 (4), 663-673. Doi: 10.1007/s11250-017-1253-4.
4. Alpaya G, Yeşilbağ K. Mastitis olgularında virüslerin rolü. *Uludağ Üniversitesi Veteriner Fakültesi Dergisi*. 2009; 28 (1), 39-46.

5. Kalińska A, Wójcik A, Slószar J, et al. Occurrence and aetiology of Staphylococcal mastitis – a review. *Animal Science Papers and Reports*. 2018; 36 (3), 263-73.
6. Wellenberg GJ, Van der Poel WHM, Van Oirschot JT. Viral infections and bovine mastitis: a review. *Veterinary Microbiology*. 2002a; 88 (1), 27-45. Doi: 10.1016/s0378-1135(02)00098-6
7. Yılmaz Z, Yeşilbaş K. Clinical and haematological findings in bovine immunodeficiency virus (BIV) infected cattle. *Turkish Journal of Veterinary and Animal Sciences*. 2008; 32 (3), 207-214.
8. Della Libera AMMP, de Souza FN, Batista CF, et al. Effects of bovine leukemia virus infection on milk neutrophil function and the milk lymphocyte profile. *Veterinary Research*. 2015; 46 (1), 1-8. Doi: 10.1186/s13567-014-0125-4
9. Shafer-Weaver KA, Corl CM, Sordillo LM. Shifts in bovine CD4+ subpopulations increase T-helper-2 compared with T-helper-1 effector cells during the postpartum period. *Journal of Dairy Science*. 1999; 82 (8), 1696-706. Doi: 10.3168/jds.S0022-0302(99)75399-3
10. Kakinuma S, Maeda Y, Ohtsuka H, et al. Bovine leukemia virus titer and leukocyte population associated with mastitis in periparturient dairy cows. *The International Journal of Applied Research in Veterinary Medicine*. 2014; 12 (3), 239-44.
11. Majumder S, Ramakrishnan MA, Nandi S. Infectious bovine rhinotracheitis: an Indian perspective. *International Journal of Current Microbiology and Applied Sciences*. 2015; 4 (10), 844-58.
12. Straub OC, (1990). Herpesviridae. Dinter Z, Morein B, (Eds), *Virus Infections of Ruminants* in (p. 69- 158). New York: Elsevier.
13. Guy JS, Potgieter LN, McCracken M, et al. Isolation of bovine herpesvirus-1 from vesicular lesions of the bovine udder. *American Journal of Veterinary Research*. 1984; 45 (4), 783-85.
14. Rola JG, Larska M, Grzeszuk M, et al. Association between antibody status to bovine herpesvirus-1 and quality of milk in dairy herds in Poland. *Journal of Dairy Science*. 2015; 98 (2), 781-9. Doi: 10.3168/jds.2014-8781
15. Herlekar DA, Shashikant CS, Gurjar AA, et al. Presence of viral and bacterial organisms in milk and their association with somatic cell counts. *Journal of Dairy Science*. 2013; 96 (10), 6336-46. Doi: 10.3168/jds.2013-6631
16. Bilge Dagalp S. Kan ve süt serumlarında IBR/IPV antikorlarının nötralizasyon testi ile saptanması ve süt örneklerinden virüs izolasyonu. *Ankara Üniversitesi Veteriner Fakültesi Dergisi*. 1998; 45, 313-21.
17. Corner AH, Greig AS, Hill DP. A histological study of the effects of the herpes virus of infectious bovine rhinotracheitis in the lactating bovine mammary gland. *Canadian Journal of Comparative Medicine and Veterinary Science*. 1967; 31 (12), 320-30.

18. Kemp R, Holliman A, Nettleton PF. Atypical bovine herpes mammillitis affecting cows and calves. *The Veterinary Record*. 2008; 163 (4), 119-21. Doi: 10.1136/vr.163.4.119
19. Janett F, Stäuber N, Schraner E, et al. Bovine herpes mammillitis: clinical symptoms and serologic course. *Schweizer Archiv für Tierheilkunde*. 2000; 142 (7), 375-80.
20. Hillerton, JE., Morgan, WF., Fransworth, R. et al. (2001). Evaluation of bovine teat condition in commercial dairy herds. *Proceedings 2th International symposium on mastitis and milk quality*. 13-15 September, Canada, (pp. 352-356).
21. Gibbs EPJ. Viral diseases of the skin of the bovine teat and udder. *Veterinary Clinics of North America: Large Animal Practice*. 1984; 6 (1), 187-202. Doi: 10.1016/s0196-9846(17)30047-2
22. Rişvanlı A, Doğan H, Şafak T, Öcal H. (2019). Memenin Savunma Sistemi: Meme Savunmasında Meme Başı ve Meme Başı Kanalının Rolü. Halis Öcal (Ed.), *İneklerde Mastitis Dışındaki Meme, Meme Başı ve Meme Derisinin Hastalıkları* içinde (s. 1-10). Ankara: Türkiye Klinikleri.
23. De-Giuli L, Magnino S, Vigo PG, et al. Development of polymerase chain reaction and restriction typing assay for the diagnosis of bovine herpesvirus 1, bovine herpesvirus 2, and bovine herpesvirus 4 infections. *Journal of Veterinary Diagnostic Investigation*. 2002; 14 (4), 353-6. Doi: 10.1177/104063870201400417
24. Campos FS, Franco AC, Oliveira MT, et al. Detection of bovine herpesvirus 2 and bovine herpesvirus 4 DNA in trigeminal ganglia of naturally infected cattle by polymerase chain reaction. *Veterinary Microbiology*. 2014; 171 (1-2), 182-8. Doi: 10.1016/j.vetmic.2014.03.012
25. Egyed L. Bovine herpesvirus type 4: a special herpesvirus. *Acta veterinaria Hungarica*. 2000; 48 (4), 501-13. Doi: 10.1556/004.48.2000.4.13
26. Izumi Y, Tsuduku S, Murakami K, et al. Characterization of bovine herpesvirus type 4 isolated from cattle with mastitis and subclinical infection by the virus among cattle. *The Journal of Veterinary Medical Science*. 2006; 68 (2), 189-93. Doi: 10.1292/jvms.68.189
27. Morán PE, Pérez SE, Odeón AC, et al. Herpesvirus bovino-4 (BoHV-4): aspectos generales de su biología y situación en la República Argentina. *Revista Argentina de Microbiología*. 2015; 47 (2), 155-66. Doi: org/10.1016/j.ram.2015.02.007
28. Donofrio G, Flammini CF, Scatozza F, et al. Detection of bovine herpesvirus 4 (BoHV-4) DNA in the cell fraction of milk of dairy cattle with history of BoHV-4 infection. *Journal of Clinical Microbiology*. 2000; 38 (12), 4668-71.
29. Wellenberg GJ, van der Poel WH, van der Vorst TJ, et al. Bovine herpesvirus 4 in bovine clinical mastitis. *The Veterinary Record*. 2000; 147 (8), 222-5. Doi: 10.1136/vr.147.8.222

30. Wellenberg GJ, Bruschke CJ, Wisselink HJ, et al. Simultaneous intrammary and intranasal inoculation of lactating cows with bovine herpesvirus 4 induced subclinical mastitis. *Veterinary microbiology*. 2002b; 86 (1-2), 115-29. Doi: 10.1016/s0378-1135(01)00496-5
31. Wellenberg GJ, Verstraten ERAM, Belak S, et al. Detection of bovine herpesvirus 4 glycoprotein B and thymidine kinase DNA by PCR assay in bovine milk. *Journal of Virological Methods*. 2001; 97 (1-2), 101-12. Doi: 10.1016/s0166-0934(01)00341-x
32. Zadoks RN, Allore HG, Barkema HW, et al. Cow and quarter level risk factors for *Streptococcus uberis* and *Staphylococcus aureus* mastitis. *Journal of Dairy Science*. 2001; 84 (12), 2649-63. Doi: 10.3168/jds.s0022-0302(01)74719-4
33. Kalman D, Janosi S, Egyed L. Role of bovine herpesvirus 4 in bacterial bovine mastitis. *Microbial Pathogenesis*. 2004; 37 (3), 125-9. Doi: 10.1016/j.micpath.2004.06.011
34. Ali H, Keefe GP, Cepica A. Bovine herpesvirus-4, a potential cause of mastitis in canadian dairy cows. *British Journal of Dairy Sciences*. 2011; 2 (3), 31-4.
35. Rışvanlı A, Kalkan C. İneklerde meme papillomatozisi ile mastitis arasındaki ilişki. *Veteriner Bilimler Dergisi*. 2001; 17 (3), 143-7.
36. Bilge Dağalp S, Dogan F, Farzan TA, et al. The genetic diversity of bovine papillomaviruses (BPV) from different papillomatosis cases in dairy cows in Turkey. *Archives of Virology*. 2017; 162 (6), 1507-18. Doi: 10.1007/s00705-017-3258-8
37. Doğan H, Aydın M, Öcal H. (2019). Meme ve Meme Başı Derisinde Viral Etkenlere Bağlı Şekillenlen Bozukluklar. Halis Öcal (Ed.), *İneklerde Mastitis Dışındaki Meme, Meme Başı ve Meme Derisinin Hastalıkları* içinde (s. 17-28). Ankara: Türkiye Klinikleri.
38. Santos EUD, Silva MAR, Pontes NE, et al. Detectin of diffrent bovine papillomavirus types and co-infectin in bloodstream of catte. *Transboundary and Emerging Diseases*. 2016; 63 (1), 103-8. Doi: 10.1111/tbed.12237
39. Lindsey CL, Almeida ME, Vicari CF, et al. Bovine papillomavirus DNA in milk, blood, urine, semen and spermatozoa of bovine papillomavirus-infected animals. *Genetics and Molecular Research*. 2009; 8 (1), 310-8. Doi: 10.4238/vol8-1gmr573
40. Hamad MA, Al-Shammari AM, Odisho SM, et al. Molecular and phylogeneti analysis of bovine papillomavirus type 1: First report in Iraqi catte. *Advances in Virology*. 2016, Article ID:2143024. Doi: 10.1155/2016/2143024
41. Öcal H, Yıldız A, Apaydın AM, et al. İnek ve düvelerde meme papillomatozisinin tedavisi ve etiyojisi üzerine çalışma. *F.Ü. Sağlık Bilimler Dergisi*. 2001; 15 (1), 23-6.

42. Apaydın AM, Aydın M, Ciftçi O, et al. Effects of *Paronychia kurdica* on teat and udder papillomatosis in cows. *Revue de Médecine Vétérinaire*. 2010; 161 (6), 267-71.
43. Terziev G, Roydev R, Kalkanov I, et al. Papillomatosis in heifers-comparative studies on surgical excision and autogenous vaccine therapies. *Trakia Journal of Sciences*. 2015; 13 (2), 274-9.
44. Kawakami Y, Kaji T, Kume T, et al. Infection of cattle with parainfluenza 3 virus with special reference to udder infection: I. virus isolation from milk. *Japanese Journal of Microbiology*. 1966; 10 (3), 159-69. Doi: 10.1111/j.1348-0421.1966.tb00304.x
45. Çomaklı S, Özdemir S. Comparative evaluation of the immune responses in cattle mammary tissues naturally infected with bovine parainfluenza virus type 3 and bovine alphaherpesvirus-1. *Pathogens*. 2019; 8 (1), 26. Doi: 10.3390/pathogens8010026
46. Waage S. Influence of new infection with bovine virus diarrhoea virus on udder health in Norwegian dairy cows. *Preventive Veterinary Medicine*. 2000; 43 (2), 123-35. Doi: 10.1016/s0167-5877(99)00084-7
47. Berends IGMA, Swart WAJM, Frankena K, et al. The effect of becoming BVDV-free on fertility and udder health in dutch dairy herds. *Preventive Veterinary Medicine*. 2008; 84 (1-2), 48-60. Doi: 10.1016/j.prevetmed.2007.11.002
48. Rola JG, Larska M, Grzeszuk M, et al. Bulk tank milk somatic cell counts in dairy herds with different bovine viral diarrhoea virus status in Poland. *Preventive Veterinary Medicine*. 2014; 116 (1-2), 183-7. Doi: 10.1016/j.prevetmed.2014.06.007
49. Tiwari A, Vanleeuwen JA, Dohoo IR, et al. Production effects of pathogens causing bovine leukosis, bovine viral diarrhea, paratuberculosis, and neosporosis. *Journal of Dairy Science*. 2007; 90 (2), 659-69. Doi: 10.3168/jds.S0022-0302(07)71548-5
50. Tiwari A, Van Leeuwen JA, Dohoo IR, et al. Effects of seropositivity for bovine leukemia virus, bovine viral diarrhoea virus, mycobacterium avium subspecies paratuberculosis, and neospore caninum on culling in dairy cattle in four canadian provinces. *Veterinary Microbiology*. 2005; 109 (3-4), 147-58. Doi: 10.1016/j.vetmic.2005.05.01
51. Kitching RP. Clinical variation in foot and mouth disease: cattle. *Revue scientifique et technique*. 2002; 21 (3), 499-504. Doi: 10.20506/rst.21.3.1343
52. Reid SM, Parida S, King DP, et al. Utility of automated real-time RT-PCR for the detection of foot-and-mouth disease virus excreted in milk. *Veterinary Research*. 2006; 37 (1), 121-32. Doi: 10.1051/vetres:2005040
53. Sharma N. Foot and mouth disease mastitis cascade in dairy cattle: a field study. *International Journal of Zoological Research*. 2008; 4 (1), 64-7. Doi: 10.3923/ijzr.2008.64.67

54. Armson B, Mioulet V, Doel C, et al. Detection of foot-and-mouth disease virus in milk samples by real-time reverse transcription polymerase chain reaction: optimisation and evaluation of a high-throughput screening method with potential for disease surveillance. *Veterinary Microbiology*. 2018; 223, 189-94. Doi: 10.1016/j.vetmic.2018.07.024
55. Frie MC, Coussens PM. Bovine leukemia virus: a major silent threat to proper immune responses in cattle. *Veterinary Immunology and Immunopathology*. 2014; 163 (3-4), 103-14. Doi: 10.1016/j.vetimm.2014.11.014
56. Sandev N, Koleva M, Binev R, et al. Influence of enzootic bovine leukosis virus upon the incidence of subclinical mastitis in cows at a different stage of infection. *Veterinarski Arhiv*. 2004; 74 (6), 411-6.
57. Watanabe A, Murakami H, Kakinuma S, et al. Association between bovine leukemia virus proviral load and severity of clinical mastitis. *The Journal of Veterinary Medical Sciencej*. 2019; 81 (10), 1431-7. Doi: 10.1292/jvms.19-0285
58. Bojarój-Nosowicz B, Kaczmarczyk E. Somatic cell count and chemical composition of milk in naturally BLV-infected cows with different phenotypes of blood leukocyte acid phosphatase. *Archives Tierz Dummerstorf*. 2006; 49 (1), 17-28. Doi: 10.5194/aab-49-17-2006
59. Yang Y, Fan W, Mao Y, et al. Bovine leukemia virus infection in cattle of China: association with reduced milk production and increased somatic cell score. *Journal of Dairy Science*. 2016; 99 (5), 3688-97. Doi: 10.3168/jds.2015-10580
60. Norby B, Bartlett PC, Byrem TM, et al. Effect of infection with bovine leukemia virus on milk production in Michigan dairy cows. *Journal of Dairy Science*. 2016; 9 (3), 2043-52. Doi: 10.3168/jds.2015-10089
61. Bhatia S, Patil S, Sood R. Bovine immunodeficiency virus: a lentiviral infection. *Indian Journal of Virology*. 2013; 24 (3), 332-341. Doi: 10.1007/s13337-013-0165-9

Bölüm 4

KİTİN-KİTOSAN: HAYVAN YETİŞTİRİCİLİĞİ VE SAĞLIĞINDA KULLANIMI

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GİRİŞ

Kitin ve Kitosan

Kitin biyopolimer yapıda, dünyada selülozdan sonra en yaygın olarak bulunan ikinci maddedir. Deniz kabuklu su ürünlerinin (yengeç, karides gibi) ana bileşenidir. Ayrıca, böceklerin iskelet, mantarların da hücre duvarlarının yapısında yer almaktadır. Dünyadaki kitin üretimi karidesten, çeşitli deniz kabuklularından, mantarlardan ve istiridyelerden sağlanmaktadır ⁽¹⁾. Böcek kabuklarının yapısında yaklaşık % 24, yengeçte % 17 ve karideste ise % 32 oranında kitin bulunmaktadır ⁽²⁾. Yapısında kitin bulunan bütün canlıların her yıl ortalama 10 milyar ton kitin üretme kapasitesine sahip olduğu bildirilmektedir ⁽³⁾.

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KAYNAKLAR

1. Guang, WY. (2002). The effect of chitosan and its derivatives on the dyeability of silk, Ph.D. Thesis, Hong Kong Polytechnic University.
2. Demir A, Seventekin N. Kitin, Kitosan ve genel kullanım alanları. *Tekstil Teknolojileri Elektronik Dergisi* 2009;3(2):92-103.
3. Kaur S, Dhillon GS. The versatile biopolymer chitosan: potential sources, evaluation of extraction methods and applications. *Crit Rev Microbiol*. 2014;40(2):155-175.
4. Gadghey KK, Bahekar A. Studies on extraction methods of chitin from crab shell and investigation of its mechanical properties. *International Journal of Mechanical Engineering and Technology* 2017;8(2):220-231.
5. Anonim (2017). Chitosan: Revolutionizing weight loss? (30/08/2019 tarihinde http://www.vanderbilt.edu/AnS/psychology/health_psychology/chitosan1.html adresinden ulaşılmıştır).
6. Anonim (2019a). History of chitin. (30/08/2019 tarihinde <http://www.primex.is/quality-and-environment/history-of-chitin/> adresinden ulaşılmıştır).
7. Anonim (2019b). The history of chitosan. (08/08/2019 tarihinde <https://www.streetdirectory.com/etoday/the-history-of-chitosan-euolup.html> adresinden ulaşılmıştır).
8. Kurita K. Chitin and chitosan: Functional biopolymers from marine crustaceans. *Mar Biotechnol*. 2006;8:203-226.
9. Sagheer FAA, Sughayer MAA, Muslim S, et al. Extraction and characterization of chitin and chitosan from marine sources in Arabian Gulf. *Carbohydr Polym*. 2009;77:410-419.
10. Jeon YJ, Park PJ, Kim SK. Antimicrobial effect of chitoooligosaccharides produced by bioreactor. *Carbohydr Polym*. 2001;44:71-76.
11. Chung Y, Su Y, Chen C, et al. Relationship between antibacterial activity of chitosan and surface characteristics of cell wall. *Acta Pharmacol Sin*. 2004;25(7):932-936.
12. Zivanovic S, Chi S, Draughon AF. Antimicrobial activity of chitosan films enriched with essential oils. *J Food Sci*. 2005;70(1):45-51.
13. Campaniello D, Bevilacqua A, Sinigaglia M, et al. Chitosan: Antimicrobial activity and potential applications for preserving minimally processed strawberries. *Food Microbiol*. 2008;25:992-1000.
14. Moreira MR, Roura SI, Ponce A. Effectiveness of chitosan edible coatings to improve microbiological and sensory quality of fresh cut broccoli. *Food Sci Technol*. 2011;44:2335-2341.
15. Pizato S, Cortez-Vega WR, De Souza JTA, et al. Effects of different edible coatings in physical, chemical and microbiological characteristics of minimally processed peaches (*Prunus Persica* L. Batsch). *J Food Saf*. 2013;33:30-39.
16. Jeon YJ, Kamil YVAJ, Shahidi F. Chitosan as an edible invisible film for quality preservation of herring and Atlantic cod. *J Agric Food Chemistry* 2002;50:5167-5178.

17. No HK, Prinyawiatkul W, Meyers SP. Comparison of shelf life of eggs coated with chitosans prepared under various deproteinization and demineralization times. *J Food Sci.* 2005;70:377-382.
18. Berger J, Reist M, Mayer JM, et al. Structure and interactions in covalently and ionically crosslinked chitosan hydrogels for biomedical applications. *Eur J Pharm Biopharm.* 2004;57:19-34.
19. de Queiroz Antonino RSCM, Fook BRPL, de Oliveira Lima VA, et al. Preparation and characterization of chitosan obtained from shells of shrimp (*Litopenaeus vannamei* Boone). *Mar Drugs* 2017;15(5):E141.
20. Kaya M, Muhtaba M, Ehrlich H, et al. On chemistry of γ -chitin. *Carbohydr Polym.* 2017;176:177-186.
21. Younes I, Rinaudo M. Chitin and chitosan preparation from marine sources. structure, properties and applications. *Mar Drugs* 2015;13:1133-1174.
22. Li Q, Dunn ET, Grandmaison EW, et al. Applications and properties of chitosan. *J Bioactive and Compatible Polym.* 1992;7:370-397.
23. Nessa F, Masum SM, Asaduzzaman M, et al. A process for the preparation of chitin and chitosan from prawn shell waste. *Bangladesh J Sci Ind Res.* 2010;45(4):323-330.
24. No HK, Meyers SP. Preparation and characterization of chitin and chitosan-A review. *J Aquat Food Prod Technol.* 1995;24(1):105-113.
25. Roberts, G. A. F. (1992). *Chitin Chemistry*. London: Macmillan Publishing.
26. Zargar V, Asghari M, Dashti A. A review on chitin and chitosan polymers: structure, chemistry, solubility, derivatives, and applications. *Chem Bio Eng Rev.* 2015;2(3):204-226.
27. Lim SH (2003). Synthesis of a fiber-reactive chitosan derivative and its application to cotton fabric as an antimicrobial finish and a dyeing-improving agent. (25/08/2019 tarihinde <http://www.lib.ncsu.edu/resolver/1840.16/5476> adresinden ulaşılmıştır).
28. Kaur S, Dhillon GS. Recent trends in biological extraction of chitin from marine shell wastes: A review. *Crit Rev Microbiol.* 2015;35(1):44-61.
29. Cheung RCF, Ng TB, Wong JH, et al. Chitosan: An update on potential biomedical and pharmaceutical applications. *Mar Drugs* 2015;13:5156-5186.
30. Percot A, Viton C, Domard A. Optimization of chitin extraction from shrimp shells. *Biomacromolecules.* 2003;4(1):12-18.
31. Ameh AO, Isa MT, Ayinla M, et al. Lactic acid demineralization of shrimp shell and chitosan synthesis. *Leonardo J Sci.* 2015;26:57-66.
32. Knaul JZ, Hudson SM, Creber KAM. Improved mechanical properties of chitosan fibers. *J Appl Polym Sci.* 1999;72:1721-1732.
33. Dhillon GS, Kaur S, Brar SK, et al. Green synthesis approach: Extraction of chitosan from fungus mycelia. *Crit Rev Biotechnol.* 2013;33(4):379-403.
34. Khanafari R, Sanatei M-Sh. Recovery of chitin and chitosan from shrimp waste by chemical and microbial methods. *Iran J Environ Health Sci Eng* 2008;5:19-24.

35. Raimbault M. (1998). General and microbiological aspects of solid substrate fermentation. *EJB Elec J Biotechnol*, 1998;1:2-15.
36. Luis CA, Huerta S, Hall GM, et al. Pilot scale lactic acid fermentation of shrimp wastes for chitin recovery. *Proc Biochem*, 2002;37:1359-1366.
37. Xie L, Hettiarachchy NS, Ju ZY, et al. Edible film coating minimize eggshell breakage and reduce post-wash bacterial contamination measured by dye penetration in eggs. *J Food Sci*. 2002;67(1):280-284.
38. Suyatma NE, Tighzert L, Copinet A. Effects of hydrophilic plasticizers on mechanical, thermal, and surface properties of chitosan films. *J Agric Food Chem*. 2005;53:3950-3957.
39. Caner C, Küçük M. 2004. Yenilebilir film ve kaplamalar: Gıdalara uygulanabilirliği. *Gıda Mühendisliği ve Gıda Sanayi Dergisi* 2004;8:30-35.
40. Küçüköner, E., Kılınççeker, O, & Dogan, S. (2003). Gıdalara Yenilebilir Kaplama Uygulamalarında Süt Ürünlerinin Kullanım Olanakları. *Süt Endüstrisinde Yeni Eğitimler Sempozyumu*, 22-23 Mayıs 2003, Bornova, İzmir, (pp. 251-256).
41. Drewnowska O, Turek B, Carstensen B, et al. Chitosan - A promising biomaterial in veterinary medicine. *Pol J Vet Sci*. 2013;16(4):843-848.
42. Şenel S, McClure SJ. Potential applications of chitosan in veterinary medicine. *Adv Drug Deliv Rev*. 2004;56:1467-1480.
43. Neuman MG, Nanau RM, Oruña-Sánchez L, et al. Hyaluronic acid and wound healing. *J Pharm Pharm Sci*. 2015;18(1):53-60.
44. Baykal Y, Karaayvaz M, Kutlu M. İnterlökinler. *T Klin J Med Sci*. 1998;18:77-81.
45. Kosaka, T, Kaneko Y, Nakada Y, et al. Effect of chitosan implantation on activation of canine macrophages and polymorphonuclear cells after surgical stress. *J Vet Med Sci*. 1996;58:963-967.
46. Okamoto Y, Shibasaki K, Minami S, et al. Evaluation of chitin and chitosan on open wound healing in dogs. *J Vet Med Sci*. 1995;57(5):851-854.
47. Tekelioglu BK, Celik M, Kucukgulmez A. Canine extremity wound treatment with chitosan extracted from shrimp shells: A case report. *J Agric Sci Technol A*. 2017;7:274-281.
48. Minami S, Okamoto Y, Hamada K, et al. (1999). Veterinary practice with chitin and chitosan. In: P. Jolles, R.A.A. Muzzarelli (Eds.), *Chitin and Chitinases* (pp. 265-277). Basel, Switzerland, Birkhauser Verlag.
49. Kurtoglu AH, Karatas A. Yara tedavisinde güncel yaklaşımlar: Modern yara örtüleri. *Ankara Ecz Fak Derg*. 2009;38(3):211-232.
50. Movaffagh J, Fazly-Bazzaz BS, Yazdi AT, et al. Wound healing and antimicrobial effects of chitosan-hydrogel/honey compounds in a rat full-thickness wound model. *Wounds* 2019; 31(9): 228-235.
51. Muzzarelli RAA. Biochemical significance of exogenous chitins and chitosans in animals and patients. *Carbohydr Polym*. 1993;20:7-16.

52. Muzzarelli RAA, Ramos V, Stanic V, et al. Osteogenesis promoted by calcium phosphate N,N-dicarboxymethyl chitosan. *Carbohydr Polym.* 1998;36: 267-276.
53. Lu JX, Prudhommeaux F, Meurier A, et al. Effects of chitosan on rat knee cartilages. *Biomaterials* 1999;20:1937-1944.
54. Mi FL, Wu YB, Shyu SS, et al. Asymmetric chitosan membranes prepared by dry/wet phase separation: A new type of wound dressing for controlled antibacterial release. *J Membr Sci.* 2003;212:237-254.
55. Ferrara L. Phytotherapy as a preventive and adjuvant for the rhinitis. *IOSR J Pharm.* 2016;6(1):6-11.
56. Lanctôt S, Fustier P, Taherian AR, et al. Effect of intramammary infusion of chitosan hydrogels at drying-off on bovine mammary gland involution. *J Dairy Sci.* 2017;100(3):2269-2281.
57. Alam MR, Kim WI, Kim JW, et al. Effects of Chitosan-oligosaccharide on diarrhoea in Hanwoo calves. *Vet Med-Czech.* 2012;57(8):385-393.
58. Asli A, Brouillette E, Ster C, et al. Antibiofilm and antibacterial effects of specific chitosan molecules on *Staphylococcus aureus* isolates associated with bovine mastitis. *PLoS One.* 2017;12(5):e0176988.
59. Tasker RAR, Ross SJ, Dohoo, SE, et al. Pharmacokinetics of an injectable sustained release formulation of morphine for use in dogs, *J. Vet. Pharmacol. Ther.* 1997;20:362-367.
60. Bowersock TL, Martin S. Vaccine delivery to animals. *Adv Drug Deliv Rev.* 1999;38:167-194.
61. Koping-Hoggard M, Ocklind G, Guan H, et al. B. Chitosan and PEI-pDNA polyplexes: in vivo gene expression after tracheal, nasal and oral administration to mice. *Proc Int Symp Control Release Bioact Mater.* 1999;26:795-796.
62. Janes KA, Calvo P, M.J. Alonso MJ. Polysaccharide colloidal particles as delivery systems for macromolecules. *Adv Drug Deliv Rev.* 2001;47:83-97.
63. Van der Lubben IM, Verhoef JC, Borchard G, et al. Chitosan and its derivatives in mucosal drug and vaccine delivery. *Eur J Pharm Sci.* 2001;14:201-207.
64. Abdel-Ghany HM, Salem MES. Effects of dietary chitosan supplementation on farmed fish; a review. *Rev Aquacult.* 2019;1:1-15.
65. Wisdom KS, Bhat IA, Kumar P, et al. Fabrication of chitosan nanoparticles loaded with aromatase inhibitors for the advancement of gonadal development in *Clarias magur* (Hamilton, 1822). *Aquaculture* 2018;497:125-133.
66. Bhat IA, Ahmad I, Mir IN, et al. Chitosan-eurycomanone nanoformulation acts on steroidogenesis pathway genes to increase the reproduction rate in fish. *J Steroid Biochem Mol Biol.* 2019a;185:237-247.
67. Bhat IA, Ahmad I, Mir IN, et al. Evaluation of the in vivo effect of chitosan conjugated eurycomanone nanoparticles on the reproductive response in female fish model. *Aquaculture* 2019b;510:392-399.
68. Kılınççeker, O., & Dogan, S. (2002). *Kaplama Ürünlerinde Tahıl Unlarının Kullanımı. Hububat Ürünleri Teknolojisi ve Kongre Sergisi*, 3-4 Ekim 2002, Gaziantep (pp. 441-450).

69. Lee JY, Park HJ, Lee CY, et al. Extending Shelf-Life of Minimally Processed Apples with Edible Coatings and Antibrowning Agents. *Lebensm-Wiss-U-Technol.* 2003;36:323-329.
70. Savini, V. (2018). *Pet-to-Man Travelling Staphylococci* (First edit). Cambridge: Academic Press.
71. Akdemir, F., Köseman, A. & Şeker, İ. (2017). Farklı sürelerde depolanan kuluçkalık tavuk yumurtalarını kitosanlı filmle kaplamanın yumurtaların depolama sırasındaki ağırlık kayıpları bazı kuluçka özellikleri civciv çıkım ağırlıkları ve yaşama güçleri üzerine etkisi. İÜBAP, Proje No: FCD-2017-703.
72. Cansız, Ö., Aday, MS. & Caner, C. (2006). Kitosan kaplama materyalinin yumurtanın kabuk mukavemetini (kalite kriterini) geliştirmede etkinliği. Türkiye 9. Gıda Kongresi, 24-26 Mayıs 2006, Bolu, (p. 875).
73. Razdan A, Petterson D, Pettersson J. Broiler chicken body weights, feed intakes, plasma lipid and small-intestinal bile acid concentrations in response to feeding of chitosan and pectin. *Br J Nutr.* 1997;78(2):283-291.
74. Tufan T, Arslan C, Durna Ö, et al. Effects of chito-oligosaccharides and L-carnitine supplementation in diets for Japanese quails on performance, carcass traits and some blood parameters. *Arq Bras Med Vet Zootec.* 2015;67(1):283-289.
75. Kamali Najafabad M, Imanpoor MR, Taghizadeh V, et al. Effect of dietary chitosan on growth performance, hematological parameters, intestinal histology and stress resistance of Caspian kutum (*Rutilus frisii kutum* Kamenskii, 1901) fingerlings. *Fish Physiol Biochem.* 2016;42(4):1063-1071.

Bölüm 5

NUTRİSYONEL GENOMİK VE VETERİNER HEKİMLİK

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GİRİŞ

Beslenme araştırmaları, yakın zamana kadar gıdalardaki besin maddelerinin eksiklikleri ve buna bağlı olarak insan sağlığının bozulması konuları üzerinde yoğunlaşmıştı. Ancak genomiklerin ortaya çıkışı, genomların yapısı, bileşimi ve işlevleri hakkında bilimsel verilerin elde edilmesi, besin maddelerinin bunları tüketen canlıların gen ve protein ekspresyonunu nasıl değiştirdiğine ilişkin anlayışı arttırarak beslenme konusuna yeni bir bakış açısı getirmiştir ⁽¹⁾.

Canlıların özellikle insan ve hayvanların sahip olduğu genetik yapı, beslenmeye bağlı olarak ortaya çıkan hastalıkları azaltan ya da arttıran bir faktördür (Şekil 1). Genetik yapıyı oluşturan genlerin sindirimdeki rolü ve besin öğelerinin oluşturduğu kalıtsal değişiklikler, besinlerin bireye özgü özellikler taşıdığını ve bu nedenle

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- Pet hayvanlarının insanlar gibi daha uzun, sağlıklı ve kaliteli yaşaması mümkün olabilecektir,
- Hatta hem insan hem de hayvanlar için bireysel diyet uygulamaları yaygınlaşabilecektir.

Gelecek için büyük umutlar vadeden bu alandaki çalışmaların multidisipliner ve yaygın olarak sürdürülmesi insanlığa önemli kazanımlar sağlayacaktır. Ayrıca, bu konularla ilgili veteriner hekimlik alanında elde edilecek başarılar da insan hekimliğine katkı sunacaktır.

KAYNAKLAR

1. Afman L, Müller M. Nutrigenomics: from molecular nutrition to prevention of disease. *J Am Diet Assoc.* 2006;106 (4): 569-76.
2. Almén MS, Nilsson EK, Jacobsson JA, et al. Genome-wide analysis reveals DNA methylation markers that vary with both age and obesity. *Gene.* 2014; 548 (1): 61-7.
3. Amaral PP, Dinger ME, Mercer TR, et al. The eucaryotic genome as an RNA machine. *Science.* 2008; 319 (5871): 1787-9.
4. Anonim. 2008. *DNA metilasyonu ve beslenme ilişkisi.* <https://learn.genetics.utah.edu/content/epigenetics/nutrition/2008>. Erişim tarihi: 05.05.2019.
5. Anonim. 2013. *Asetil molekülünün histon proteinine bağlanması.* <http://www.mbgbilim.com/epigenetik/2013>. Erişim tarihi:18.05.2019.
6. Anonim. 2015. *DNA Metilasyonu.* <https://www.jove.com/video/3170/dna-metilasyonu-bisfit-modifikasyonu-ve-analiz?language=Turkish>. 2015. Erişim tarihi: 04.05.2019.
7. Anonim. 2019a. *Dosya:Genome-tr.svg.* <https://tr.wikipedia.org/wiki/Dosya:Genome-tr.svg>. Erişim tarihi: 03.02.2019.
8. Anonim. 2019b. *What is Epigenetics.* <https://www.whatisepigenetics.com/non-coding-rna/> Erişim tarihi: 05.05.2019.
9. Anonim. 2019c. *Kraliçe arının meydana gelişi.* <https://www.elifguveloglu.com/single-post/2018/05/11/Nutrigenomik-Beslenme>. Erişim tarihi: 17.5.2019.
10. Anonim. 2019d. *DNA'dan mRNA kodlanması ve RNA modifiasyonu.* <https://www.whatisepigenetics.com/non-coding-rna/>. Erişim tarihi: 05.05.2019.
11. Arena S, Renzone G, Scaloni A. A multi-approach peptidomic analysis of hen egg white reveals novel putative bioactive molecules. *J Proteomics.* 2020; 9 (215): 103646.

12. Başaran E, Aras S, Cansaran-Duman D. Genomik, proteomik, metabolik kavramlarına genel bakış ve uygulama alanları. *Türk Hij Den Biyol Derg.* 2010; 67 (2): 85-96.
13. Bertram HC, Bach Knudsen KE, Serena A, et al. NMR-based metabolomic studies reveal changes in the biochemical profile of plasma and urine from pigs fed high-fibre rye bread. *Brit J Nutr.* 2006; 95: 955-962.
14. Caramia G. Childhood feeding, chronic-degenerative disease in adults, and nutrigenomics. *Pediatr Med Chir.* 2007; 29 (6): 309-20.
15. Choi SW, Friso S. Epigenetics: A new bridge between nutrition and health. *Adv Nutr.* 2010; 1 (1): 8-16.
16. Collins LJ, Cheni XS. Ancestral RNA: The biology of the eucaryotic ancestor. *RNA Biol* 2009; 6 (5): 495-502.
17. Connolly S, Dona A, Wilkinson-White L, et al. Relationship of the blood metabolome to subsequent carcass traits at slaughter in feedlot Wagyu crossbred steers. *Sci Rep.* 2019; 9 (1): 15139.
18. Coşkun T. Nütrisyonel genomik. *Çocuk Sağlığı ve Hastalıkları Dergisi.* 2007; 50: 47-66
19. Dawson, K.A. Nutrigenomics: feeding the genes for improved fertility. *Animal Reprod Sci.* 2006; 96: 312-322.
20. Dervishi E, Serrano C, Joy M, et al. Effect of the feeding system on the fatty acid composition, expression of the Delta9-desaturase, Peroxisome Proliferator Activated Receptor Alpha, Gamma, and Sterol Regulatory Element Binding Protein 1 genes in the semitendinous muscle of light lambs of the Rasa Aragonesa breed. *BMC Vet Res.* 2010; 22: 6-40.
21. Elango N, Hunt BG, Goodisman MA, et al. DNA methylation is widespread and associated with differential gene expression in castes of the honeybee, *Apis mellifera*. *Proc Natl Acad Sci USA.* 2009; 106: 11206-11.
22. El-Sohemy A. Nutrigenetics. *Forum Nutr.* 2007; 60: 25-30.
23. Foret S, Kucharski R, Pittelkow Y, et al. Epigenetic regulation of the honey bee transcriptome: unraveling the nature of methylated genes. *BMC Genomics.* 2009; 10: 472.
24. Fumagalli M, Moltke I, Grarup N, et al. Greenlandic Inuit show genetic signatures of diet and climate adaptation. *Science.* 2015; 349: 1343-1347.
25. Giuseppe A, James L. An Emerging Role for Metabolomics in Nutrition Science. *J Nutrigenet Nutrigenomics.* 2013; 6: 181-200.
26. Goodacre R. Metabolomics-the way forward. *Metabolomics.* 2005; 1: 1-2.
27. Herrera-Marcos LV, Lou-Bonafonte JM, Arnal C, et al. Transcriptomics and the Mediterranean diet: A systematic review. *Nutrients.* 2017; 9: 9 (5). pii: E472.
28. Izmirli M. Epigenetik mekanizmalar ve kanser tedavisinde epigenetik yaklaşımlar. *Van Tıp Dergisi.* 2013; 20 (1): 48-51.
29. Jiménez-Chillaron JC, Díaz R, Martínez D, et al. The role of nutrition on epigenetic modifications and their implications on health. *Biochimie.* 2012; 94 (11): 2242-63.

30. Joseph SJ, Pratt SL, Pavan E, et al. Omega-6 fat supplementation alters lipogenic gene expression in bovine subcutaneous adipose tissue. *Gene Regul. Syst. Bio.* 2010; 19 (4): 91-101.
31. Kaput J, Rodriguez RL. Nutritional genomics: the next frontier in the post-genomic era. *Physiol Genomics.* 2004; 16 (2): 166-77.
32. Kauwell GPA. Emerging concepts in nutrigenomics: a preview of what is to come. *Nutr Clin Pract.* 2005; 20 (1): 75-87.
33. Keyes MK, Jang H, Mason JB, et al. Older age and dietary folate are determinants of genomic and p16- specific DNA methylation in mouse colon. *J Nutr.* 2007; 137:1713-7.
34. Kucharski R, Maleszka J, Foret S, et al. Nutritional control of reproductive status in honeybees via DNA methylation. *Science.* 2008; 319:1827-30.
35. Law PP, Holland ML. DNA methylation at the crossroads of gene and environment interactions. *Essays Biochem.* 2019; 63 (6): 717-726.
36. Li L, Yang M, Li C, et al. Understanding the toxin effects of β -Zearalenol and HT-2 on bovine granulosa cells using iTRAQ-based proteomics. *Animals (Basel).* 2020; 10 (1). pii: E130
37. JJ, Dann HM, Janovick Guretzky NA, et al. Plane of nutrition prepartum alters hepatic gene expression and function in dairy cows as assessed by longitudinal transcript and metabolic profiling. *Physiol Genomics.* 2006; 27: 29-41.
38. Luzzatto L, Pandolfi PP. Causality and Chance in the Development of Cancer. *N Engl J Med.* 2015; 373 (1): 84-88.
39. Mathieson I, Lazaridis I, Rohland N, et al. Genome-wide patterns of selection in 230 ancient Eurasians. *Nature.* 2015; 528: 499-503.
40. Menezes EB, Velho ALC, Santos F, et al. Uncovering sperm metabolome to discover biomarkers for bull fertility. *BMC Genomics.* 2019; 20 (1): 714.
41. Miggiano GA, De Sanctis R. Nutritional genomics: toward a personalized diet. *Clin Ter.* 2006; 157 (4): 355-61.
42. Mobuchon L, Le Guillou S, Marthey S, et al. Sunflower oil supplementation affects the expression of miR20a-5p and miR-142-5p in the lactating bovine mammary gland. *PLoS ONE* 2017; 12 (12): e0185511.
43. Neiberghs HL, Johnson KA. Alpha Beef Cattle Nutrition Symposium: nutrition and the genome. *J Anim Sci.* 2012; 90 (7): 2308-16.
44. Newgard CB. Metabolomics and Metabolic Diseases: Where Do We Stand? *Cell Metab.* 2017; 25 (1): 43-56.
45. Nowacka-Woszek J. Nutrigenomics in livestock-recent advances. *J Appl Genet.* 2019; 61 (1): 93-103.
46. Ommen B, Broek T, Hoogh I, et al. Systems biology of personalized nutrition. *Nutrition Reviews.* 2017; 75 (8): 579-599
47. Oppermann U. Why is epigenetics important in understanding the pathogenesis of inflammatory musculoskeletal diseases? *Arthritis Res Ther.* 2013; 15 (2): 209.

48. Ordovas JM. The quest for cardiovascular health in the genomic era: nutrigenetics and plasma lipoproteins. *Proc Nutr Soc.* 2004; 63 (1): 145-52.
49. Ordovas JM, Corella D. Nutritional genomics. *Annu Rev Genomics Hum Genet.* 2004; 5: 71-118.
50. Qudeer AA, Byung PY, Hae YC, et al. Epigenetic modifications of gene expression by lifestyle and environment. *Arch. Pharm. Res.* 2017; 40:1219-1237.
51. Rao L, Puschner B, Prolla TA. Gene expression profiling of low selenium status in the mouse intestine: transcriptional activation of genes linked to DNA damage, cell cycle control and oxidative stress. *J. Nutr.* 2001; 131: 3175-3181.
52. Rebel JM, Van Hemert S, Hoekman AJ, et al. Maternal diet influences gene expression in intestine of offspring in chicken (*Gallus gallus*). *Comp. Bioc. hem. Physiol. A Mol. Integr. Physiol.* 2006; 145 (4): 502-508
53. Sarıgöl Z, Ündeğer Bucurğat Ü. Epigenetik değişiklikler ve beslenme ilişkisi. *Türkiye Klinikleri J Pharm Sci.* 2014; 3 (2): 74-80.
54. Sasaki H, Matsui Y. Epigenetic events in mammalian germ-cell development: reprogramming and beyond. *Nat Rev Genet.* 2008; 9: 129-40.
55. Saxena R, Saxena VK, Tripathi V, et al. Dynamics of gene expression of hormones involved in the growth of broiler chickens in response to the dietary protein and energy changes. *Gen Comp Endocrinol.* 2019; 288:113377.
56. Seçer S, Özden A, Güllüoğlu S, ve ark. Diyetle ilişkili hastalıkların önlenmesi ve hayat kalitesinin iyileştirilmesi için genotiplemeyle dayalı kişiye özel beslenme. *Akademik Gıda.* 2011; 9 (4): 51-59.
57. Selçuk S. 2018. *Nütrisyonel genetik nedir? genetiğin beslenme üzerine etkileri nelerdir? kişiselleştirilmiş beslenme nedir?* <https://www.birbes.com/nutrisyonel-genetik-nedir-genetigin-beslenme-uzerine-etkileri-nelerdir-5328/>. Erişim tarihi: 11.02.2020.
58. Simopoulos AP. Genetic variation and dietary response: nutrigenetics/nutrigenomics. *Asia Pacific J Clin Nutr.* 2002; 11: 117-128.
59. Song S, Hooiveld GJ, Zhang W, et al. Comparative Proteomics Provides Insights into Metabolic Responses in Rat Liver to Isolated Soy and Meat Proteins. *J Proteome Res.* 2016; 15 (4): 1135-42.
60. Trujillo E, Davis C, Milner J. Nutrigenomics, Proteomics, Metabolomics, and the practice of dietetics. *J. Am. Diet. Assoc.* 2006; 106: 403-413.
61. Tucker KL. Methylated cytosine and the brain: A new base for neuroscience. *Neuron.* 2001; 30 (3): 649-52
62. Usanma Koban B, Vural Tuzcular Z, Gönenç I, ve ark. Beslenme, diğer çevresel faktörler ve mikrobiyotanın obezite epigenetiğine etkileri. *Jour Turk Fam Phy.* 2017; 8 (3): 108-117.
63. Vahid F, Zand H, Nosrat-Mirshekarlou E, et al. The role dietary of bioactive compounds on the regulation of histone acetylases and deacetylases: A review. *Gene.* 2015; 562 (1): 8-15.

64. Van Dijk SJ, Feskens EJ, Bos MB, et al. Consumption of a high monounsaturated fat diet reduces oxidative phosphorylation gene expression in peripheral blood mononuclear cells of abdominally overweight men and women. *J. Nutr.* 2012; 142: 1219–1225.
65. Vargas-Bello-Pérez E, Geldsetzer-Mendoza C, Cancino-Padilla N, et al. Effects of Dietary Vegetable Oils on Mammary Lipid-Related Genes in Holstein Dairy Cows. *Animals (Basel)*. 2019; 10 (1). pii: E57.
66. Waddington CH. The epigenotype. 1942. *Int J Epidemiol.* 2012; 41 (1): 10-3.
67. Wakabayashi K, Okamura M, Tsutsumi S, et al. The peroxisome proliferator-activated receptor gamma/retinoid X receptor alpha heterodimer targets the histone modification enzyme PR-Set7/Setd8 gene and regulates adipogenesis through a positive feedback loop. *Mol Cell Biol.* 2009; 29 (13): 3544–55.
68. Weihrauch-Blüher S, Richter M, Staeger MS. Body weight regulation, socioeconomic status and epigenetic alterations. *Metabolism.* 2018; (18) 30072-6.
69. Williams MB, Watts SA. Current basis and future directions of zebrafish nutrigenomics. *Genes Nutr.* 2019; 14:34.
70. Wilson AG. Epigenetic regulation of gene expression in the inflammatory response and relevance to common diseases. *J Periodontol.* 2008; 79 (8 Suppl):1514-9.
71. Yılmaz S. 2019. *Nutrisyonel genomik*. <https://acikders.ankara.edu.tr/mod/resource/view.php?id=16735>. Erişim tarihi: 18.5.2019.