



2. Bölüm

İSKEMİ REPERFÜZYONDA DİKKAT EDİLMESİ GEREKEN HUSUSLAR

Ümmü Gülsen BOZOK¹

Mustafa ARSLAN²

Ayşegül KÜÇÜK³

Preklinik çalışmalarında fizyolojik ve patolojik mekanizmaları öğrenmek, patolojik mekanizmalara tanı koymak, yeni tanı ve tedavi yöntemleri geliştirmek amacıyla hayvan modelleri oluşturulur (1). Hayvan modeli oluştururken; hayvanın türü, cinsi, yaşı, cinsiyeti ve çevresel faktörler çok iyi belirlenmelidir.

2.1. Tür-Cins

Hayvan türünün seçiminde, elde edilen sonuçların kliniğe uyarlanabilirliği, hayvanın anatomik ve fizyolojik özellikleri göz önünde bulundurulmalıdır. Üreme fizyolojisi ve büyümeye hızları iyi değerlendirilmelidir. Seçilen türün yapılacak cerrahi işleme uygunluğu, hayvanın anestezije dayanıklılığı önceden belirlenmelidir.

Hayvan modeli oluştururken cins ve suş farklılıklarını da önemlidir. Bir cinste alınan sonuçların başka bir cinste alınamayabileceği akılda tutulmalıdır. Suşlar arasında derin genetik farklılıklar bulunmaktadır. Özellikle transgenik çalışmalararda, sadece tek gen mutasyonu değil, genetik arka plan da dikkate alınmalıdır. Beyin iskemisi başta olmak üzere iskemi reperfüzyon (I/R) çalışmalarında doğru sonuç alabilmek için, birden fazla suşun sonuçları değerlendirilmelidir (2).

¹ Öğr. Gör. Dr., Kütahya Sağlık Bilimleri Üniversitesi Tıp Fakültesi, Fizyoloji AD., gulsenozsoy07@gmail.com

² Doç. Dr., Gazi Üniversitesi Tıp Fakültesi, Anesteziyoloji ve Reanimasyon AD., mustarslan@gmail.com

³ Prof. Dr., Kütahya Sağlık Bilimleri Üniversitesi Tıp Fakültesi, Fizyoloji AD., kucukaysegul@hotmail.com

- verici sonuçlardan sonra, insan dışı primatlarda yapılan çalışmalar (avantajlar: jirensefyalik, insan durumuna daha yakın) düşünülmelidir.
2. Cins seçilmelidir. İdeal olarak, sonuçlar diğer cinslerle doğrulanmalıdır.
 3. Genç erkek hayvanlar tercih edilebilir (avantajlar: homojenlik, tekrarlanabilirlik). Dişi, yaşlı veya komorbid hayvanlarda ek çalışmalar yapılabilir (avantaj: klinik duruma daha yakın).
 4. İ/R modeli seçilmelidir. Seçilen İ/R modeli, beklenen klinik durumla patofizyolojik olarak alakalı olmalıdır ve çalışılan ilaç veya müdahale varsayılan mekanizmasıyla eşleşmelidir.
 5. Anestezik ve anestezi yöntemi seçilmelidir. İnhalasyon anestezisi, i.v. veya i.p. ilaç uygulamasına tercih edilebilir. Anesteziklerin İ/R modelinde çalışılacak olan ilaç ile etkileşimine dikkat edilmelidir.
 6. Farmakokinetik ve doz-yanıt eğrileri belirlenmelidir. Bunlardan dozaj ve terapötik zaman penceresilarındaki bilgiler elde edilebilir.
 7. Deney esnasında fizyolojik parametreler kontrol edilmelidir. Özellikle hipotermiden kaçınılmalıdır.
 8. Sonuç analizi için iskemik alanın boyutu, fonksiyon skorları, histolojik, biyokimyasal ve moleküler değerlendirme yapılabilir.

KAYNAKLAR

1. Kaya M, Çevik A. Hayvan deneylerinde planlanma ve model seçimi. Deneysel Tip Araştırma Enstitüsü Dergisi 2011;1(2): 36-9.
2. Braeuninger S, Kleinschmitz C. Rodent models of focal cerebral ischemia: procedural pitfalls and translational problems. Exp Transl Stroke Med 2009;1:8. doi: 10.1186/2040-7378-1-8.
3. Fagundes DJ, Taha MO. Animal disease model: choice's criteria and current animals specimens. Acta Cirúrgica Bras 2004;19(1):59-65.
4. Camacho P, Fan H, Liu Z, He JQ. Small mammalian animal models of heart disease. Am J Cardiovasc Dis 2016;6(3):70-80.
5. Milani-Nejad N, Janssen PM. Small and large animal models in cardiac contraction research: advantages and disadvantages. Pharmacol Ther 2014;141(3):235-49.
6. Yeğen BC, Gören MZ. Biyomedikal Araştırmalarda Deney Hayvanı Temel Bilgiler ve Etik İlkeler. Yüce Yayımlı. 2005; 54-64.
7. Durukan A, Tatlisumak T. Acute ischemic stroke: overview of major experimental rodent models, pathophysiology, and therapy of focal cerebral ischemia. Pharmacol Biochem Behav 2007;87(1):179-97.
8. Rumajogee P, Bregman T, Miller SP, Yager JY, Fehlings MG. Rodent Hypoxia-Ischemia Models for Cerebral Palsy Research: A Systematic Review. Front Neurol 2016;7:57. doi: 10.3389/fneur.2016.00057.
9. Kim SK, Cho KO, Kim SY. White Matter Damage and Hippocampal Neurodegeneration Induced by Permanent Bilateral Occlusion of Common Carotid Artery in the Rat: Comparison between Wistar and WW Dawley Strain. Korean J Physiol Pharmacol 2008;12(3):89-94.

10. Verdouw PD, van den Doel MA, de Zeeuw S, Duncker DJ. Animal models in the study of myocardial ischaemia and ischaemic syndromes. *Cardiovasc Res* 1998;39(1):121-35.
11. Wang J, Zhang P, Tang Z. Animal models of transient ischemic attack: a review. *Acta Neurol Belg* 2020;120(2):267-75.
12. Kher A, Meldrum KK, Wang M, Tsai BM, Pitcher JM, Meldrum DR. Cellular and molecular mechanisms of sex differences in renal ischemia-reperfusion injury. *Cardiovasc Res* 2005;67(4):594-603.
13. Müller V, Losonczy G, Heemann U, Vannay A, Fekete A, Reusz G, et al. Sexual dimorphism in renal ischemia-reperfusion injury in rats: possible role of endothelin. *Kidney Int* 2002;62(4):1364-71.
14. Park KM, Kim JI, Ahn Y, Bonventre AJ, Bonventre JV. Testosterone is responsible for enhanced susceptibility of males to ischemic renal injury. *J Biol Chem* 2004;279(50):52282-92.
15. Wang M, Tsai BM, Kher A, Baker LB, Wairiuko GM, Meldrum DR. Role of endogenous testosterone in myocardial proinflammatory and proapoptotic signaling after acute ischemia-reperfusion. *Am J Physiol Heart Circ Physiol* 2005;288(1):H221-6.
16. Takaoka M, Yuba M, Fujii T, Ohkita M, Matsumura Y. Oestrogen protects against ischaemic acute renal failure in rats by suppressing renal endothelin-1 overproduction. *Clin Sci (Lond)* 2002;103 Suppl 48:434S-7S.
17. Lee TM, Su SF, Tsai CC, Lee YT, Tsai CH. Cardioprotective effects of 17 beta-estradiol produced by activation of mitochondrial ATP-sensitive K(+)Channels in canine hearts. *J Mol Cell Cardiol* 2000;32(7):1147-58.
18. Er F, Michels G, Gassanov N, Rivero F, Hoppe UC. Testosterone induces cytoprotection by activating ATP-sensitive K⁺ channels in the cardiac mitochondrial inner membrane. *Circulation* 2004;110(19):3100-7.
19. Patten RD, Pourati I, Aronovitz MJ, Baur J, Celestin F, Chen X, et al. 17beta-estradiol reduces cardiomyocyte apoptosis in vivo and in vitro via activation of phospho-inositol-3 kinase/Akt signaling. *Circ Res* 2004;95(7):692-9.
20. Nuedling S, Kahlert S, Loebbert K, Meyer R, Vetter H, Grohé C. Differential effects of 17beta-estradiol on mitogen-activated protein kinase pathways in rat cardiomyocytes. *FEBS Lett* 1999;454(3):271-6.
21. Wang M, Baker L, Tsai BM, Meldrum KK, Meldrum DR. Sex differences in the myocardial inflammatory response to ischemia-reperfusion injury. *Am J Physiol Endocrinol Metab* 2005;288(2):E321-6.
22. Fraser H, Davidge ST, Clanachan AS. Activation of Ca(2+)-independent nitric oxide synthase by 17beta-estradiol in post-ischemic rat heart. *Cardiovasc Res* 2000;46(1):111-8.
23. Simoncini T, Hafezi-Moghadam A, Brazil DP, Ley K, Chin WW, Liao JK. Interaction of oestrogen receptor with the regulatory subunit of phosphatidylinositol-3-OH kinase. *Nature* 2000;407(6803):538-41.
24. Fekete A, Vannay A, Vér A, Vásárhelyi B, Müller V, Ouyang N, et al. Sex differences in the alterations of Na(+), K(+)-ATPase following ischaemia-reperfusion injury in the rat kidney. *J Physiol* 2004;555(Pt 2):471-80.
25. Cross HR, Lu L, Steenbergen C, Philipson KD, Murphy E. Overexpression of the cardiac Na+/Ca²⁺ exchanger increases susceptibility to ischemia/reperfusion injury in male, but not female, transgenic mice. *Circ Res* 1998;83(12):1215-23.
26. Sugishita K, Li F, Su Z, Barry WH. Anti-oxidant effects of estrogen reduce [Ca²⁺]i during metabolic inhibition. *J Mol Cell Cardiol* 2003;35(3):331-6.
27. Sugishita K, Su Z, Li F, Philipson KD, Barry WH. Gender influences [Ca²⁺]i during metabolic inhibition in myocytes overexpressing the Na(+)-Ca²⁺ exchanger. *Circulation* 2001;104(17):2101-6.

28. Dubey RK, Jackson EK, Keller PJ, Imthurn B, Rosselli M. Estradiol metabolites inhibit endothelin synthesis by an estrogen receptor-independent mechanism. *Hypertension* 2001;37(2 Pt 2):640-4.
29. Minamoto K, Pinsky DJ, Fujita T, Naka Y. Timing of nitric oxide donor supplementation determines endothelin-1 regulation and quality of lung preservation for transplantation. *Am J Respir Cell Mol Biol* 2002;26(1):14-21.
30. Lee TM, Chou TF, Tsai CH. Differential role of K(ATP) channels activated by conjugated estrogens in the regulation of myocardial and coronary protective effects. *Circulation* 2003;107(1):49-54.
31. Rahgozar M, Willgoss DA, Gobé GC, Endre ZH. ATP-dependent K⁺ channels in renal ischemia reperfusion injury. *Ren Fail* 2003;25(6):885-96.
32. Ceballos G, Figueiroa L, Rubio I, Gallo G, Garcia A, Martinez A, et al. Acute and nongenomic effects of testosterone on isolated and perfused rat heart. *J Cardiovasc Pharmacol* 1999;33(5):691-7.
33. Harris NR, Rumbaut RE. Age-related responses of the microcirculation to ischemia-reperfusion and inflammation. *Pathophysiology* 2001;8(1):1-10.
34. Sengupta P. The Laboratory Rat: Relating Its Age With Human's. *Int J Prev Med* 2013;4(6):624-30.
35. Sengupta P. A Scientific Review of Age Determination for a Laboratory Rat: How old is it in comparison with Human age? *Biomed Int* 2012;2:81-9.
36. Spear LP. The adolescent brain and age-related behavioral manifestations. *Neurosci Biobehav Rev* 2000;24(4):417-63.
37. Maggioni AP, Maseri A, Fresco C, Franzosi MG, Mauri F, Santoro E, et al. Age-related increase in mortality among patients with first myocardial infarctions treated with thrombolysis. The Investigators of the Gruppo Italiano per lo Studio della Sopravvivenza nell'Infarto Miocardico (GISSI-2). *N Engl J Med* 1993;329(20):1442-8.
38. Lesnfsky EJ, Gallo DS, Ye J, Whittingham TS, Lust WD. Aging increases ischemia-reperfusion injury in the isolated, buffer-perfused heart. *J Lab Clin Med* 1994;124(6):843-51.
39. Ataka K, Chen D, Levitsky S, Jimenez E, Feinberg H. Effect of aging on intracellular Ca²⁺, pH_i, and contractility during ischemia and reperfusion. *Circulation* 1992;86(5 Suppl):II371-6.
40. Drozdowski L, Thomson AB. Aging and the intestine. *World J Gastroenterol* 2006;12(47):7578-84.
41. Kusaka J, Koga H, Hagiwara S, Hasegawa A, Kudo K, Noguchi T. Age-dependent responses to renal ischemia-reperfusion injury. *J Surg Res* 2012;172(1):153-8.
42. Okaya T, Blanchard J, Schuster R, Kuboki S, Husted T, Caldwell CC, et al. Age-dependent responses to hepatic ischemia/reperfusion injury. *Shock* 2005;24(5):421-7.
43. Boraschi D, Aguado MT, Dutel C, Gorony J, Louis J, Grubeck-Loebenstein B, et al. The gracefully aging immune system. *Sci Transl Med* 2013;5(185):185ps8. doi: 10.1126/scitranslmed.3005624.
44. Zalewski CK. Aging of the Human Vestibular System. *Semin Hear* 2015;36(3):175-96.
45. Cannon JG, Fiatarone MA, Meydani M, Gong J, Scott L, Blumberg JB, et al. Aging and dietary modulation of elastase and interleukin-1 beta secretion. *Am J Physiol* 1995;268(1 Pt 2):R208-13.
46. Sohal RS, Ku HH, Agarwal S, Forster MJ, Lal H. Oxidative damage, mitochondrial oxidant generation and antioxidant defenses during aging and in response to food restriction in the mouse. *Mech Ageing Dev* 1994;74(1-2):121-33.
47. Azhar S, Cao L, Reaven E. Alteration of the adrenal antioxidant defense system during aging in rats. *J Clin Invest* 1995;96(3):1414-24.
48. Xia E, Rao G, Van Remmen H, Heydari AR, Richardson A. Activities of antioxidant enzymes in various tissues of male Fischer 344 rats are altered by food restriction. *J Nutr* 1995;125(2):195-201.

49. Sabatino F, Masoro EJ, McMahan CA, Kuhn RW. Assessment of the role of the glucocorticoid system in aging processes and in the action of food restriction. *J Gerontol* 1991;46(5):B171-9.
50. House SD, Ruch S, Koscienski WF 3rd, Rocholl CW, Moldow RL. Effects of the circadian rhythm of corticosteroids on leukocyte-endothelium interactions in the AM and PM. *Life Sci* 1997;60(22):2023-34.
51. Williams RM, Singh J, Sharkey KA. Innervation and mast cells of the rat exorbital lacrimal gland: the effects of age. *J Auton Nerv Syst* 1994;47(1-2):95-108.
52. Shepherd RK, Duling BR. Inosine-induced vasoconstriction is mediated by histamine and thromboxane derived from mast cells. *Am J Physiol* 1996;270(2 Pt 2):H560-6.
53. Lee JY, Lee MY, Chung SM, Chung JH. Chemically induced platelet lysis causes vasoconstriction by release of serotonin. *Toxicol Appl Pharmacol* 1998;149(2):235-42.
54. Myśliwska J, Bryl E, Foerster J, Myśliwski A. The upregulation of TNF alpha production is not a generalised phenomenon in the elderly between their sixth and seventh decades of life. *Mech Ageing Dev* 1999;107(1):1-14.
55. Roth-Isigkeit A, Schwarzenberger J, v Borstel T, Gehring H, Ocklitz E, Wagner K, et al. Perioperative cytokine release during coronary artery bypass grafting in patients of different ages. *Clin Exp Immunol* 1998;114(1):26-32.
56. van Ruiven R, Meijer GW, van Zutphen LFM, Ritskes-Hoitinga J. Adaptation period of laboratory animals after transport: A review. *Scand J Lab Anim Sci* 1996;23(4):185-90.
57. Obernier JA, Baldwin RL. Establishing an appropriate period of acclimatization following transportation of laboratory animals. *ILAR J* 2006;47(4):364-9.
58. Tuli JS, Smith JA, Morton DB. Stress measurements in mice after transportation. *Lab Anim* 1995;29(2):132-8.
59. Mistlberger RE, Rusak B. Circadian Rhythms in Mammals. Formal Properties and Environmental Influences. In: (ed), Meir H Kryger, Thomas Roth, William C. Dement, Principles and Practice of Sleep Medicine: Fifth Edition. Elsevier. 2011;363-375.
60. Weinert D, Eimert H, Erkert HG, Schneyer U. Resynchronization of the circadian corticosterone rhythm after a light/dark shift in juvenile and adult mice. *Chronobiol Int* 1994;11(4):222-31.
61. Turner JG, Parrish JL, Hughes LF, Toth LA, Caspary DM. Hearing in laboratory animals: strain differences and nonauditory effects of noise. *Comp Med* 2005;55(1):12-23.
62. Hawkins P. Recognizing and assessing pain, suffering and distress in laboratory animals: a survey of current practice in the UK with recommendations. *Lab Anim* 2002;36(4):378-95.
63. Animal Research Review. ARRP Guideline 20. Guidelines for the Housing of Rats in Scientific Institutions. Animal Welfare Branch, NSW Department of Primary Industries, Locked Bag 21, Orange NSW. 2007;1-72.
64. Schmitt U, Hiemke C. Strain differences in open-field and elevated plus-maze behavior of rats without and with pretest handling. *Pharmacol Biochem Behav* 1998;59(4):807-11.
65. Gouveia K, Hurst JL. Reducing mouse anxiety during handling: effect of experience with handling tunnels. *PLoS One* 2013;8(6):e66401. doi: 10.1371/journal.pone.0066401.
66. Percie du Sert N, Alfieri A, Allan SM, Carswell HV, Deuchar GA, Farr TD, et al. The IMPROVE Guidelines (Ischaemia Models: Procedural Refinements Of in Vivo Experiments). *J Cereb Blood Flow Metab* 2017;37(11):3488-517.
67. Sanger GJ, Holbrook JD, Andrews PL. The translational value of rodent gastrointestinal functions: a cautionary tale. *Trends Pharmacol Sci* 2011;32(7):402-9.
68. Longo VD, Mattson MP. Fasting: molecular mechanisms and clinical applications. *Cell Metab* 2014;19(2):181-92.
69. Gargiulo S, Greco A, Gramanzini M, Esposito S, Affuso A, Brunetti A, et al. Mice anesthesia, analgesia, and care, Part I: anesthetic considerations in preclinical research. *ILAR J* 2012;53(1):E55-69.

70. Saha JK, Xia J, Grondin JM, Engle SK, Jakubowski JA. Acute hyperglycemia induced by ketamine/xylazine anesthesia in rats: mechanisms and implications for preclinical models. *Exp Biol Med (Maywood)* 2005;230(10):777-84.
71. Gaines Das R, North D. Implications of experimental technique for analysis and interpretation of data from animal experiments: outliers and increased variability resulting from failure of intraperitoneal injection procedures. *Lab Anim* 2007;41(3):312-20.
72. Vijn PC, Sneyd JR. I.v. anaesthesia and EEG burst suppression in rats: bolus injections and closed-loop infusions. *Br J Anaesth* 1998;81(3):415-21.
73. Flecknell P. *Laboratory Animal Anaesthesia*. Third edition. Elsevier. 2009;1-304.
74. Zhu M, Nehra D, Ackerman JJH, Yablonskiy DA. On the role of anesthesia on the body/brain temperature differential in rats. *J Therm Biol* 2004;29(7-8):599-603.
75. Marion DW. Controlled normothermia in neurologic intensive care. *Crit Care Med* 2004;32(2 Suppl):S43-5.
76. Yenari MA, Han HS. Neuroprotective mechanisms of hypothermia in brain ischaemia. *Nat Rev Neurosci* 2012;13(4):267-78.
77. Zausinger S, Baethmann A, Schmid-Elsaesser R. Anesthetic methods in rats determine outcome after experimental focal cerebral ischemia: mechanical ventilation is required to obtain controlled experimental conditions. *Brain Res Brain Res Protoc* 2002;9(2):112-21.
78. Albrecht M, Henke J, Tacke S, Markert M, Guth B. Effects of isoflurane, ketamine-xylazine and a combination of medetomidine, midazolam and fentanyl on physiological variables continuously measured by telemetry in Wistar rats. *BMC Vet Res* 2014;10:198.