

### GİRİŞ

İskemi-reperfüzyon hasarı patofizyolojisinin iyi anlaşılmasının miyokardiyal hibernasyon, akut kalp yetmezliği, serebral disfonksiyon, gastrointestinal disfonksiyon, sistemik inflamatuvar yanıt sendromu(SIRS) ve çoklu organ yetmezliği (MOD) gibi hekimler için önemli sorunlara yeni tedavi girişimlerinin geliştirilmesine imkan sağlayacağına inanıyoruz.<sup>(1)</sup> Bu derlemede, iskemi-reperfüzyon hasarının temel patofizyolojisinden, özellikle reaktif oksijen türlerinin ve hücre ölüm yollarının tutulumundan ve antioksidanlar üzerine odaklanan son gelişmelerden bahsedilecektir.

### PATOFİZYOLOJİ

Arteryal kan akımında ki azalmaya bağlı oluşan iskemi hipoksiye ve mitokondrinin elektron transfer zincirinde fonksiyon bozulmasına öncülük eder. Mitokondrinin ATP üretimindeki düşüş anaerobik metabolizmayı, Na-K pompalarını ve ribozomların ayrılmasını uyarır. Anaerobik metabolizmaya bağlı olarak ATP ve antioksidanların az üretilmesi laktik asid retansiyonu ve metabolik asidoz ile sonuçlanır. Hücre zarındaki  $\text{Na}^+$ - $\text{K}^+$ -ATP, Na-H ve kalsiyum pompalarının bozulması; hücre içinde sodyum, hidrojen ve

kalsiyum birikmesine neden olur ki bu da hipe-rosmolariteyi arttırarak hücre içine su girişi ve hücre şişmesi ile sonuçlanır. Hidrojen retansiyonu da hücre PH'ını düşürerek enzim salınmasına ve nükleer kromotin kümelenmesine neden olur. Ribozomlardaki ayrılma, protein sentezini azaltır. Reperfüzyon aşamasıyla iskemik dokuya kan akışıyla oksijenizasyon sağlanır. İskemik dokudaki antioksidan ajanların düşük konsantrasyonlarına bağlı serbest oksijen radikalleri oluşumu artar. Serbest oksijen radikallerine bağlı oluşan oksidatif stres; endotelial disfonksiyon, DNA hasarı ve lokal inflamatuvar yanıt oluşmasıyla sonuçlanır. İnflamatuvar kaskad ve oksidatif stres sitokin fırtınasına bağlı hücre yapısındaki hasar ve hücre ölümüyle sonuçlanabilir <sup>(2)</sup>.(şekil-1)

### OKSİDATİF STRESİN İSKEMİ REPERFÜZYON HASARINDA Kİ YERİ:

Oksidatif stres enzimatik yada enzimatik olmayan yollardan gelişebilir. En bilinen enzimatik kaynaklar ksantin oksidaz, NADPH oksidaz, mitokondriyal elektron transfer zinciri ve nitrik oksit sentazdır (NOS). Ksantin oksidaz, NADPH oksidaz, ve mitokondriyal elektron transfer zinciri, barsaklar akciğer, kalp, beyin, kas, mide,

lendirebilir. Renal IR yaralanmasının yeni tanı biyolojik belirteçlerini temsil edebilirler<sup>(103)</sup>

Diyabetik böbrek, artmış oksidatif stres ve bozulmuş nükleer faktör eritroid 2 ile ilişkili faktör 2 (Nrf2)/heme oksijenaz-1 (HO-1) sin-yalizasyonu ile ilişkili iskemi/reperfüzyon (I/R) yaralanmasına daha duyarlıdır.<sup>(102)</sup>

### Sonuç

Şu anda, MI hastalarının I/R yaralanmasına karşı önlem olarak soğutulması, soğutma birkaç potansiyel reperfüzyon yaralanma yolunu değiştirdiğinden miyokard kurtarma için en büyük potansiyele sahip görünmektedir. Bu devam eden çabalar başarısız olursa, daha fazla müdahaleyi dahil etmeye cesaret eden adaptif tasarıma sahip daha büyük denemeler ileriye doğru olmalıdır.

**Anahtar Kelimeler:** iskemi, reperfüzyon, antioksidanlar

### KAYNAKLAR

1. Wua M.Y, Yianga G.T, LiaocW.T ,et al. Current Mechanistic Concepts in Ischemia and Reperfusion Injury. *Physiol Biochem.* 2018;46(4):1650-1667. doi: 10.1159/000489241. Epub 2018 Apr 20.
2. Takiya CM, Morales MM: Bone Marrow-Derived Mononuclear Cell Therapy Accelerates Renal Ischemia Reperfusion Injury Recovery by Modulating Inflammatory, Antioxidant and Apoptotic Related Molecules. *Cell Physiol Biochem* 2017;41:1736-1752.
3. Granger DN, Kvietys PR: Reperfusion injury and reactive oxygen species: The evolution of a concept. *Redox Biol* 2015;6:524-551
4. De Pascali F, Hemann C, Samons K, et al: Hypoxia and reoxygenation induce endothelial nitric oxide synthase uncoupling in endothelial cells through tetrahydropterin depletion and S-glutathionylation. *Biochemistry* 2014;53:3679-3688.
5. Moens AL, Champion HC, Claeys MJ, et al: High-Dose Folic Acid Pretreatment Blunts Cardiac Dysfunction During Ischemia Coupled to Maintenance of High-Energy Phosphates and Reduces Postreperfusion Injury. *Circulation* 2008;117:1810-1819.
6. Perkins K-AA, Pershad S, Chen Q, et al: The effects of modulating eNOS activity and coupling in ischemia/reperfusion (I/R). *Naunyn Schmiedebergs Arch Pharmacol* 2012;385:27
7. Kalogeris T, Baines CP, Krenz M, et al: Ischemia/Reperfusion. *Compr Physiol.* 2016 Dec 6;7(1):113-170. doi: 10.1002/cphy.c160006.
8. Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: A delay of lethal cell injury in ischemic myocardium. *Circulation* 74: 1124-1136, 1986.
9. Depre C, Park JY, Shen Y T, et al. Molecular mechanisms mediating preconditioning following chronic ischemia differ from those in classical second window. *Am J Physiol Heart Circ Physiol* 299: H752-H762, 2010.
10. Depre C, Vatner SF. Cardioprotection in stunned and hibernating myocardium. *Heart Fail Rev* 12: 307-317, 2007.
11. Abela CB, Homer-Vanniasinkham S. Clinical implications of ischaemia-reperfusion injury. *Pathophysiology* 9: 229-240, 2003.
12. Burne MJ, Haq M, Matsuse H, et al Genetic susceptibility to renal ischemia reperfusion injury revealed in a murine model. *Transplantation* 69: 1023-1025, 2000.
13. Baumgartner WA, Williams GM, Fraser CD, Jr, et al Cardiopulmonary bypass with profound hypothermia. An optimal preservation method for multiorgan procurement. *Transplantation* 47, 123-127, 1989.
14. Taylor MJ, Baicu SC. Current state of hypothermic machine perfusion preservation of organs: The clinical perspective. *Cryobiology* 60: S2035, 2010.
15. Gaboury JP, Johnston B, Niu X. Mechanisms underlying acute mast cell-induced leukocyte rolling and adhesion in vivo. *J Immunol* 154: 804-813, 1995.
16. Barnabei MS, Palpant NJ, Metzger JM. Influence of genetic background on ex vivo and in vivo cardiac function in several commonly used inbred mouse strains. *Physiol Genomics* 42: 103-113, 2010.
17. Barone FC, Knudsen DJ, Nelson AH, et al. Mouse strain differences in susceptibility to cerebral ischemia are related to cerebral vascular anatomy. *J Cereb Blood Flow Metab* 13: 683-692, 1993.
18. Burne MJ, Haq M, Matsuse H, et al. Genetic susceptibility to renal ischemia reperfusion injury revealed in a murine model. *Transplantation* 69: 1023-1025, 2000.
19. Candilio L, Hausenloy DJ, Yellon DM. Remote ischemic conditioning: A clinical trial's update. *J Cardiovasc Pharmacol Ther* 16: 304-312, 2011.
20. Ordy JM, Wengenack TM, Bialobok P, et al. Selective vulnerability and early progression of hippocampal CA1 pyramidal cell degeneration and GFAP-positive astrocyte reactivity in the rat four-vessel occlusion model of transient global ischemia. *Exp Neurol* 119: 128-139, 1993.
21. Bluhmki E, Chamorro A, Davalos A, et al. Stroke treatment with alteplase given 3.0-4.5h after onset of acute ischemic stroke (ECASSIII): Additional outcomes and subgroup analysis of a randomised controlled trial. *Lancet Neurol* 8: 1095-1102, 2009.
22. Hacke W, Donnan G, Fieschi C, et al. Association of outcome with early stroke treatment: Pooled analysis of ATLANTIS, ECASS, and NINDS rt-PA stroke trials. *Lancet* 363: 768-74, 2004.
23. Kristi'an T. Metabolic stages, mitochondria and calcium in hypoxic/ischemic brain damage. *Cell Calcium* 36: 221-233, 2004.
24. Lee JM, Grabb MC, Zipfel GJ, et al. Brain tissue responses to ischemia. *J Clin Invest* 106: 723-731, 2000.
25. Adibhatha RM, Hatche, JF. Lipid oxidation and peroxidation in CNS health and disease: From molecu-

- lar mechanisms to therapeutic opportunities. *Antiox Redox Signal* 12: 125-169, 2010.
26. Damle SS, Moore EE, Babu AN, et al. Hemoglobin-based oxygen carrier induces heme oxygenase-1 in the heart and lung but not brain. *J Am Coll Surg* 208: 592-598, 2009.
  27. Choi DW. The role of glutamate neurotoxicity in hypoxic-ischemic neuronal death. *Annu Rev Neurosci* 13: 171-182, 1990.
  28. Boersma E, Maas AC, Deckers JW, et al. Early thrombolytic treatment in acute myocardial infarction: Reappraisal of the golden hour. *Lancet* 348: 771-775, 1996.
  29. LATE\_Study\_Group. Late Assessment of Thrombolytic Efficacy (LATE) study with alteplase 6-24 hours after onset of acute myocardial infarction. *Lancet* 342: 759-66, 1993.
  30. Wittnich C. Age-related differences in myocardial metabolism affects response to ischemia. Age in heart tolerance to ischemia. *Am J Cardiovasc Pathol* 4: 175-180, 1992.
  31. McDougal WS. Renal perfusion/reperfusion injuries. *J Urol* 140: 1325-30, 1988.
  32. Humphreys MR, Castle EP, Lohse CM, et al. Renal ischemia time in laparoscopic surgery: An experimental study in a porcine model. *Int J Urol* 16: 105-109, 2009.
  33. Cerqueira NE, Hussni CA, Yoshida WB. Pathophysiology of mesenteric ischemia/reperfusion: A review. *Acta Cir Bras* 20: 336-343, 2005.
  34. Ikeda H, Suzuki Y, Suzuki M et al. Apoptosis is a major mode of cell death caused by ischaemia and ischaemia/reperfusion injury to the rat intestinal epithelium. *Gut* 42: 530-537, 1998.
  35. Kinross J, Warren O, Basson S, et al. Intestinal ischemia/reperfusion injury: Defining the role of the gut microbiome. *Biomark Med* 3: 175-192, 2009.
  36. Souza DG, Vieira AT, Soares AC, et al. The essential role of the intestinal microbiota in facilitating acute inflammatory responses. *J Immunol* 173: 4137-4146, 2004.
  37. Sorkine P, Szold O, Halpern P, et al. Gut decontamination reduces bowel ischemia-induced lung injury in rats. *Chest* 112: 491-495, 1997.
  38. Yoshiya K, Lapchak PH, Thai TH., et al. Depletion of gut commensal bacteria attenuates intestinal ischemia/reperfusion injury. *Am J Physiol Gastrointest Liver Physiol* 301: G1020-G1030, 2011.
  39. Wagers AJ, Conboy IM. Cellular and molecular signatures of muscle regeneration: Current concepts and controversies in adult myogenesis. *Cell* 122: 659-667, 2005.
  40. Liu L, Kuberski P. Molecular mechanisms of leukocyte recruitment: Organ-specific mechanisms of action. *Thromb Haemost* 89: 213-220, 2003.
  41. Burns AR, Smith CW, Walker DC. Unique structural features that influence neutrophil emigration into the lung. *Physiol Rev* 83: 3093-336, 2003.
  42. Jong HR, Ko GJ, Wasowska BA, et al. The interaction between ischemia-reperfusion and immune responses in the kidney. *J Mol Med* 87: 859-864, 2009
  43. Yilmaz G, Granger DN. Cell adhesion molecules and ischemic stroke. *Neurol Res* 30: 783-793, 2008.
  44. Carden DL, Granger DN. Pathophysiology of ischemia-reperfusion injury. *J Pathol* 190: 255-266, 2000.
  45. Esme H, Fidan H, Koken T, Solak O. Effect of lung ischemia-reperfusion on oxidative stress parameters of remote tissues. *Eur J Cardio-Thoracic Surg* 29: 294-298, 2006.
  46. He C, Klionsky DJ. Regulation mechanisms and signaling pathways of autophagy. *Annu Rev Genet* 43: 67-93, 2009.
  47. Santora RJ, Lie ML, Grigoryev DN, et al. Therapeutic distant organ effects of regional hypothermia during mesenteric ischemia-reperfusion injury. *J Vasc Surg* 52: 1003-1014, 2010.
  48. Sorkine P, Szold O, Halpern P, et al. Gut decontamination reduces bowel ischemia-induced lung injury in rats. *Chest* 112: 491-495, 1997.
  49. Vega VL, Mardones L, Maldonado M, et al. Xanthine oxidase released from reperfused hind limbs mediate kupffer cell activation, neutrophil sequestration, and hepatic oxidative stress in rats subjected to tourniquet shock. *Shock* 14: 565-571, 2000.
  50. Courties G, Moskowitz MA, Nahrendorf M. The innate-immune system after ischemic injury: Lessons to be learned from the heart and brain. *JAMA Neurol* 71: 233-236, 2014.
  51. Deitch EA. Gut-origin sepsis: Evolution of a concept. *Surgeon* 10: 350-356, 2012.
  52. Stallion A, Kou TD, Latfi SQ, et al. Ischemia/reperfusion: A clinically relevant model of intestinal injury yielding systemic inflammation. *J Pediatr Surg* 40: 470-477, 2005.
  53. Deitch EA, Xu D, Kaise VL. Role for the gut in the development of injury- and shock-induced SIRS and MODS: The gut-lymph hypothesis, a review. *Front Biosci* 11: 520-528, 2006.
  54. Women's Health Research: Progress, Pitfalls, and Promise. Report Brief. c Institute of Medicine. Committee on Women's Health Research, National Academies Press, Washington, DC, 2010.
  55. Boengler K, Schulz R, Heusch G. Loss of cardioprotection with ageing. *Cardiovasc Res* 83: 247-261, 2009.
  56. Endres M, Ahmadi M, Kruman I. Folate deficiency increases postischemic brain injury. *Stroke* 36: 321-325, 2005.
  57. Feng M, Wang H, Wang Q, Guan W. Matrix metalloproteinase 9 promotes liver recovery from ischemia and reperfusion injury. *J Surg Res* 180: 156-161, 2013.
  58. Gundewar S, Calvert JW, Elrod JW, et al. Cytoprotective effects of N,N,N-trimethylsphingosine during ischemia-reperfusion injury are lost in the setting of obesity and diabetes. *Am J Physiol Heart Circ Physiol* 293: H2462-H2471, 2007.
  59. Roerecke M, Rehm J. Alcohol consumption, drinking patterns, and ischemic heart disease: A narrative review of meta-analyses and a systematic review and meta-analysis of the impact of heavy drinking occasions on risk for moderate drinkers. *BMC Med* 12: 182, 2014
  60. Rosen SE, Henry S, Bond R, et al. Sex-specific disparities in risk factors for coronary heart disease. *Curr Atheroscler*

- roscler Rep 17(8): 523, 2015.
61. Wever KE, Hooijmans CR, Rixen NP, et al. (2015) Determinants of the efficacy of cardiac ischemic preconditioning: a systematic review and meta-analysis of animal studies. *PLoS One* 10:e0142021. doi:10.1371/journal.pone.0142021
  62. Liu GS, Thornton J, Van Winkle D et al. (1991) Protection against infarction afforded by preconditioning is mediated by A1 adenosine receptors in rabbit heart. *Circulation* 84:350–356. doi:10.1161/01.CIR.84.1.350
  63. Marber MS, Latchman DS, Walker JM, et al. (1993) Cardiac stress protein elevation 24 hours after brief ischemia or heat stress is associated with resistance to myocardial infarction. *Circulation* 88:1264–1272. doi:10.1161/01.CIR.88.3.1264
  64. Kwan JC, Gao L, Macdonald PS, et al. (2015) Cardio-protective signalling by glyceryl trinitrate and cariporide in a model of donor heart preservation. *Heart Lung Circ* 24:306–318. doi:10.1016/j.hlc.2014.10.001
  65. Currie RW, Karmazyn M, Kloc M, et al. (1988) Heat-shock response is associated with enhanced postischemic ventricular recovery. *Circ Res* 63:543–549. doi:10.1161/01.RES.63.3.543
  66. Przyklenk K, Bauer B, Ovize M, et al. (1993) Regional ischemic 'preconditioning' protects remote virgin myocardium from subsequent sustained coronary occlusion. *Circulation* 87:893–899. doi:10.1161/01.CIR.87.3.893
  67. Whittaker P, Przyklenk K (1994) Reduction of infarct size in vivo with ischemic preconditioning: mathematical evidence for protection via non-ischemic tissue. *Basic Res Cardiol* 89:6–15. doi:10.1007/BF00788673
  68. Przyklenk K, Whittaker P (2013) Genesis of remote conditioning: action at a distance—hypotheses non fingo? *J Cardiovasc Med (Hagerstown)* 14:180–186. doi:10.2459/JCM.0b013e328358c8eb
  69. Cabrera-Fuentes HA, Alba-Alba C, Aragonés J, et al. (2016) Meeting report from the 2nd international symposium on new frontiers in cardiovascular research. Protecting the cardiovascular system from ischemia: between bench and bedside. *Basic Res Cardiol* 111:7. doi:10.1007/s00395-0150527-0
  70. Hausenloy DJ, Yellon DM (2008) Remote ischaemic preconditioning: underlying mechanisms and clinical application. *Cardiovasc Res* 79:377–386. doi:10.1093/cvr/cvn114
  71. Heusch G, Botker HE, Przyklenk K, et al. (2015) Remote ischemic conditioning. *J Am Coll Cardiol* 65:177–195. doi:10.1016/j.jacc.2014.10.031
  72. Pickard JM, Botker HE, Crimi G, et al. (2015) Remote ischemic conditioning: from experimental observation to clinical application: report from the 8th Biennial Hatter Cardiovascular Institute Workshop. *Basic Res Cardiol* 110:453. doi:10.1007/s00395-014-0453-6
  73. Sivaraman V, Pickard JM, Hausenloy DJ (2015) Remote ischaemic conditioning: cardiac protection from afar. *Anaesthesia* 70:732–748. doi:10.1111/anae.12973
  74. Bromage DJ, Pickard JM, Rosello X, et al. (2016) Remote ischaemic conditioning reduces infarct size in animal in vivo models of ischaemia reperfusion injury: a systematic review and meta-analysis. *Cardiovasc Res*. doi:10.1093/cvr/cvw143
  75. Piper HM, Garcia-Dorado D, Ovize M (1998) A fresh look at reperfusion injury. *Cardiovasc Res* 38:291–300. doi:10.1016/S0008-6363(98)00033-9
  76. Hausenloy DJ, Botker HE, Engstrom T, (2016) Targeting reperfusion injury in patients with ST-segment elevation myocardial infarction: trials and tribulations. *Eur Heart J*. doi:10.1093/eurheartj/ehw14
  77. Heusch G, Gersh BJ (2016) The pathophysiology of acute myocardial infarction and strategies of protection beyond reperfusion: a continual challenge. *Eur Heart J*. doi:10.1093/eurheartj/ehw224
  78. Heusch G, Rassaf T (2016) Time to give up on cardioprotection? A critical appraisal of clinical studies on ischemic pre-, post-, and remote conditioning. *Circ Res* 119:676–695. doi:10.1161/CIRCRESA-HA.116.308736
  79. Hausenloy DJ, Lecour S, Yellon DM (2011) Reperfusion injury salvage kinase and survivor activating factor enhancement prosurvival signaling pathways in ischemic postconditioning: two sides of the same coin. *Antioxid Redox Signal* 14:893–907. doi:10.1089/ars.2010.3360
  80. Hausenloy DJ, Yellon DM (2004) New directions for protecting the heart against ischaemia-reperfusion injury: targeting the Reperfusion Injury Salvage Kinase (RISK)-pathway. *Cardiovasc Res* 61:448–460. doi:10.1016/j.cardiores.2003.09.024
  81. Hausenloy DJ, Yellon DM (2007) Reperfusion injury salvage kinase signalling: taking a RISK for cardioprotection. *Heart Fail Rev* 12:217–234. doi:10.1007/s10741-007-9026-1
  82. Lecour S (2009) Activation of the protective Survivor Activating Factor Enhancement (SAFE) pathway against reperfusion injury: does it go beyond the RISK pathway? *J Mol Cell Cardiol* 47:32–40. doi:10.1016/j.yjmcc.2009.03.019
  83. Lecour S (2009) Multiple protective pathways against reperfusion injury: a SAFE path without Aktion? *J Mol Cell Cardiol* 46:607–609. doi:10.1016/j.yjmcc.2009.01.003
  84. Schulman D, Latchman DS, Yellon DM (2002) Urocortin protects the heart from reperfusion injury via upregulation of p42/ p44 MAPK signaling pathway. *Am J Physiol Heart Circ Physiol* 283:H1481–H1488. doi:10.1152/ajpheart.01089.2001
  85. Lecour S, Smith RM, Woodward B, et al. (2002) Identification of a novel role for sphingolipid signaling in TNF alpha and ischemic preconditioning mediated cardioprotection. *J Mol Cell Cardiol* 34:509–518. doi:10.1006/jmcc.2002.1533
  86. Lecour S, Suleman N, Deuchar GA, (2005) Pharmacological preconditioning with tumor necrosis factor-alpha activates signal transducer and activator of transcription-3 at reperfusion without involving classic prosurvival kinases (Akt and extracellular signal-regulated kinase). *Circulation* 112:3911–3918. doi:10.1161/CIRCULATIONAHA.105.581058
  87. Frias MA, Lecour S, James RW, (2012) High density lipoprotein/sphingosine-1-phosphate-induced cardioprotection: role of STAT3 as part of the SAFE pat-

- hway. JAKSTAT 1:92–100. doi:10.4161/jkst.19754
88. Ludman AJ, Hausenloy DJ, Babu G, et al. (2011) Failure to recapture cardioprotection with high-dose atorvastatin in coronary artery bypass surgery: a randomised controlled trial. *Basic Res Cardiol* 106:1387–1395. doi:10.1007/s00395-011-0209-5
  89. Ludman AJ, Yellon DM, Hasleton J, et al. (2011) Effect of erythropoietin as an adjunct to primary percutaneous coronary intervention: a randomised controlled clinical trial. *Heart* 97:1560–1565. doi:10.1136/hrt.2011.223867
  90. Alburquerque-Bejar JJ, Barba I, Inserte J, et al. (2015) Combination therapy with remote ischaemic conditioning and insulin or exenatide enhances infarct size limitation in pigs. *Cardiovasc Res* 107:246–254. doi:10.1093/cvr/cvv171
  91. Lei P, Bai T, Sun Y (2018) Mechanisms of Ferroptosis and Relations with Regulated Cell Death: A Review. *Front. Physiol.* 10:139. doi:10.3389/fphys.2019.00139
  92. Kezic A, Spasojevic I, Lezaic V et al. (2016) Mitochondria-Targeted Antioxidants: Future Perspectives in Kidney Ischemia Reperfusion Injury; A Review. *Oxidative Medicine and Cellular Longevity* Volume 2016, Article ID 2950503, 1-12 <http://dx.doi.org/10.1155/2016/2950503>
  93. Tyagi S, Tya S, Singh N Diabetes abolish cardioprotective effects of remote ischemic conditioning: evidences and possible mechanisms. *J Physiol Biochem.* 2019 Feb;75(1):19–28. doi: 10.1007/s13105-019-00664-w. Epub 2019 Feb 7
  94. Reiter RJ. Oxidative processes and antioxidant defense mechanisms in the aging brain. *FASEB J* 1995; 9: 526-533.
  95. Karihtala P, Soini Y. Reactive oxygen species and antioxidant mechanisms in human tissues and their relation to malignancies. *APMIS* 2007;115:81-103.
  96. Cherubini A, Ruggiero C, Morand C, et al. 2008; 15: 1236-1248. Mickle DA, Weisel RD. Future directions of vitamin E and its analogues in minimizing myocardial ischemia-reperfusion injury. *Can J Cardiol* 1993; 9: 89-93.
  97. Mickle DA, Weisel RD. Future directions of vitamin E and its analogues in minimizing myocardial ischemia-reperfusion injury. *Can J Cardiol* 1993; 9: 89-93.
  98. Virág L, Szabó C. The therapeutic potential of poly(ADP-ribose) polymerase inhibitors. *Pharmacol Rev* 2002; 54: 375-429
  99. Lasheen N., Elaylat W., Elrefai M. et al. Possible role of garlic oil in ameliorating renal injury after liver ischemia/reperfusion in rats. *J Physiol Pharmacol.* 2019 Oct;70(5). doi:10.26402/jpp.2019.5.12. Epub 2020 Jan 30. PMID: 32009629
  100. Tian F, Liu R, Fan C. Et al. Effects of Thymoquinone on Small-Molecule Metabolites in a Rat Model of Cerebral Ischemia Reperfusion Injury Assessed using MALDI-MSI. *Metabolites.* 2020 Jan 7;10(1). pii: E27. doi: 10.3390/metabo10010027. PMID:31936061
  101. Sener G, Sehirli AO, Keyer-Uysal M, Arbak S, Ersoy Y, Yeğen BC. The protective effect of melatonin on renal ischemia-reperfusion injury in the rat. *J Pineal Res* 2002; 32: 120-126.
  102. Shi S, Lei S, Tang C. Et al. Melatonin attenuates acute kidney ischemia/reperfusion injury in diabetic rats by activation of the SIRT1/Nrf2/HO-1 signaling pathway. *Biosci Rep.* 2019 Jan 15;39(1). pii: BSR20181614. doi: 10.1042/BSR20181614. Print 2019 Jan 31. PMID: 30578379
  103. Banaei S. Novel role of microRNAs in renal ischemia reperfusion injury. *Ren Fail.* 2015 Aug;37(7):1073-9. doi: 10.3109/0886022X.2015.1055697. Epub 2015 Jun 9. Review. PMID: 26056732
  104. Sener G, Tosun O, Sehirli AO, Kaçmaz A, Arbak S, Ersoy Y, Ayanoğlu-Dülger G. Melatonin and N-acetylcysteine have beneficial effects during hepatic ischemia and reperfusion. *Life Sci* 2003; 72: 2707-2718
  105. Sener G, Kaçmaz A, User Y, Ozkan S, Tilki M, Yeğen BC. Melatonin ameliorates oxidative organ damage induced by acute intra-abdominal compartment syndrome in rats. *J Pineal Res* 2003 ;35: 163-168.
  106. Erkanli K, Kayalar N, Erkanli G, Ercan F, Sener G, Kirali K. Melatonin protects against ischemia/reperfusion injury in skeletal muscle. *J Pineal Res* 2005; 39: 238-242
  107. Kaçmaz A, User EY, Sehirli AO, Tilki M, Ozkan S, Sener G. Protective effect of melatonin against ischemia/reperfusion-induced oxidative remote organ injury in the rat. *Surg Today.* 2005; 35:744-750.
  108. Sener G, Sert G, Ozer Sehirli A, Arbak S, Gedik N, Ayanoğlu-Dülger G. Melatonin protects against pressure ulcer-induced oxidative injury of the skin and remote organs in rats. *J Pineal Res* 2006; 40:280-287.
  109. Sener G, Tuğtepe H, Yüksel M, Cetinel S, Gedik N, Yeğen BC. Resveratrol improves ischemia/reperfusion-induced oxidative renal injury in rats. *Arch Med Res* 2006; 37: 822-829.
  110. Toklu H, Alican I, Ercan F, et al. The beneficial effect of resveratrol on rat bladder contractility and oxidant damage following ischemia/reperfusion. *Pharmacology* 2006; 78:44-50.
  111. Dulundu E, Ozel Y, Topaloglu U, et al. Alpha-lipoic acid protects against hepatic ischemia-reperfusion injury in rats. *Pharmacology.* 2007; 79:163-170.
  112. Sehirli O, Sener E, Cetinel S, et al. Alpha-lipoic acid protects against renal ischaemia-reperfusion injury in rats. *Clin Exp Pharmacol Physiol* 2008; 35:249-255
  113. Chen Z., Ding T., Ma C. Dexmedetomidine (DEX) protects against hepatic ischemia/reperfusion (I/R) injury by suppressing inflammation and oxidative stress in NLRC5 deficient mice. *Biochem Biophys Res Commun.* 2017 Nov 18;493(2):1143-1150. doi: 10.1016/j.bbrc.2017.08.017. Epub 2017 Aug 4.
  114. Lauz Medeiros SH, de Oliveira Menezes A, Zogbi L, et al. N-Acetylcysteine Use in Hepatic Ischemia/Reperfusion in Rats Minimizing Bowel Injury. *Transplant Proc.* 2016 Sep;48(7):2371-2374. doi: 10.1016/j.transproceed.2016.06.003.
  115. Sehirli A, Sener G, Satiroglu H, et al. Protective effect of N-acetylcysteine on renal ischemia/reperfusion injury in the rat. *J Nephrol* 2003; 16: 75-80.
  116. Sehirli O, Ozel Y, Dulundu E, et al. Grape seed extract treatment reduces hepatic ischemia-reperfusion injury in rats. *Phytother Res* 2008; 22: 43-48

117. Amini N, Sarkaki A. , Dianat M. Et al. Protective effects of naringin and trimetazidine on remote effect of acute renal injury on oxidative stress and myocardial injury through Nrf-2 regulation. *Pharmacol Rep.* 2019 Nov;71(6):1059-1066. doi: 10.1016/j.pharep.2019.06.007. Epub 2019 Jun 14
118. Sener G, Sehirli O, Velioglu A., Montelukast protects against renal ischemia/ reperfusion injury in rats. *Pharmacol Res.* 2006; 54: 65-71.
119. Sener G, Sener E, Sehirli O, Oğünç AV, Cetinel S, Gedik N, Sakarcan A. Ginkgo biloba extract ameliorates ischemia reperfusion-induced renal injury in rats. *Pharmacol Res* 2005; 52: 216-22.
120. Kabasakal L, Sehirli O, Cetinel S, et al. Protective effect of aqueous garlic extract against renal ischemia/reperfusion injury in rats. *J Med Food.* 2005; 8:319-326.
121. Sener G, Sehirli O, Ipçi Y, et al. Aqueous garlic extract alleviates ischaemia-reperfusion-induced oxidative hepatic injury in rats. *J Pharm Pharmacol.* 2005; 57: 145-150
122. Sener G, Sert G, Ozer Sehirli A, Arbak S, Uslu B, Gedik N, Ayanoglu-Dulger G. Pressure ulcer-induced oxidative organ injury is ameliorated by beta-glucan treatment in rats. *Int Immunopharmacol.* 2006; 6: 724-732.
123. Kaçmaz A, Polat A, User Y, Tilki M, Ozkan S, Sener G. Octreotide: a new approach to the management of acute abdominal hypertension. *Peptides.* 2003; 24: 1381-1386.
124. Sener G, Sehirli O, Ercan F, Sirvanci S, Gedik N, Kaçmaz A. Protective effect of MESNA (2-mercaptoethane sulfonate) against hepatic ischemia/reperfusion injury in rats. *Surg Today.* 2005; 35:575-580.
125. Sener G, Akgün U, Satiroğlu H, Topaloğlu U, Keyer-Uysal M. The effect of pentoxifylline on intestinal ischemia/reperfusion injury. *Fundam Clin Pharmacol.* 2001; 15: 19-22