

## **Bölüm 2**

# **HÜCRE ÖLÜMÜNE GENEL BAKIŞ VE GÜNCEL HÜCRE ÖLÜMÜ MEKANİZMALARI**

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### **GİRİŞ**

Vücudumuzda yaşam döngüsünü tamamlayıp sağlıklı yaşlanmış hücreler ve hasar görmüş hücreler de dahil olmak üzere günde milyonlarca hücre yerlerine sağlıklı yeni hücrelerin gelmesi amacıyla ölmektedir. Hücre ölümü, ölü ya da ölmekte olan hücre tanımını ve hücre ölümü çeşitlerini bilimsel çalışma ve yayınlanan bilimsel makalelere dayandırarak sınıflandıran ve bu sınıflandırmayı seriler halinde güncel bilgiler ışığında yayınlayan bir “Hücre Ölümü Adlandırma Komitesi- The Nomenclature Committee on Cell Death (NCCD) mevcuttur. Komisyonun amacı hücre ölümü alanında devam eden bilimsel çalışmaların çıktılarını göz önünde tutarak hücre ölümü konusunda kabul görmüş bir isimlendirme ve sınıflandırma sağlamaktır (1). Bu amaç doğrultusunda farklı hücre ölümü çeşitleri farklı açılardan (örneğin morfolojik, biyokimyasal, fonksiyonel ve terapötik uygulamalar) değerlendirilmektedir ve bu konuda araştırmacılar arasında tartışma ve görüş paylaşımı ortamı yaratılmıştır. Böylece ortak bir dil ile oluşturulmuş esnek kavramlar geliştirilmiştir ve hücre ölümü çalışan bilim insanları arasında iletişim kurulması sağlanmıştır (2). Komite 2005 yılından bu yana beş derleme makale yayınlamıştır (1–5). Komitenin 2005 ve 2009 yıllarında yayınlanan derlemelerinde hücre ölümü litetratüründeki kavramlardan ve hücre ölümünün sınıflandırılmasından bahsedilmiştir (2,5). Bir sonraki derleme makalesinde ise (4) ise hücre ölümü alt tiplerinin moleküler düzeyde meydana gelen süreçleri açıklanmıştır. 2015 yılındaki derlemede komite hücre ölümüne farklı bir bakış açısı kazandırmak adına hücre ölümünü “gerekli” (essential) ya da “yardımcı” (accessory) yönden değerlendirmiştir. Bir diğer deyişle hücre ölümü mekanizmalarının sırasıyla etiyolojik ve morfolojik /biyokimyasal yönden ne şekilde meydana geldiği özetlenmiştir (3). Komitenin 2018 yılında yayınladıkları son derlemede ise hücre ölümünün moleküler mekanizmalarına derinlemesine değinilmiştir (1). Bu kitap bölümünde hücre ölümü araştırmalarının sonuçlarını içeren güncel literatür bulguları derlenmiştir ve hücre ölümü alanında yapılan çalışmalar özetlenmiştir.

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bu süreçlerin her zaman hücrenin “ölümü” ile sonuçlanmayan süreçler olduğu vurgulanmaktadır. Regüle edilen (regulated) hücre ölümü süreçlerine benzer morfolojik ve biyokimyasal özellikler göstermekle beraber sözkonusu olaylar sonucunda hücrelerin ölümü gerçekleşmeyebilmektedir (1).

### **1. Hücresel Senesens (Cellular senescence)**

Senesens-ilişkili sekretuar fenotip (SASP- senescence-associated secretory phenotype) de dahil olmak üzere, hücrelerin geridönüşümsüz bir şekilde spesifik olarak morfolojik ve biyokimyasal bazı özelliklerinde kayıp meydana gelmesi sonucunda gelişir ve regüle edilen (regulated) hücre ölümü çeşidi değildir (112,113).

### **2. Mitotik Katastrof (Mitotic catastrophe)**

Mitotik katastrof, fazla miktarda meydana gelen DNA hasarı, mitotik süreçle ilgili problemler ve mitotik kontrol noktalarının yetersizliği nedeniyle mitozu tamamlayamayan hücrelerin çoğalmasını ve hayatta kalmasını engelleyen onkosupresif bir mekanizmadır (114,115). Regüle edilen (regulated) hücre ölümü çeşidi değildir. Endojen (DNA replikasyon mekanizması sırasında meydana gelen stres ve DNA replikasyonunda veya kromozom ayrılması sırasında rol alan faktörlerin düzensiz aktivitesi) ve egzojen (ksenobiyotikler) tetikleyiciler tarafından indüklenilen bir süreçtir (116,117). Örneğin; mitotik katastrof süreci sonucunda regüle edilen bir hücre çeşidi olan “mitotik ölüm” diye adlandırılan bir süreçle hücreler ölebilmektedir.

**Anahtar Kelimeler:** Hücre Ölümü, Sınıflandırma, Programlı, Ani gelişen

### **KAYNAKÇA**

1. Galluzzi L, Vitale I, Aaronson SA, et al. Molecular mechanisms of cell death: Recommendations of the Nomenclature Committee on Cell Death 2018. *Cell Death Differ.* 2018 Mar;25(3):486-541. doi: 10.1038/s41418-017-0012-4.
2. Kroemer G, Galluzzi L, Vandenabeele P, et al. Classification of Cell Death 2009. *Cell Death Differ.* 2009 Jan;16(1):3-11. doi: 10.1038/cdd.2008.150.
3. Galluzzi L, Bravo-San Pedro JM, Vitale I, et al. Essential versus accessory aspects of cell death: Recommendations of the NCCD 2015. *Cell Death Differ.* 2015 Jan;22(1):58-73. doi: 10.1038/cdd.2014.137.
4. Galluzzi L, Vitale I, Abrams JM, et al. Molecular definitions of cell death subroutines: Recommendations of the Nomenclature Committee on Cell Death 2012. *Cell Death Differ.* 2012 Jan;19(1):107-20. doi: 10.1038/cdd.2011.96.
5. Kroemer G, El-Deiry WS, Golstein P, et al. Classification of cell death: Recommendations of the nomenclature committee on cell death. *Cell Death Differ.* 2005 Nov;12 Suppl 2:1463-7.
6. Kepp O, Galluzzi L, Lipinski M, et al. Cell death assays for drug discovery. *Nat Rev Drug Discov.* 2011 Mar;10(3):221-37. doi: 10.1038/nrd3373
7. Mazzearello P. A unifying concept: The history of cell theory. *Nat Cell Biol.* 1999 May;1(1):E13-5.
8. Vaux DL. Apoptosis timeline. *Cell Death Differ.* 2002 Apr;9(4):349-54.
9. Vogt C. Untersuchungen über die Entwicklungsgeschichte der Geburtshelerkroete (Alytes obs-

- tetricians). Solothurn: Jent und Gassman, 1842, p. 130.
10. Lockshin RA, Williams CM. Programmed cell death-II. Endocrine potentiation of the breakdown of the intersegmental muscles of silkmoths. *J Insect Physiol.* Volume 10, Issue 4, August 1964, Pages 643-649.
  11. Tang D, Kang R, Berghe T Vanden, et al. The molecular machinery of regulated cell death. *Cell Res.* 2019 May;29(5):347-364. doi: 10.1038/s41422-019-0164-5.
  12. Kerr JFR, Wyllie AH, Currie AR. Apoptosis: A basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer.* 1972 Aug;26(4):239-57.
  13. Schweichel J.U, Merker H. J. The morphology of various types of cell death in prenatal tissues. *Teratology.* 1973 Jun;7(3):253-66.
  14. Clarke PGH. Developmental cell death: morphological diversity and multiple mechanisms. *Anat Embryol (Berl).* 1990;181(3):195-213.
  15. Ohsumi Y. Historical landmarks of autophagy research. *Cell Res.* 2014 Jan;24(1):9-23. doi: 10.1038/cr.2013.169.
  16. Takeshige K, Baba M, Tsuboi S, et al. Autophagy in yeast demonstrated with proteinase-deficient mutants and conditions for its induction. *J Cell Biol.* 1992 Oct;119(2):301-11.
  17. Baba M, Takeshige K, Baba N, et al. Ultrastructural analysis of the autophagic process in yeast: Detection of autophagosomes and their characterization. *J Cell Biol.* 1994 Mar;124(6):903-13.
  18. Tanida I, Ueno T, Kominami E. LC3 conjugation system in mammalian autophagy. *Int J Biochem Cell Biol.* 2004 Dec;36(12):2503- 18.
  19. Poon IKH, Lucas CD, Rossi AG, et al. Apoptotic cell clearance: Basic biology and therapeutic potential. *Nat Rev Immunol.* 2014 Mar;14(3):166-80. doi: 10.1038/nri3607
  20. Hochreiter-Hufford A, Ravichandran KS. Clearing the dead: Apoptotic cell sensing, recognition, engulfment, and digestion. *Cold Spring Harb Perspect Biol.* 2013 Jan 1;5(1):a008748. doi: 10.1101/cshperspect.a008748.
  21. Underhill DM, Goodridge HS. Information processing during phagocytosis. *Nat Rev Immunol.* 2012 Jun 15;12(7):492-502. doi: 10.1038/nri3244.
  22. Overholtzer M, Mailloux AA, Mouneimne G, et al. A Nonapoptotic Cell Death Process, Entosis, that Occurs by Cell-in-Cell Invasion. *Cell.* 2007 Nov 30;131(5):966-79.
  23. Klionsky DJ, Abdalla FC, Abeliovich H, et al. Guidelines for the use and interpretation of assays for monitoring autophagy. *Autophagy.* 2012 Apr;8(4):445-544.
  24. Galluzzi L, Aaronson SA, Abrams J, et al. Guidelines for the use and interpretation of assays for monitoring cell death in higher eukaryotes. *Cell Death Differ.* 2009 Aug;16(8):1093-107. doi: 10.1038/cdd.2009.44.
  25. Vanden Berghe T, Grootjans S, Goossens V, et al. Determination of apoptotic and necrotic cell death in vitro and in vivo. *Methods.* 2013 Jun 1;61(2):117-29. doi: 10.1016/j.ymeth.2013.02.011.
  26. Galluzzi L, Maiuri MC, Vitale I, et al. Cell death modalities: Classification and pathophysiological implications. *Cell Death Differ.* 2007 Jul;14(7):1237-43.
  27. Majno G, Joris I. Apoptosis, oncosis, and necrosis: An overview of cell death. *Am J Pathol.* 1995 Jan;146(1):3-15.
  28. Kroemer G, Levine B. Autophagic cell death: The story of a misnomer. *Nat Rev Mol Cell Biol.* 2008 Dec;9(12):1004-10. doi: 10.1038/nrm2529.
  29. Denton D, Nicolson S, Kumar S. Cell death by autophagy: Facts and apparent artefacts. *Cell Death Differ.* 2012 Jan;19(1):87-95. doi: 10.1038/cdd.2011.
  30. Jänicke RU, Sprengart ML, Wati MR, et al. Caspase-3 is required for DNA fragmentation and morphological changes associated with apoptosis. *J Biol Chem.* 1998 Apr 17;273(16):9357-60.
  31. Coleman ML, Sahai EA, Yeo M, et al. Membrane blebbing during apoptosis results from caspase-mediated activation of ROCK I. *Nat Cell Biol.* 2001 Apr;3(4):339-45.
  32. Jouan-Lanhouet S, Arshad MI, Piquet-Pellorce C, et al. TRAIL induces necroptosis involving RIPK1/RIPK3-dependent PARP-1 activation. *Cell Death Differ.* 2012 Dec;19(12):2003-14. doi: 10.1038/cdd.2012.90.
  33. Banik S, Akter M, Corpus Bondad SE, et al. Carvacrol inhibits cadmium toxicity through combating against caspase dependent/independent apoptosis in PC12 cells. *Food Chem Toxicol.*

- 2019 Dec;134:110835. doi: 10.1016/j.fct.2019.110835.
34. Belmokhtar CA, Hillion J, Ségal-Bendirdjian E. Staurosporine induces apoptosis through both caspase-dependent and caspase-independent mechanisms. *Oncogene*. 2001 Jun 7;20(26):3354-62.
  35. Casares N, Pequignot MO, Tesniere A, et al. Caspase-dependent immunogenicity of doxorubicin-induced tumor cell death. *J Exp Med*. 2005 Dec 19;202(12):1691-701.
  36. Asano K, Miwa M, Miwa K, et al. Masking of phosphatidylserine inhibits apoptotic cell engulfment and induces autoantibody production in mice. *J Exp Med*. 2004 Aug 16;200(4):459-67.
  37. Lauber K, Blumenthal SG, Waibel M, et al. Clearance of apoptotic cells: Getting rid of the corpses. *Mol Cell*. 2004 May 7;14(3):277-87.
  38. Fadok VA, Bratton DL, Henson PM. Phagocyte receptors for apoptotic cells: Recognition, uptake, and consequences. *J Clin Invest*. 2001 Oct;108(7):957-62.
  39. Hugues S, Mougneau E, Ferlin W, et al. Tolerance to islet antigens and prevention from diabetes induced by limited apoptosis of pancreatic  $\beta$  cells. *Immunity*. 2002 Feb;16(2):169-81.
  40. Green DR, Oguin TH, Martinez J. The clearance of dying cells: Table for two. *Cell Death Differ*. 2016 Jun;23(6):915-26. doi: 10.1038/cdd.2015.172.
  41. Fadok VA, Savill JS, Haslett C, et al. Different populations of macrophages use either the vitronectin receptor or the phosphatidylserine receptor to recognize and remove apoptotic cells. *J Immunol*. 1992 Dec 15;149(12):4029-35.
  42. Elmore S. Apoptosis: A Review of Programmed Cell Death. *Toxicol Pathol*. 2007 Jun;35(4):495-516.
  43. Gardai SJ, McPhillips KA, Frasch SC, et al. Cell-surface calreticulin initiates clearance of viable or apoptotic cells through trans-activation of LRP on the phagocyte. *Cell*. 2005 Oct 21;123(2):321-34.
  44. Yatim N, Cullen S, Albert ML. Dying cells actively regulate adaptive immune responses. *Nat Rev Immunol*. 2017 Apr;17(4):262-275. doi: 10.1038/nri.2017.9.
  45. Green DR, Ferguson T, Zitvogel L, Kroemer G. Immunogenic and tolerogenic cell death. *Nat Rev Immunol*. 2009 May;9(5):353-63. doi: 10.1038/nri2545
  46. Nowak AK, Lake RA, Marzo AL, et al. Induction of Tumor Cell Apoptosis In Vivo Increases Tumor Antigen Cross-Presentation, Cross-Priming Rather than Cross-Tolerizing Host Tumor-Specific CD8 T Cells. *J Immunol*. 2003 May 15;170(10):4905-13
  47. Lugade AA, Moran JP, Gerber SA, et al. Local Radiation Therapy of B16 Melanoma Tumors Increases the Generation of Tumor Antigen-Specific Effector Cells That Traffic to the Tumor. *J Immunol*. 2005 Jun 15;174(12):7516-23.
  48. Petrovski G, Ayna G, Majaj G, et al. Phagocytosis of cells dying through autophagy induces inflammasome activation and IL-1 $\beta$  release in human macrophages. *Autophagy*. 2011;7(3):321-30.
  49. Ayna G, Krysko D V., Kaczmarek A, ATP release from dying autophagic cells and their phagocytosis are crucial for inflammasome activation in macrophages *PLoS One*. 2012;7(6):e40069. doi: 10.1371/journal.pone.0040069.
  50. Galluzzi L, Buqué A, Kepp O, et al. Immunogenic cell death in cancer and infectious disease. *Nat Rev Immunol*. 2017 Feb;17(2):97-111. doi: 10.1038/nri.2016.107.
  51. Thompson CB. Apoptosis in the pathogenesis and treatment of disease. *Science*. 1995 Mar 10;267(5203):1456-62.
  52. Zitvogel L, Casares N, Péquignot MO, et al. Immune response against dying tumor cells. *Adv Immunol*. 2004;84:131-79.
  53. Tang D, Kang R, Coyne CB, et al. PAMPs and DAMPs: Signal 0s that spur autophagy and immunity. *Immunol Rev*. 2012 Sep;249(1):158-75. doi: 10.1111/j.1600-065X.2012.01146.x.
  54. Hou W, Zhang Q, Yan Z, et al. Strange attractors: DAMPs and autophagy link tumor cell death and immunity. *Cell Death Dis*. 2013 Dec 12;4:e966. doi: 10.1038/cddis.2013.493.
  55. Yatim N, Jusforgues-Saklani H, Orozco S, et al. RIPK1 and NF- $\kappa$ B signaling in dying cells determines cross-priming of CD8+ T cells. *Science*. 2015 Oct 16;350(6258):328-34. doi: 10.1126/science.aad0395.

56. Ahn J, Xia T, Rabasa Capote A, et al. Extrinsic Phagocyte- Dependent STING Signaling Dictates the Immunogenicity of Dying Cells. *Cancer Cell*. 2018 May 14;33(5):862-873.e5. doi: 10.1016/j.ccell.2018.03.027.
57. Ma Y, Aymeric L, Locher C, et al. Contribution of IL-17- producing gamma delta T cells to the efficacy of anticancer chemotherapy. *J Exp Med*. 2011 Mar 14;208(3):491-503. doi: 10.1084/jem.20100269.
58. Ren J, Jia X, Zhao Y, et al. The RIP3-RIP1-NF- $\kappa$ B signaling axis is dispensable for necroptotic cells to elicit cross-priming of CD8+ T cells. *Cell Mol Immunol*. 2017 Jul;14(7):639-642. doi: 10.1038/cmi.2017.31.
59. Obeid M, Tesniere A, Ghiringhelli F, et al. Calreticulin exposure dictates the immunogenicity of cancer cell death. *Nat Med*. 2007 Jan;13(1):54-61.
60. Michaud M, Martins I, Sukkurwala AQ, et al. Autophagy- dependent anticancer immune responses induced by chemotherapeutic agents in mice. *Science*. 2011 Dec 16;334(6062):1573-7. doi: 10.1126/science.1208347.
61. Vacchelli E, Ma Y, Baracco EE, et al. Chemotherapy-induced antitumor immunity requires formyl peptide receptor 1. *Science*. 2015 Nov 20;350(6263):972-8. doi: 10.1126/science.aad0779.
62. Apetoh L, Ghiringhelli F, Tesniere A, et al. Toll-like receptor 4- dependent contribution of the immune system to anticancer chemotherapy and radiotherapy. *Nat Med*. 2007 Sep;13(9):1050-9.
63. Yang M, Li C, Zhu S, et al. TFAM is a novel mediator of immunogenic cancer cell death. *Oncoimmunology*. 2018 Feb 15;7(6):e1431086. doi: 10.1080/2162402X.2018.1431086.
64. Kang R, Chen R, Zhang Q et al. HMGB1 in health and disease. *Mol Aspects Med*. 2014 Dec;40:1-116. Doi: 10.1016/j.mam.2014.05.001.
65. Kazama H, Ricci JE, Herndon JM, et al. Induction of Immunological Tolerance by Apoptotic Cells Requires Caspase- Dependent Oxidation of High-Mobility Group Box-1 Protein. *Immunity*. 2008 Jul 18;29(1):21-32. doi: 10.1016/j.immuni.2008.05.013.
66. Li C, Zhang Y, Cheng X, et al. PINK1 and PARK2 Suppress Pancreatic Tumorigenesis through Control of Mitochondrial Iron- Mediated Immunometabolism. *Dev Cell*. 2018 Aug 20;46(4):441-455.e8. doi: 10.1016/j.devcel.2018.07.012.
67. Ito T, Kawahara KI, Okamoto K, et al. Proteolytic cleavage of high mobility group box 1 protein by thrombin-thrombomodulin complexes. *Arterioscler Thromb Vasc Biol*. 2008 Oct;28(10):1825-30. doi: 10.1161/ATVBAHA.107.150631.
68. Yu Y, Tang D, Kang R. Oxidative stress-mediated HMGB1 biology. *Front Physiol*. 2015 Apr 7;6:93. doi: 10.3389/fphys.2015.00093.
69. Vaux DL, Korsmeyer SJ. Cell death in development. *Cell*. 1999 Jan 22;96(2):245-54.
70. Franz TA, Kidson SH. Mapping of interdigital apoptosis in the chick and duck hindlimb. *Embryology*. 93(2):85-94. 1997.
71. Marti A, Ritter PM, Jäger R, et al. Mouse mammary gland involution is associated with cytochrome c release and caspase activation. *Mech Dev*. 2001 Jun;104(1-2):89-98.
72. LeBlanc A, Liu H, Goodyer C, et al. Caspase-6 role in apoptosis of human neurons, amyloidogenesis, and Alzheimer's disease. *J Biol Chem*. 1999 Aug 13;274(33):23426-36.
73. Duprez L, Wirawan E, Berghe T Vanden, et al. Major cell death pathways at a glance. *Microbes Infect*. 2009 Nov;11(13):1050-62. doi: 10.1016/j.micinf.2009.08.013.
74. Movassagh M, Foo RSY. Simplified apoptotic cascades. *Heart Fail Rev*. 2008 Jun;13(2):111-9.
75. Adams JM, Cory S. Life-or-death decisions by the Bcl-2 protein family. *Trends Biochem Sci*. 2001 Jan;26(1):61-6.
76. Adrain C, Martin SJ. The mitochondrial apoptosome: A killer unleashed by the cytochrome seas. *Trends Biochem Sci*. 2001 Jun;26(6):390-7.
77. Spierings DC, De Vries EG, Vellenga E, et al. Tissue distribution of the death ligand TRAIL and its receptors. *J Histochem Cytochem*. 2004 Jun;52(6):821-31.
78. Vousden KH, Lu X. Live or let die: The cell's response to p53. *Nat Rev Cancer*. 2002 Aug;2(8):594-604.

79. Curtin JF, Cotter TG. Live and let die: Regulatory mechanisms in Fas-mediated apoptosis. *Cell Signal*. 2003 Nov;15(11):983-92.
80. Coşkun G, Özgür H. Apoptoz ve Nekrozun Moleküler Mekanizması. *Arşiv Kaynak Tarama Derg*. 2011, 20:145.
81. Martinvalet D, Zhu P, Lieberman J. Granzyme A induces caspase- independent mitochondrial damage, a required first step for apoptosis. *Immunity*. 2005 Mar;22(3):355-70.
82. Vercammen D, Vandenabeele P, Beyaert R, et al. Tumour necrosis factor-induced necrosis versus anti-Fas-induced apoptosis in L929 cells. *Cytokine*. 1997 Nov;9(11):801-8.
83. Vercammen D, Brouckaert G, Denecker G, et al. Dual signaling of the Fas receptor: Initiation of both apoptotic and necrotic cell death pathways. *J Exp Med*. 1998 Sep 7;188(5):919-30.
84. Galluzzi L, Kepp O, Krautwald S, et al. Molecular mechanisms of regulated necrosis. *Semin Cell Dev Biol*. 2014 Nov;35:24-32. doi: 10.1016/j.semcdb.2014.02.006.
85. Degtrev A, Hitomi J, Germscheid M, et al. Identification of RIP1 kinase as a specific cellular target of necrostatins. *Nat Chem Biol*. 2008 May;4(5):313-21. doi: 10.1038/nchembio.83.
86. Kaczmarek A, Vandenabeele P, Krysko D V. Necroptosis: The Release of Damage-Associated Molecular Patterns and Its Physiological Relevance. *Immunity*. 2013 Feb 21;38(2):209-23. doi: 10.1016/j.immuni.2013.02.003.
87. Weinlich R, Oberst A, Beere HM, et al. Necroptosis in development, inflammation and disease. *Nat Rev Mol Cell Biol*. 2017 Feb;18(2):127-136. doi: 10.1038/nrm.2016.149.
88. Dixon SJ. Ferroptosis: bug or feature? *Immunol Rev*. 2017 May;277(1):150-157. doi: 10.1111/imr.12533.
89. Stockwell BR, Friedmann Angeli JP, Bayir H, et al. Ferroptosis: A Regulated Cell Death Nexus Linking Metabolism, Redox Biology, and Disease. *Cell*. 2017 Oct 5;171(2):273-285. doi: 10.1016/j.cell.2017.09.021.
90. Dixon SJ, Lemberg KM, Lamprecht MR, et al. Ferroptosis: An iron-dependent form of nonapoptotic cell death. *Cell*. 2012 May 25;149(5):1060-72. doi: 10.1016/j.cell.2012.03.042.
91. Linkermann A, Skouta R, Himmerkus N, et al. Synchronized renal tubular cell death involves ferroptosis. *Proc Natl Acad Sci U S A*. 2014 Nov 25;111(47):16836-41. doi: 10.1073/pnas.1415518111.
92. Kim SE, Zhang L, Ma K, et al. Ultrasmall nanoparticles induce ferroptosis in nutrient-deprived cancer cells and suppress tumour growth. *Nat Nanotechnol*. 2016 Nov;11(11):977-985. doi: 10.1038/nnano.2016.164.
93. Jorgensen I, Miao EA. Pyroptotic cell death defends against intracellular pathogens. *Immunol Rev*. 2015 May;265(1):130-42. doi: 10.1111/imr.12287.
94. Bergsbaken T, Fink SL, Cookson BT. Pyroptosis: Host cell death and inflammation. *Nat Rev Microbiol*. 2009 Feb;7(2):99-109. doi: 10.1038/nrmicro2070.
95. Zychlinsky A, Prevost MC, Sansonetti PJ. *Shigella flexneri* induces apoptosis in infected macrophages. *Nature*. 1992 Jul 9;358(6382):167-9
96. Shi J, Gao W, Shao F. Pyroptosis: Gasdermin-Mediated Programmed Necrotic Cell Death. *Trends Biochem Sci*. 2017 Apr;42(4):245-254. doi: 10.1016/j.tibs.2016.10.004.
97. Shi J, Zhao Y, Wang Y, Gao W, Ding J, et al. Inflammatory caspases are innate immune receptors for intracellular LPS. *Nature*. 2014 Oct 9;514(7521):187-92. doi: 10.1038/nature13683.
98. Aziz M, Jacob A, Wang P. Revisiting caspases in sepsis. *Cell Death Dis*. 2014 Nov 20;5:e1526. doi: 10.1038/cddis.2014.488.
99. Vanden Berghe T, Demon D, Bogaert P, et al. Simultaneous targeting of IL-1 and IL-18 is required for protection against inflammatory and septic shock. *Am J Respir Crit Care Med*. 2014 Feb 1;189(3):282-91. doi: 10.1164/rccm.201308-1535OC.
100. Fatokun AA, Dawson VL, Dawson TM. Parthanatos: Mitochondrial-linked mechanisms and therapeutic opportunities. *British Br J Pharmacol*. 2014 Apr;171(8):2000-16. doi: 10.1111/bph.12416.
101. Wan Q, Liu J, Zheng Z, et al. Regulation of myosin activation during cell-cell contact for-

- mation by Par3-Lgl antagonism: Entosis without matrix detachment. *Mol Biol Cell*. 2012 Jun;23(11):2076-91. doi: 10.1091/mbc.E11-11-0940.
102. Durgan J, Tseng YY, Hamann JC, et al. Mitosis can drive cell cannibalism through entosis. *Elife*. 2017 Jul 11;6. pii: e27134. doi: 10.7554/eLife.27134.
103. Krishna S, Overholtzer M. Mechanisms and consequences of entosis. *Cell Mol Life Sci*. 2016 Jun;73(11-12):2379-86. doi: 10.1007/s00018-016-2207-0.
104. Florey O, Kim S, Overholtzer M. Entosis: Cell-in-Cell Formation that Kills Through Entotic Cell Death. *Curr Mol Med*. 2015;15(9):861-6.
105. Pérez E, Bergmann A. Intercellular cannibalism fuels tumor growth. *Cell Death Differ*. 2017 May;24(5):759-760. doi: 10.1038/cdd.2017.39.
106. Brinkmann V, Zychlinsky A. Neutrophil extracellular traps: Is immunity the second function of chromatin? *J Cell Biol*. 2012 Sep 3;198(5):773-83. doi: 10.1083/jcb.201203170.
107. Remijsen Q, Kuijpers TW, Wirawan E, et al. Dying for a cause: NETosis, mechanisms behind an antimicrobial cell death modality. *Cell Death Differ*. 2011 Apr;18(4):581-8. doi: 10.1038/cdd.2011.1.
108. Aits S, Jäättelä M. Lysosomal cell death at a glance. *J Cell Sci*. 2013 May 1;126(Pt 9):1905-12. doi: 10.1242/jcs.091181.
109. Serrano-Puebla A, Boya P. Lysosomal membrane permeabilization in cell death: new evidence and implications for health and disease. *Ann N Y Acad Sci*. 2016 May;1371(1):30-44. doi: 10.1111/nyas.12966.
110. Song X, Zhu S, Xie Y, et al. JTC801 Induces pH-dependent Death Specifically in Cancer Cells and Slows Growth of Tumors in Mice. *Gastroenterology*. 2018 Apr;154(5):1480-1493. Doi: 10.1053/j.gastro.2017.12.004.
111. Holze C, Michaudel C, MacKowiak C, et al. Oxeiptosis, a ROS- induced caspase-independent apoptosis-like cell-death pathway article. *Nat Immunol*. 2018 Feb;19(2):130-140. doi: 10.1038/s41590-017-0013-y.
112. Campisi J. Aging, Cellular Senescence, and Cancer. *Annu Rev Physiol*. 2013;75:685-705. doi: 10.1146/annurev-physiol-030212-183653.
113. Sharpless NE, Sherr CJ. Forging a signature of in vivo senescence. *Nat Rev Cancer*. 2015 Jul;15(7):397-408. doi: 10.1038/nrc3960.
114. Castedo M, Perfettini JL, Roumier T, et al. Cell death by mitotic catastrophe: A molecular definition. *Oncogene*. 2004 Apr 12;23(16):2825- 37.
115. Vitale I, Galluzzi L, Castedo M, et al. Mitotic catastrophe: a mechanism for avoiding genomic instability. *Nat Rev Mol Cell Biol*. 2011 Jun;12(6):385-92. doi: 10.1038/nrm3115.
116. Dominguez-Brauer C, Thu KL, Mason JM, et al. Targeting Mitosis in Cancer: Emerging Strategies. *Mol Cell*. 2015 Nov 19;60(4):524-36. doi: 10.1016/j.molcel.2015.11.006.
117. Neelsen KJ, Zanini IMY, Herrador R, et al. Oncogenes induce genotoxic stress by mitotic processing of unusual replication intermediates. *J Cell Biol*. 2013 Mar 18;200(6):699-708. doi: 10.1083/jcb.201212058.