

**ALZHEIMER HASTALIĞINDA NHE6:
ENDOZOMAL PH DÜZENLENMESİ VE
MOLEKÜLER MEKANİZMALAR**

İrem Gülfem AKIN

Üsküdar Üniversitesi



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*Bu kitap yazara ait "1013327" referans numaralı, "NHE6 Gen Modülasyonunun Alzheimer ile İlişkili Genlerin Anlatımı Üzerine Etkisinin Araştırılması" başlıklı doktora tezinden üretilmiştir.

ISBN **Sayfa ve Kapak Tasarımı**
978-625-362-148-3 Akademisyen Dizgi Ünitesi

Kitap Adı **Yayıncı Sertifika No**
Alzheimer Hastalığında NHE6: 47518
Endozomal pH Düzenlenmesi ve
Baskı ve Cilt
Moleküler Mekanizmalar Vadi Matbaacılık

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Yazar **DOI**
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Kütüphane Kimlik Kartı
Akın, İrem Gülfem.
Alzheimer Hastalığında NHE6: Endozomal pH Düzenlenmesi ve Moleküler
Mekanizmalar / İrem Gülfem Akın; ed. Muhsin Konuk.
Ankara : Akademisyen Yayınevi Kitabevi, 2026.
91 s. ; 135x210 mm.
Kaynakça var.
ISBN 9786253621483

GENEL DAĞITIM
Akademisyen Yayınevi

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Yenişehir / Ankara
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ÖNSÖZ

Alzheimer hastalığı, genetik yatkınlık, hücre içi protein trafiği, endozomal-lizozomal işlevler ve nörodejeneratif süreçlerin birbiriyle etkileşimi sonucunda gelişen kompleks bir hastalıktır. Son yıllarda yapılan çalışmalar, endozomal pH dengesinin ve bu dengenin düzenlenmesinde görev alan proteinlerin Alzheimer hastalığının moleküler mekanizmalarının anlaşılmasında önemli bir yere sahip olduğunu göstermektedir.

Bu kitap, “NHE6 Gen Modülasyonunun Alzheimer ile İlişkili Genlerin Anlatımı Üzerine Etkisinin Araştırılması” başlıklı doktora tez çalışmam temel alınarak hazırlanmıştır. Kitapta Alzheimer hastalığının temel klinik, patolojik ve genetik özelliklerinin yanı sıra endolizozomal sistem, hücre içi pH düzenlenmesi ve *SLC9A6* geni tarafından kodlanan NHE6 proteininin hücresel işlevleri ele alınmıştır. Ayrıca NHE6’nın Christianson sendromu, epilepsi, Parkinson hastalığı ve Alzheimer hastalığı ile ilişkisi değerlendirilmiş; NHE6 ve endozomal pH düzenlenmesinin terapötik hedef olarak taşıdığı potansiyel, mevcut literatür bulguları çerçevesinde tartışılmıştır.

Doktora eğitimim ve bu çalışmanın hazırlanması sürecinde bilgi ve deneyimleriyle bana yol gösteren saygıdeğer danışmanım Prof. Dr. Muhsin Konuk’a teşekkürlerimi sunarım. Ayrıca akademik çalışmalarım boyunca desteklerini esirgemeyen sevgili eşime ve aileme içtenlikle teşekkür ederim.

Dr. İrem Gülfem Akın

KISALTMALAR

Kısaltmalar	Açıklama
A β	: Amiloid-beta
AEP	: Asparaginil endopeptidaz
AH	: Alzheimer hastalığı
AMPA	: α -Amino-3-hidroksi-5-metil-4 izoksazolpropiyonik asit
AMPAR	: AMPA reseptörü
AP2	: Adaptör protein kompleksi 2
ApoE	: Apolipoprotein E
ApoE4	: Apolipoprotein E4
APP	: Amiloid öncü protein
AT2	: Anjiyotensin II tip 2 reseptörü
BACE1	: Beta-bölgesi APP parçalayıcı enzim 1
BDNF	: Beyin kaynaklı nörotrofik faktör
BOS	: Beyin omurilik sıvısı
CKI ϵ	: Kazein kinaz 1 epsilon
CME	: Klatrin aracılı endositoz
<i>CSNK1E</i>	: Kazein kinaz 1 epsilon geni
DNA	: Deoksiribonükleik asit
EE	: Erken endozom
eNHE	: Endozomal Na ⁺ /H ⁺ değiştirici
FDA	: Amerika Birleşik Devletleri Gıda ve İlaç Dairesi
GABA	: Gama-aminobütirik asit
GDP	: Guanozin difosfat
GTP	: Guanozin-5'-trifosfat
HDAC	: Histon deasetilaz
HDAC4	: Histon deasetilaz 4
HEK293	: İnsan embriyonik böbrek 293 hücreleri
IGF1	: İnsülin benzeri büyüme faktörü 1
IRS	: İnsülin reseptörü substratı
<i>IRS1</i>	: İnsülin reseptörü substrat 1 geni

KEGG	: “ <i>Kyoto Encyclopedia of Genes and Genomes</i> ”
LE	: Geç endozom
LTP	: Uzun süreli potansiyasyon
MRG	: Manyetik rezonans görüntüleme
MSS	: Merkezi sinir sistemi
NFT	: Nörofibriler yumak
NHE	: Na^+/H^+ değiştirici
NHE6	: Na^+/H^+ değiştirici 6
NMDAR	: N-metil-D-aspartat reseptörü
PET	: Pozitron emisyon tomografisi
PH	: Parkinson hastalığı
PHF	: Eşleşmiş sarmal filament
pHi	: Hücre içi pH
pHo	: Hücre dışı pH
<i>PSEN1</i>	: Presenilin 1 geni
<i>PSEN2</i>	: Presenilin 2 geni
PSD95	: Postsinaptik yoğunluk proteini 95
pTrkB	: Fosforile TrkB
RACK1	: Aktive protein kinaz C reseptörü 1
RE	: Geri dönüşüm endozomu
RNA	: Ribonükleik asit
SE	: Sinyal endozomları
SLC9A6	: “ <i>SLC9A6 solute carrier family 9 member A6</i> ” geni
SNARE	: Çözünür NSF bağlanma proteini reseptörü
SNP	: Tek nükleotid polimorfizmi
SV2	: Sinaptik vezikül proteini 2
TM	: Transmembran
TrkB	: Tropomiyozin reseptör kinaz B
V-ATPaz	: Vakuoler tip H^+ -ATPaz
β CTF	: Karboksil-terminal APP parçacığı

İÇİNDEKİLER

GİRİŞ	1
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BÖLÜM 1

ALZHEIMER HASTALIĞINA GENEL BAKIŞ	5
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1. 1. Alzheimer Hastalığı	5
1.2. Alzheimer Hastalığının Patofizyolojisi	9
1.2.1. Senil plaklar	9
1.2.2. Nörofibriler yumaklar	10
1.2.3. Sinaptik kayıplar	10
1.2.4. Oksidatif stres	11
1.3. Alzheimer Hastalığının Genetik Temeli	12

BÖLÜM 2

ENDOLİZOZOMAL SİSTEM	15
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2.1. Endolizozomal Sistem ve Hücre İçi pH Düzenlemesi	15
2.2. Na^+/H^+ Değiştiriciler	22
2.3. NHE6 Proteini ve Hücrel Fonksiyonları	23

BÖLÜM 3

ALZHEIMER HASTALIĞI İLE İLİŞKİLİ BAZI GENLER VE MOLEKÜLER SİNYAL YOLAKLARI	31
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3.1. <i>APP</i> Geni	31
3.2. <i>PSEN1</i> Geni	33
3.3. <i>IRS1</i> Geni	35
3.4. <i>CK1ε</i> Geni	36

BÖLÜM 4**NHE6 VE NÖROLOJİK HASTALIKLAR39**

4.1. Christianson Sendromu 39

4.2. NHE6 ve Epilepsi 40

4.3. NHE6 ve Parkinson Hastalığı 41

4.4. NHE6 ve Alzheimer Hastalığı 42

BÖLÜM 5**NHE6’NIN ALZHEIMER HASTALIĞI İÇİN TERAPÖTİK HEDEF OLARAK DEĞERLENDİRİLMESİ45**

5.1. Endozomal pH’ın Terapötik Hedef Olarak Önemi 46

5.2. *SLC9A6* Gen Anlatımının APP Trafikğine Etkisi 47

5.3. NHE6 ve APP-BACE1 Etkileşimi 49

5.4. NHE6 Modülasyonunun A β Üretimine Etkisi 50

5.5. Endozomal Alkalileştirici Ajanlar 54

5.6. NHE6’nın Farmakolojik Olarak Hedeflenebilirliği 55

5.7. Terapötik Yaklaşımın Sınırlılıkları 57

BÖLÜM 6**TARTIŞMA VE GENEL DEĞERLENDİRME61****BÖLÜM 7****GELECEK PERSPEKTİFLERİ65****KAYNAKÇA69**

KAYNAKÇA

1. Rostagno, A. A. Pathogenesis of Alzheimer's Disease. *Int. J. Mol. Sci.* **24**, (2023).
2. Orlowski, J. & Grinstein, S. Na⁺/H⁺ exchangers. *Compr. Physiol.* **1**, 2083–2100 (2011).
3. Pedersen, S. F. & Counillon, L. The SLC9A-C Mammalian Na⁺/H⁺ Exchanger Family: Molecules, Mechanisms, and Physiology. *Physiol. Rev.* **99**, 2015–2113 (2019).
4. Casey, J. R., Grinstein, S. & Orlowski, J. Sensors and regulators of intracellular pH. *Nat. Rev. Mol. Cell Biol.* **11**, 50–61 (2010).
5. Chesler, M. Regulation and modulation of pH in the brain. *Physiol. Rev.* **83**, 1183–1221 (2003).
6. Sinning, A. & Hübner, C. A. Minireview: pH and synaptic transmission. *FEBS Lett.* **587**, 1923–1928 (2013).
7. Li, P. A. & Siesjö, B. K. Role of hyperglycaemia-related acidosis in ischaemic brain damage. *Acta Physiol. Scand.* **161**, 567–580 (1997).
8. Schuchmann, S. *et al.* Experimental febrile seizures are precipitated by a hyperthermia-induced respiratory alkalosis. *Nat. Med.* **12**, 817–823 (2006).
9. Patak, J., Faraone, S. V. & Zhang-James, Y. Sodium hydrogen exchanger 9 NHE9 (SLC9A9) and its emerging roles in neuropsychiatric comorbidity. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **183**, 289–305 (2020).
10. Nakamura, N., Tanaka, S., Teko, Y., Mitsui, K. & Kanazawa, H. Four Na⁺/H⁺ exchanger isoforms are distributed to Golgi and post-Golgi compartments and are involved in organelle pH regulation. *J. Biol. Chem.* **280**, 1561–1572 (2005).
11. Ohgaki, R. *et al.* The Na⁺/H⁺ exchanger NHE6 in the endosomal recycling system is involved in the development of apical bile canalicular surface domains in HepG2 cells. *Mol. Biol. Cell* **21**, 1293–1304 (2010).
12. Ouyang, Q. *et al.* Christianson syndrome protein NHE6 modulates TrkB endosomal signaling required for neuronal circuit development. *Neuron* **80**, 97–112 (2013).
13. Roxrud, I., Raiborg, C., Gilfillan, G. D., Strømme, P. & Stenmark, H. Dual degradation mechanisms ensure disposal of NHE6 mutant protein associated with neurological disease. *Exp. Cell Res.* **315**, 3014–3027 (2009).
14. Malo, M. E. & Fliegel, L. Physiological role and regulation of the Na⁺/H⁺ exchanger. *Can. J. Physiol. Pharmacol.* **84**, 1081–1095 (2006).
15. Kittler, J. T. *et al.* Constitutive endocytosis of GABAA receptors by an association with the adaptin AP2 complex modulates inhibitory synaptic currents in hippocampal neurons. *J. Neurosci.* **20**, 7972–7977 (2000).
16. Hanley, J. G. The Regulation of AMPA Receptor Endocytosis by Dynamic Protein-Protein Interactions. *Front. Cell. Neurosci.* **12**, (2018).

17. Kurosinski, P., Guggisberg, M. & Götz, J. Alzheimer's and Parkinson's disease - Overlapping or synergistic pathologies? *Trends Mol. Med.* **8**, 3–5 (2002).
18. Mignot, C. *et al.* Novel mutation in SLC9A6 gene in a patient with Christ-
ianson syndrome and retinitis pigmentosa. *Brain Dev.* **35**, 172–176 (2013).
19. Pescosolido, M. F. *et al.* Complex Neurological Phenotype in Female Car-
riers of NHE6 Mutations. *Mol. Neuropsychiatry* **5**, 98–108 (2019).
20. Jiang, T., Yu, J.-T., Tian, Y. & Tan, L. Epidemiology and etiology of Alzhei-
mer's disease: from genetic to non-genetic factors. *Curr. Alzheimer Res.* **10**,
852–867 (2013).
21. Ross, C. A. & Poirier, M. A. Protein aggregation and neurodegenerative di-
sease. *Nat. Med.* **10 Suppl**, S10 (2004).
22. Bonifacino, J. S. & Traub, L. M. Signals for sorting of transmembrane prote-
ins to endosomes and lysosomes. *Annu. Rev. Biochem.* **72**, 395–447 (2003).
23. Haug Yu, W. *et al.* Macroautophagy--a novel Beta-amyloid peptide-gene-
rating pathway activated in Alzheimer's disease. *J. Cell Biol.* **171**, 87–98
(2005).
24. Cirrito, J. R. *et al.* Endocytosis is required for synaptic activity-dependent
release of amyloid-beta in vivo. *Neuron* **58**, 42–51 (2008).
25. Sun, J. & Roy, S. The physical approximation of APP and BACE-1: A key
event in alzheimer's disease pathogenesis. *Dev. Neurobiol.* **78**, 340–347
(2018).
26. Strømme, P. *et al.* X-linked Angelman-like syndrome caused by Slc9a6 kno-
ckout in mice exhibits evidence of endosomal-lysosomal dysfunction. *Brain*
134, 3369–3383 (2011).
27. Lee, Y. *et al.* Early lysosome defects precede neurodegeneration with amy-
loid- β and tau aggregation in NHE6-null rat brain. *Brain* **145**, 3187–3202
(2022).
28. Prasad, H. & Rao, R. The Na⁺/H⁺ exchanger NHE6 modulates endosomal
pH to control processing of amyloid precursor protein in a cell culture model
of Alzheimer disease. *J. Biol. Chem.* **290**, 5311–5327 (2015).
29. Prasad, H. & Rao, R. Amyloid clearance defect in ApoE4 astrocytes is re-
versed by epigenetic correction of endosomal pH. *Proc. Natl. Acad. Sci. U.*
S. A. **115**, E6640–E6649 (2018).
30. Fernandez, M. A. *et al.* Loss of endosomal exchanger NHE6 leads to patho-
logical changes in tau in human neurons. *Stem Cell Reports* **17**, 2111–2126
(2022).
31. De-Paula, V. J., Radanovic, M., Diniz, B. S. & Forlenza, O. V. Alzheimer's
Disease. *Subcell. Biochem.* **65**, 329–352 (2012).
32. Ferretti, M. T. *et al.* Sex differences in Alzheimer disease — the gateway
to precision medicine. *Nature Reviews Neurology* 2018 14:8 **14**, 457–469
(2018).
33. Livingston, G. *et al.* Dementia prevention, intervention, and care: 2020 re-
port of the Lancet Commission. *The Lancet* **396**, 413–446 (2020).

34. Yiannopoulou, K. G. & Papageorgiou, S. G. Current and Future Treatments in Alzheimer Disease: An Update. *J. Cent. Nerv. Syst. Dis.* **12**, (2020).
35. Haines, J. L. Alzheimer Disease: Perspectives from Epidemiology and Genetics. *Journal of Law, Medicine & Ethics* **46**, 694–698 (2018).
36. 2019 Alzheimer's disease facts and figures. *Alzheimer's & Dementia* **15**, 321–387 (2019).
37. Mucke, L. Alzheimer's disease. *Nature* 2009 461:7266 **461**, 895–897 (2009).
38. Piaceri, I., Nacmias, B. & Sorbi, S. Genetics of familial and sporadic Alzheimer's disease. *Frontiers in Bioscience - Elite* **5 E**, 167–177 (2013).
39. Tiwari, S., Atluri, V., Kaushik, A., Yndart, A. & Nair, M. Alzheimer's disease: pathogenesis, diagnostics, and therapeutics. *Int. J. Nanomedicine* **14**, 5541–5554 (2019).
40. Lanoiselée, H. M. *et al.* APP, PSEN1, and PSEN2 mutations in early-onset Alzheimer disease: A genetic screening study of familial and sporadic cases. *PLoS Med.* **14**, e1002270 (2017).
41. Gunnarsson, L. G. & Bodin, L. Occupational Exposures and Neurodegenerative Diseases—A Systematic Literature Review and Meta-Analyses. *International Journal of Environmental Research and Public Health* 2019, Vol. 16, Page 337 **16**, 337 (2019).
42. Huynh, T. P. V., Davis, A. A., Ulrich, J. D. & Holtzman, D. M. Apolipoprotein E and Alzheimer's disease: the influence of apolipoprotein E on amyloid- β and other amyloidogenic proteins: Thematic Review Series: ApoE and Lipid Homeostasis in Alzheimer's Disease. *J. Lipid Res.* **58**, 824–836 (2017).
43. Najm, R., Jones, E. A. & Huang, Y. Apolipoprotein E4, inhibitory network dysfunction, and Alzheimer's disease. *Molecular Neurodegeneration* 2019 14:1 **14**, 1–13 (2019).
44. Gordon, B. A. *et al.* Spatial patterns of neuroimaging biomarker change in individuals from families with autosomal dominant Alzheimer's disease: a longitudinal study. *Lancet Neurol.* **17**, 241–250 (2018).
45. Kwok, J. B. Role of epigenetics in Alzheimer's and Parkinson's disease. *Epigenomics* **2**, 671–682 (2010).
46. Qazi, T. J., Quan, Z., Mir, A. & Qing, H. Epigenetics in Alzheimer's Disease: Perspective of DNA Methylation. *Molecular Neurobiology* 2017 55:2 **55**, 1026–1044 (2017).
47. Mastroeni, D. *et al.* Epigenetic changes in Alzheimer's disease: Decrements in DNA methylation. *Neurobiol. Aging* **31**, 2025–2037 (2010).
48. Monteiro, A. R., Barbosa, D. J., Remião, F. & Silva, R. Alzheimer's disease: Insights and new prospects in disease pathophysiology, biomarkers and disease-modifying drugs. *Biochem. Pharmacol.* **211**, 115522 (2023).
49. Yokoyama, A. S., Rutledge, J. C. & Medici, V. DNA methylation alterations in Alzheimer's disease. *Environ. Epigenet.* **3**, (2017).

50. Chouliaras, L. *et al.* Consistent decrease in global DNA methylation and hydroxymethylation in the hippocampus of Alzheimer's disease patients. *Neurobiol. Aging* **34**, 2091–2099 (2013).
51. Bradley-Whitman, M. A. & Lovell, M. A. Epigenetic changes in the progression of Alzheimer's disease. *Mech. Ageing Dev.* **134**, 486–495 (2013).
52. Serrano-Pozo, A., Frosch, M. P., Masliah, E. & Hyman, B. T. Neuropathological Alterations in Alzheimer Disease. *Cold Spring Harb. Perspect. Med.* **1**, a006189 (2011).
53. Singh, S. K., Srivastav, S., Yadav, A. K., Srikrishna, S. & Perry, G. Overview of Alzheimer's Disease and Some Therapeutic Approaches Targeting A β by Using Several Synthetic and Herbal Compounds. *Oxid. Med. Cell. Longev.* **2016**, 7361613 (2016).
54. Spire-Jones, T. L. & Hyman, B. T. The Intersection of Amyloid Beta and Tau at Synapses in Alzheimer's Disease. *Neuron* **82**, 756–771 (2014).
55. Cras, P. *et al.* Senile plaque neurites in Alzheimer disease accumulate amyloid precursor protein. *Proceedings of the National Academy of Sciences* **88**, 7552–7556 (1991).
56. Perl, D. P. Neuropathology of Alzheimer's Disease. *Mount Sinai Journal of Medicine: A Journal of Translational and Personalized Medicine* **77**, 32–42 (2010).
57. Armstrong, R. The molecular biology of senile plaques and neurofibrillary tangles in Alzheimer's disease. *Folia Neuropathol.* (2009).
58. Tabaton, M. & Piccini, A. Role of water-soluble amyloid- β in the pathogenesis of Alzheimer's disease. *Int. J. Exp. Pathol.* **86**, 139–145 (2005).
59. Brion, J.-P. & Brion, J.-P. Neurofibrillary Tangles and Alzheimer's Disease. *Eur. Neurol.* **40**, 130–140 (1998).
60. Metaxas, A. & Kempf, S. J. Neurofibrillary tangles in Alzheimer's disease: Elucidation of the molecular mechanism by immunohistochemistry and tau protein phospho-proteomics. *Neural Regen. Res.* **11**, 1579–1581 (2016).
61. Overk, C. R. & Masliah, E. Pathogenesis of synaptic degeneration in Alzheimer's disease and Lewy body disease. *Biochem. Pharmacol.* **88**, 508–516 (2014).
62. Lleó, A. *et al.* Changes in Synaptic Proteins Precede Neurodegeneration Markers in Preclinical Alzheimer's Disease Cerebrospinal Fluid. *Molecular & Cellular Proteomics* **18**, 546–560 (2019).
63. Tarawneh, R. *et al.* Diagnostic and Prognostic Utility of the Synaptic Marker Neurogranin in Alzheimer Disease. *JAMA Neurol.* **73**, 561–571 (2016).
64. Ito, F., Sono, Y. & Ito, T. Measurement and Clinical Significance of Lipid Peroxidation as a Biomarker of Oxidative Stress: Oxidative Stress in Diabetes, Atherosclerosis, and Chronic Inflammation. *Antioxidants* **2019**, Vol. 8, Page 72 **8**, 72 (2019).
65. Forman, H. J. & Zhang, H. Targeting oxidative stress in disease: promise and limitations of antioxidant therapy. *Nature Reviews Drug Discovery* **2021** **20**, 689–709 (2021).

66. Savelieff, M. G. *et al.* Development of multifunctional molecules as potential therapeutic candidates for Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis in the last decade. *Chem. Rev.* **119**, 1221–1322 (2019).
67. Circu, M. L. & Aw, T. Y. Reactive oxygen species, cellular redox systems, and apoptosis. *Free Radic. Biol. Med.* **48**, 749–762 (2010).
68. Van Cauwenberghe, C., Van Broeckhoven, C. & Sleegers, K. The genetic landscape of Alzheimer disease: clinical implications and perspectives. *Genetics in Medicine* **18**, 421–430 (2016).
69. Khanahmadi, M., Farhud, D. & Malmir, M. Genetic of Alzheimer's Disease: A Narrative Review Article. *Iran. J. Public Health* (2015).
70. Li, N. M. *et al.* Mutations of beta-amyloid precursor protein alter the consequence of Alzheimer's disease pathogenesis. *Neural Regen. Res.* **14**, 658–665 (2019).
71. Tew, J. & Goate, A. M. Genetics of β -Amyloid Precursor Protein in Alzheimer's Disease. *Cold Spring Harb. Perspect. Med.* **7**, a024539 (2017).
72. Cai, Y., An, S. S. A. & Kim, S. Mutations in presenilin 2 and its implications in Alzheimer's disease and other dementia-associated disorders. *Clin. Interv. Aging* **10**, 1163–1172 (2015).
73. Dai, M.-H. *et al.* The genes associated with early-onset Alzheimer's disease. *Oncotarget* **9**, 15132–15143 (2018).
74. Kelleher, R. J. & Shen, J. Presenilin-1 mutations and Alzheimer's disease. *Proceedings of the National Academy of Sciences* **114**, 629–631 (2017).
75. Walker, E. S., Martinez, M., Brunkan, A. L. & Goate, A. Presenilin 2 familial Alzheimer's disease mutations result in partial loss of function and dramatic changes in A β 42/40 ratios. *J. Neurochem.* **92**, 294–301 (2005).
76. Kim, J., Basak, J. M. & Holtzman, D. M. The Role of Apolipoprotein E in Alzheimer's Disease. *Neuron* **63**, 287–303 (2009).
77. Liu, C. C., Kanekiyo, T., Xu, H. & Bu, G. Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nature Reviews Neurology* **2013** 9:2 **9**, 106–118 (2013).
78. Giau, V. Van, Bagyinszky, E., An, S. S. A. & Kim, S. Y. Role of apolipoprotein E in neurodegenerative diseases. *Neuropsychiatr. Dis. Treat.* **11**, 1723–1737 (2015).
79. Hu, Y. B., Dammer, E. B., Ren, R. J. & Wang, G. The endosomal-lysosomal system: from acidification and cargo sorting to neurodegeneration. *Transl. Neurodegener.* **4**, (2015).
80. Neefjes, J. & van der Kant, R. Stuck in traffic: an emerging theme in diseases of the nervous system. *Trends Neurosci.* **37**, 66–76 (2014).
81. Tan, J. & Evin, G. B-site APP-cleaving enzyme 1 trafficking and Alzheimer's disease pathogenesis. *J. Neurochem.* **120**, 869–880 (2012).
82. Wu, J. *et al.* Arc/Arg3.1 Regulates an Endosomal Pathway Essential for Activity-Dependent β -Amyloid Generation. *Cell* **147**, 615 (2011).

83. Lamb, C. A., Dooley, H. C. & Tooze, S. A. Endocytosis and autophagy: Shared machinery for degradation. *BioEssays* **35**, 34–45 (2013).
84. Tooze, S. A., Abada, A. & Elazar, Z. Endocytosis and Autophagy: Exploitation or Cooperation? *Cold Spring Harb. Perspect. Biol.* **6**, (2014).
85. Huotari, J. & Helenius, A. Endosome maturation. *EMBO J.* **30**, 3481–3500 (2011).
86. Diering, G. H. & Numata, M. Endosomal pH in neuronal signaling and synaptic transmission: Role of Na⁺/H⁺ exchanger NHE5. *Front. Physiol.* **4 JAN**, 70666 (2014).
87. Verkhovskii, R. A. *et al.* Current Principles, Challenges, and New Metrics in pH-Responsive Drug Delivery Systems for Systemic Cancer Therapy. *Pharmaceutics* **15**, (2023).
88. Villarroel-Campos, D., Gastaldi, L., Conde, C., Caceres, A. & Gonzalez-Billault, C. Rab-mediated trafficking role in neurite formation. *J. Neurochem.* **129**, 240–248 (2014).
89. Laifenfeld, D. *et al.* Rab5 Mediates an Amyloid Precursor Protein Signaling Pathway That Leads to Apoptosis. *The Journal of Neuroscience* **27**, 7141 (2007).
90. Hsu, V. W. & Prekeris, R. Transport at the recycling endosome. *Curr. Opin. Cell Biol.* **22**, 528–534 (2010).
91. Li, X. & DiFiglia, M. The recycling endosome and its role in neurological disorders. *Prog. Neurobiol.* **97**, 127–141 (2012).
92. Ulery, P. G. *et al.* Modulation of beta-amyloid precursor protein processing by the low density lipoprotein receptor-related protein (LRP). Evidence that LRP contributes to the pathogenesis of Alzheimer's disease. *J. Biol. Chem.* **275**, 7410–7415 (2000).
93. Kang, D. E. *et al.* Modulation of amyloid beta-protein clearance and Alzheimer's disease susceptibility by the LDL receptor-related protein pathway. *J. Clin. Invest.* **106**, 1159–1166 (2000).
94. Rajendran, L. *et al.* Efficient inhibition of the Alzheimer's disease beta-secretase by membrane targeting. *Science* **320**, 520–523 (2008).
95. HS, K. *et al.* Carboxyl-terminal fragment of Alzheimer's APP destabilizes calcium homeostasis and renders neuronal cells vulnerable to excitotoxicity. *FASEB J.* **14**, 1508–1517 (2000).
96. Cataldo, A. M. *et al.* A β localization in abnormal endosomes: Association with earliest A β elevations in AD and Down syndrome. *Neurobiol. Aging* **25**, 1263–1272 (2004).
97. Ling, D., Song, H. J., Garza, D., Neufeld, T. P. & Salvaterra, P. M. Abeta42-Induced Neurodegeneration via an Age-Dependent Autophagic-Lysosomal Injury in *Drosophila*. *PLoS One* **4**, 4201 (2009).
98. Nixon, R. A. Endosome function and dysfunction in Alzheimer's disease and other neurodegenerative diseases. *Neurobiol. Aging* **26**, 373–382 (2005).
99. Cataldo, A. M., Barnett, J. L., Pieroni, C. & Nixon, R. A. Increased neuronal endocytosis and protease delivery to early endosomes in sporadic Alz-

- heimer's disease: neuropathologic evidence for a mechanism of increased beta-amyloidogenesis. *J. Neurosci.* **17**, 6142–6151 (1997).
100. Ginsberg, S. D. *et al.* Microarray analysis of hippocampal CA1 neurons implicates early endosomal dysfunction during Alzheimer's disease progression. *Biol. Psychiatry* **68**, 885–893 (2010).
 101. Cataldo, A. M. *et al.* Endocytic pathway abnormalities precede amyloid beta deposition in sporadic Alzheimer's disease and Down syndrome: differential effects of APOE genotype and presenilin mutations. *Am. J. Pathol.* **157**, 277–286 (2000).
 102. Bonnet, U., Scherbaum, N. & Wiemann, M. The endogenous alkaloid harmaline: Acidifying and activity-reducing effects on hippocampal neurons in vitro. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **32**, 362–367 (2008).
 103. Majumdar, A., Capetillo-Zarate, E., Cruz, D., Gouras, G. K. & Maxfield, F. R. Degradation of Alzheimer's amyloid fibrils by microglia requires delivery of CIC-7 to lysosomes. *Mol. Biol. Cell* **22**, 1664 (2011).
 104. Lee, J. H. *et al.* Lysosomal proteolysis and autophagy require presenilin 1 and are disrupted by Alzheimer-related PS1 mutations. *Cell* **141**, 1146–1158 (2010).
 105. Zhao, L. *et al.* Structural analysis of asparaginyl endopeptidase reveals the activation mechanism and a reversible intermediate maturation stage. *Cell Research* **24**, 344–358 (2014).
 106. Forester, B. P. *et al.* Age-related changes in brain energetics and phospholipid metabolism. *NMR Biomed.* **23**, 242–250 (2010).
 107. Zhang, Z. *et al.* Cleavage of tau by asparagine endopeptidase mediates the neurofibrillary pathology in Alzheimer's disease. *Nat. Med.* **20**, 1254–1262 (2014).
 108. Wolfe, D. M. *et al.* Autophagy failure in Alzheimer's disease and the role of defective lysosomal acidification. *Eur. J. Neurosci.* **37**, 1949–1961 (2013).
 109. Chintapalli, V. R. *et al.* Transport proteins NHA1 and NHA2 are essential for survival, but have distinct transport modalities. *Proc. Natl. Acad. Sci. U. S. A.* **112**, 11720–11725 (2015).
 110. Fuster, D. G. & Alexander, R. T. Traditional and emerging roles for the SLC9 Na⁺/H⁺ exchangers. *Pflugers Arch.* **466**, 61–76 (2014).
 111. Windler, F. *et al.* The solute carrier SLC9C1 is a Na⁺/H⁺-exchanger gated by an S4-type voltage-sensor and cyclic-nucleotide binding. *Nat. Commun.* **9**, (2018).
 112. Huang, Y., Chen, W., Dotson, D. L., Beckstein, O. & Shen, J. Mechanism of pH-dependent activation of the sodium-proton antiporter NhaA. *Nat. Commun.* **7**, (2016).
 113. Maes, M., Rimon, A., Kozachkov-Magrisso, L., Friedler, A. & Padan, E. Revealing the Ligand Binding Site of NhaA Na⁺/H⁺ Antiporter and Its pH Dependence. *J. Biol. Chem.* **287**, 38150 (2012).
 114. Wakabayashi, S., Hisamitsu, T., Pang, T. & Shigekawa, M. Kinetic dissection of two distinct proton binding sites in Na⁺/H⁺ exchangers by measurement of reverse mode reaction. *J. Biol. Chem.* **278**, 43580–43585 (2003).

115. Wakabayashi, S., Hisamitsu, T., Pang, T. & Shigekawa, M. Mutations of Arg440 and Gly455/Gly456 oppositely change pH sensing of Na⁺/H⁺ exchanger 1. *J. Biol. Chem.* **278**, 11828–11835 (2003).
116. Touret, N., Poujeol, P. & Counillon, L. Second-site revertants of a low-sodium-affinity mutant of the Na⁺/H⁺ exchanger reveal the participation of TM4 into a highly constrained sodium-binding site. *Biochemistry* **40**, 5095–5101 (2001).
117. Slepkov, E. R., Rainey, J. K., Sykes, B. D. & Fliegel, L. Structural and functional analysis of the Na⁺/H⁺ exchanger. *Biochem. J.* **401**, 623–633 (2007).
118. Omasits, U., Ahrens, C. H., Müller, S. & Wollscheid, B. Protter: interactive protein feature visualization and integration with experimental proteomic data. *Bioinformatics* **30**, 884–886 (2014).
119. Miyazaki, E., Sakaguchi, M., Wakabayashi, S., Shigekawa, M. & Mihara, K. NHE6 protein possesses a signal peptide destined for endoplasmic reticulum membrane and localizes in secretory organelles of the cell. *J. Biol. Chem.* **276**, 49221–49227 (2001).
120. Numata, M., Petrecca, K., Lake, N. & Orlowski, J. Identification of a mitochondrial Na⁺/H⁺ exchanger. *J. Biol. Chem.* **273**, 6951–6959 (1998).
121. Ohgaki, R., Fukura, N., Matsushita, M., Mitsui, K. & Kanazawa, H. Cell surface levels of organellar Na⁺/H⁺ exchanger isoform 6 are regulated by interaction with RACK1. *J. Biol. Chem.* **283**, 4417–4429 (2008).
122. Pulakat, L. *et al.* Ligand-dependent complex formation between the Angiotensin II receptor subtype AT2 and Na⁺/H⁺ exchanger NHE6 in mammalian cells. *Peptides (N.Y.)*. **26**, 863–873 (2005).
123. Gao, L. & Zucker, I. H. AT2 receptor signaling and sympathetic regulation. *Curr. Opin. Pharmacol.* **11**, 124–130 (2011).
124. Gendron, L., Payet, M. D. & Gallo-Payet, N. The angiotensin type 2 receptor of angiotensin II and neuronal differentiation: from observations to mechanisms. *J. Mol. Endocrinol.* **31**, 359–372 (2003).
125. Guimond, M. O. & Gallo-Payet, N. How does angiotensin AT(2) receptor activation help neuronal differentiation and improve neuronal pathological situations? *Front. Endocrinol. (Lausanne)*. **3**, (2012).
126. Hill, J. K. *et al.* Vestibular Hair Bundles Control pH with (Na⁺, K⁺)/H⁺ Exchangers NHE6 and NHE9. *The Journal of Neuroscience* **26**, 9944 (2006).
127. Liu, L., Schlesinger, P. H., Slack, N. M., Friedman, P. A. & Blair, H. C. High capacity Na⁺/H⁺ exchange activity in mineralizing osteoblasts. *J. Cell. Physiol.* **226**, 1702–1712 (2011).
128. Lein, E. S. *et al.* Genome-wide atlas of gene expression in the adult mouse brain. *Nature* **445**, 168–176 (2007).
129. Deane, E. C. *et al.* Enhanced recruitment of endosomal Na⁺/H⁺ exchanger NHE6 into Dendritic spines of hippocampal pyramidal neurons during NMDA receptor-dependent long-term potentiation. *J. Neurosci.* **33**, 595–610 (2013).

130. Park, M. *et al.* Plasticity-induced growth of dendritic spines by exocytic trafficking from recycling endosomes. *Neuron* **52**, 817–830 (2006).
131. Park, M., Penick, E. C., Edwards, J. G., Kauer, J. A. & Ehlers, M. D. Recycling endosomes supply AMPA receptors for LTP. *Science* **305**, 1972–1975 (2004).
132. Chao, M. V. Neurotrophins and their receptors: a convergence point for many signalling pathways. *Nat. Rev. Neurosci.* **4**, 299–309 (2003).
133. Huang, E. J. & Reichardt, L. F. Neurotrophins: roles in neuronal development and function. *Annu. Rev. Neurosci.* **24**, 677–736 (2001).
134. Rex, C. S. *et al.* Brain-Derived Neurotrophic Factor Promotes Long-Term Potentiation-Related Cytoskeletal Changes in Adult Hippocampus. *The Journal of Neuroscience* **27**, 3017 (2007).
135. Koleske, A. J. Molecular mechanisms of dendrite stability. *Nat. Rev. Neurosci.* **14**, 536–550 (2013).
136. Das, U. *et al.* Activity-induced convergence of APP and BACE-1 in acidic microdomains via an endocytosis-dependent pathway. *Neuron* **79**, 447–460 (2013).
137. Mellman, I. The importance of being acid: the role of acidification in intracellular membrane traffic. *Journal of Experimental Biology* **172**, 39–45 (1992).
138. Kondapalli, K. C., Prasad, H. & Rao, R. An inside job: How endosomal Na⁺/H⁺ exchangers link to autism and neurological disease. *Front. Cell. Neurosci.* **8**, 172 (2014).
139. Ohgaki, R., Van Ijzendoorn, S. C. D., Matsushita, M., Hoekstra, D. & Kanazawa, H. Organellar Na⁺/H⁺ exchangers: Novel players in organelle pH regulation and their emerging functions. *Biochemistry* **50**, 443–450 (2011).
140. Brett, C. L., Wei, Y., Donowitz, M. & Rao, R. Human Na⁺/H⁺ exchanger isoform 6 is found in recycling endosomes of cells, not in mitochondria. *Am. J. Physiol. Cell Physiol.* **282**, (2002).
141. Kondapalli, K. C., Prasad, H. & Rao, R. An inside job: how endosomal Na⁽⁺⁾/H⁽⁺⁾ exchangers link to autism and neurological disease. *Front. Cell. Neurosci.* **8**, (2014).
142. Brett, C. L., Tukaye, D. N., Mukherjee, S. & Rao, R. The yeast endosomal Na^(K+)/H⁺ exchanger Nhx1 regulates cellular pH to control vesicle trafficking. *Mol. Biol. Cell* **16**, 1396–1405 (2005).
143. Xinhan, L. *et al.* Na⁺/H⁺ exchanger isoform 6 (NHE6/SLC9A6) is involved in clathrin-dependent endocytosis of transferrin. *Am. J. Physiol. Cell Physiol.* **301**, (2011).
144. Futai, M. *et al.* Luminal Acidification of Diverse Organelles by V-ATPase in Animal Cells. *Journal of Experimental Biology* **203**, 107–116 (2000).
145. Choquet, D. & Triller, A. The Dynamic Synapse. *Neuron* **80**, 691–703 (2013).
146. Zhao, H. *et al.* Emerging roles of Na⁺/H⁺ exchangers in epilepsy and developmental brain disorders. *Prog. Neurobiol.* **138–140**, 19–35 (2016).

147. Verpelli, C., Galimberti, I., Gomez-Mancilla, B. & Sala, C. Molecular basis for prospective pharmacological treatment strategies in intellectual disability syndromes. *Dev. Neurobiol.* **74**, 197–206 (2014).
148. Müller, U. C., Deller, T. & Korte, M. Not just amyloid: physiological functions of the amyloid precursor protein family. *Nature Reviews Neuroscience* **2017 18:5** **18**, 281–298 (2017).
149. APP - NCBI Gene. APP. <https://www.ncbi.nlm.nih.gov/gene/351>.
150. APP, KEGG ORTHOLOGY: K04520. <https://www.kegg.jp/entry/K04520>.
151. Haass, C., Kaether, C., Thinakaran, G. & Sisodia, S. Trafficking and proteolytic processing of APP. *Cold Spring Harb. Perspect. Med.* **2**, (2012).
152. Hefter, D. *et al.* Amyloid Precursor Protein Protects Neuronal Network Function after Hypoxia via Control of Voltage-Gated Calcium Channels. *J. Neurosci.* **36**, 8356–8371 (2016).
153. Mockett, B. G., Richter, M., Abraham, W. C. & Mülle, U. C. Therapeutic Potential of Secreted Amyloid Precursor Protein APPs α . *Front. Mol. Neurosci.* **10**, (2017).
154. Sasaguri, H. *et al.* APP mouse models for Alzheimer's disease preclinical studies. *EMBO J.* **36**, 2473–2487 (2017).
155. Selkoe, D. J. & Hardy, J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol. Med.* **8**, 595–608 (2016).
156. Hefter, D., Ludewig, S., Draguhn, A. & Korte, M. Amyloid, APP, and Electrical Activity of the Brain. *Neuroscientist* **26**, 231–251 (2020).
157. PSEN1 - NCBI Gene. PSEN1. <https://www.ncbi.nlm.nih.gov/gene/5663>.
158. PSEN1, KEGG ORTHOLOGY: K04505. <https://www.kegg.jp/entry/K04505+K04522>.
159. Yang, Y., Bagyinszky, E. & An, S. S. A. Presenilin-1 (PSEN1) Mutations: Clinical Phenotypes beyond Alzheimer's Disease. *Int. J. Mol. Sci.* **24**, (2023).
160. Chen, Q. & Schubert, D. Presenilin-interacting proteins. *Expert Rev. Mol. Med.* **4**, 1–18 (2002).
161. Czech, C., Tremp, G. & Pradier, L. Presenilins and Alzheimer's disease: biological functions and pathogenic mechanisms. *Prog. Neurobiol.* **60**, 363–384 (2000).
162. Kelleher, R. J. & Shen, J. Presenilin-1 mutations and Alzheimer's disease. *Proc. Natl. Acad. Sci. U. S. A.* **114**, 629 (2017).
163. IRS1 - NCBI Gene. IRS1 insulin receptor substrate 1 [Homo sapiens (human)] - Gene - NCBI. <https://www.ncbi.nlm.nih.gov/gene/3667>.
164. IRS1, KEGG ORTHOLOGY: K16172. <https://www.kegg.jp/entry/K16172+K07187+K17445+K17446>.
165. Machado-Neto, J. A. *et al.* Insulin Substrate Receptor (IRS) proteins in normal and malignant hematopoiesis. *Clinics (Sao Paulo)* **73**, (2018).
166. Yong, H. L. & White, M. F. Insulin receptor substrate proteins and diabetes. *Arch. Pharm. Res.* **27**, 361–370 (2004).

167. Sun, X. J. *et al.* Structure of the insulin receptor substrate IRS-1 defines a unique signal transduction protein. *Nature* **352**, 73–76 (1991).
168. White, M. F., Maron, R. & Kahn, C. R. Insulin rapidly stimulates tyrosine phosphorylation of a Mr-185,000 protein in intact cells. *Nature* **318**, 183–186 (1985).
169. CSNK1E - NCBI Gene. CSNK1E casein kinase 1 epsilon [Homo sapiens (human)] - Gene - NCBI. <https://www.ncbi.nlm.nih.gov/gene/1454>.
170. CSNK1E, KEGG ORTHOLOGY: K08960. <https://www.kegg.jp/entry/K08960>.
171. Ko, C. H. & Takahashi, J. S. Molecular components of the mammalian circadian clock. *Hum. Mol. Genet.* **15 Spec No 2**, (2006).
172. Klingaman, E. A., Palmer-Bacon, J., Bennett, M. E. & Rowland, L. M. Sleep Disorders Among People With Schizophrenia: Emerging Research. *Curr. Psychiatry Rep.* **17**, (2015).
173. Pritchett, D. *et al.* Evaluating the links between schizophrenia and sleep and circadian rhythm disruption. *J. Neural Transm. (Vienna)* **119**, 1061–1075 (2012).
174. Pinacho, R. *et al.* Altered CSNK1E, FABP4 and NEFH protein levels in the dorsolateral prefrontal cortex in schizophrenia. *Schizophr. Res.* **177**, 88–97 (2016).
175. Schwab, C. *et al.* Casein kinase 1 delta is associated with pathological accumulation of tau in several neurodegenerative diseases. *Neurobiol. Aging* **21**, 503–510 (2000).
176. Gilfillan, G. D. *et al.* SLC9A6 Mutations Cause X-Linked Mental Retardation, Microcephaly, Epilepsy, and Ataxia, a Phenotype Mimicking Angelman Syndrome. *Am. J. Hum. Genet.* **82**, 1003–1010 (2008).
177. Pescosolido, M. F. *et al.* Genetic and phenotypic diversity of NHE6 mutations in Christianson syndrome. *Ann. Neurol.* **76**, 581–593 (2014).
178. Garbern, J. Y. *et al.* A mutation affecting the sodium/proton exchanger, SLC9A6, causes mental retardation with tau deposition. *Brain* **133**, 1391–1402 (2010).
179. Jiao, J. P. *et al.* Missense variants in SLC9A6 cause partial epilepsy without neurodevelopmental delay. *Orphanet J. Rare Dis.* **20**, 380 (2025).
180. Lan, Y. *et al.* Case Report: Christianson Syndrome Caused by SLC9A6 Mutation: From Case to Genotype-Phenotype Analysis. *Front. Genet.* **12**, (2021).
181. Prasad, H. Genes for endosomal pH regulators NHE6 and NHE9 are dysregulated in the substantia nigra in Parkinson's disease. *Gene* **927**, 148737 (2024).
182. Okochi, R., Nihei, Y. & Ito, D. NDPACX: a newly defined X-linked Parkinsonian syndrome associated with SLC9A6 hemizygote mutation. *Brain Commun.* **7**, (2025).
183. Xian, X. *et al.* Reversal of ApoE4-induced recycling block as a novel prevention approach for Alzheimer's disease. *Elife* **7**, (2018).

184. Pohlkamp, T. *et al.* NHE6 depletion corrects ApoE4-mediated synaptic impairments and reduces Amyloid plaque load. *Elife* **10**, (2021).
185. Huang, N. *et al.* Targeting the HDAC4-NHE6-endosomal pH axis restores amyloid- β clearance and cognitive function in Alzheimer's disease mice. *J. Nanobiotechnology* **24**, (2026).
186. Masereel, B., Pochet, L. & Laeckmann, D. An overview of inhibitors of Na⁺/H⁺ exchanger. *Eur. J. Med. Chem.* **38**, 547–554 (2003).
187. Uria-Avellanal, C. & Robertson, N. J. Na⁺/H⁺ Exchangers and Intracellular pH in Perinatal Brain Injury. *Transl. Stroke Res.* **5**, 79–98 (2014).
188. Pedersen, S. F. & Counillon, L. The SLC9A-C mammalian Na⁺/H⁺ exchanger family: Molecules, mechanisms, and physiology. *Physiol. Rev.* **99**, 2015–2113 (2019).
189. Azzopard, D. *et al.* Prognosis of newborn infants with hypoxic-ischemic brain injury assessed by phosphorus magnetic resonance spectroscopy. *Pediatr. Res.* **25**, 445–451 (1989).
190. Blumberg, R. M., Cady, E. B., Wigglesworth, J. S., McKenzie, J. E. & Edwards, A. D. Relation between delayed impairment of cerebral energy metabolism and infarction following transient focal hypoxia-ischaemia in the developing brain. *Exp. Brain Res.* **113**, 130–137 (1997).
191. Hope, P. L. *et al.* CEREBRAL ENERGY METABOLISM STUDIED WITH PHOSPHORUS NMR SPECTROSCOPY IN NORMAL AND BIRTH-ASPHYXIATED INFANTS. *The Lancet* **324**, 366–370 (1984).
192. Robertson, N. J. *et al.* Cerebral intracellular lactic alkalosis persisting months after neonatal encephalopathy measured by magnetic resonance spectroscopy. *Pediatr. Res.* **46**, 287–296 (1999).
193. Robertson, N. J., Cowan, F. M., Cox, I. J. & Edwards, A. D. Brain alkaline intracellular pH after neonatal encephalopathy. *Ann. Neurol.* **52**, 732–742 (2002).
194. Barkovich, A. J. *et al.* MR Imaging, MR Spectroscopy, and Diffusion Tensor Imaging of Sequential Studies in Neonates with Encephalopathy. *AJNR Am. J. Neuroradiol.* **27**, 533 (2006).
195. Cowan, F. *et al.* Origin and timing of brain lesions in term infants with neonatal encephalopathy. *Lancet* **361**, 736–742 (2003).
196. Vornov, J. J., Thomas, A. G. & Jo, D. Protective effects of extracellular acidosis and blockade of sodium/hydrogen ion exchange during recovery from metabolic inhibition in neuronal tissue culture. *J. Neurochem.* **67**, 2379–2389 (1996).
197. Matsumoto, Y. *et al.* Na⁺/H⁺ Exchanger Inhibitor, SM-20220, Is Protective Against Excitotoxicity in Cultured Cortical Neurons. *Stroke* **35**, 185–190 (2004).
198. Lee, C. *et al.* A two-domain elevator mechanism for sodium/proton antiport. *Nature* **501**, 573–577 (2013).

199. Phillis, J. W., Ren, J. & O'Regan, M. H. Inhibition of Na⁺/H⁺ exchange by 5-(N-ethyl-N-isopropyl)-amiloride reduces free fatty acid efflux from the ischemic reperfused rat cerebral cortex. *Brain Res.* **884**, 155–162 (2000).
200. Pilitsis, J. G., Diaz, F. G., O'Regan, M. H. & Phillis, J. W. Inhibition of Na⁺/H⁺ exchange by SM-20220 attenuates free fatty acid efflux in rat cerebral cortex during ischemia-reperfusion injury. *Brain Res.* **913**, 156–158 (2001).
201. Hwang, I. K. *et al.* Late expression of Na⁺/H⁺ exchanger 1 (NHE1) and neuroprotective effects of NHE inhibitor in the gerbil hippocampal CA1 region induced by transient ischemia. *Exp. Neurol.* **212**, 314–323 (2008).
202. Cengiz, P. *et al.* Inhibition of Na⁺/H⁺ exchanger isoform 1 is neuroprotective in neonatal hypoxic ischemic brain injury. *Antioxid. Redox Signal.* **14**, 1803–1813 (2011).
203. Sevigny, J. *et al.* The antibody aducanumab reduces A β plaques in Alzheimer's disease. *Nature* **537**, 50–56 (2016).