

Flavonoids

Editors

Ayşegül ÇEBİ

E. Gülçeri GÜLEÇ PEKER



© Copyright 2025

Printing, broadcasting and sales rights of this book are reserved to Academician Bookstore House Inc. All or parts of this book may not be reproduced, printed or distributed by any means mechanical, electronic, photocopying, magnetic paper and/or other methods without prior written permission of the publisher. Tables, figures and graphics cannot be used for commercial purposes without permission. This book is sold with banderol of Republic of Türkiye Ministry of Culture.

ISBN	Page and Cover Design
978-625-375-806-6	Typesetting and Cover Design by Akademisyen
Book Title	Publisher Certificate Number
Flavonoids	47518
Editors	Printing and Binding
Ayşegül ÇEBİ ORCID iD: 0000-0003-3804-7966 E. Gülçeri GÜLEÇ PEKER ORCID iD: 0000-0001-7244-0281	Vadi Printingpress
Publishing Coordinator	Bisac Code
Yasin DİLMEN	SOC000000
	DOI
	10.37609/akya.3936

Library ID Card
Flavonoids / ed. Ayşegül Çebi, E. Gülçeri Güleç Peker.
Ankara : Akademisyen Yayınevi Kitabevi, 2025.
162 p. : figure, table. ; 160x235 mm.
Includes References.
ISBN 9786253758066

GENERAL DISTRIBUTION

Akademisyen Kitabevi AŞ

Halk Sokak 5 / A Yenışehir / Ankara

Tel: 0312 431 16 33

siparis@akademisyen.com

www.akademisyen.com

PREFACE

Nature has long been the ultimate pharmacy, providing humanity with a vast reservoir of bioactive compounds that form the backbone of modern pharmacognosy and nutritional science. Among these, flavonoids and related polyphenolic compounds stand out as one of the most diverse and biologically significant groups of secondary metabolites. This book, designed as a comprehensive reference for researchers, clinicians, and students, aims to bridge the gap between fundamental phytochemical structures and their expanding therapeutic horizons.

The volume opens with a foundational exploration of flavonoids, establishing a clear understanding of their general characteristics and classification. Building upon this framework, the subsequent chapters embark on a molecule-specific journey. From the “king of flavonoids,” Quercetin, to the therapeutic potential of Naringenin and the ecological complexities of Isoflavonoids, each chapter dissects the unique molecular architecture and biological mechanisms of these compounds.

Distinguished contributors have provided rigorous analyses of key phytochemicals, including Apigenin, Kaempferol, and the Catechin family, elucidating their roles in cellular signaling and disease prevention. Furthermore, the inclusion of compounds such as Resveratrol, Cyanidin, Rutin, and Anthocyanins broadens the scope to cover critical antioxidants that are pivotal in combating oxidative stress and chronic inflammation.

By synthesizing current research on molecular regulation with practical health applications, this book offers more than just a chemical inventory; it provides a roadmap for future research in functional foods and pharmaceutical development. We extend our deepest gratitude to the esteemed authors who have poured their expertise into these pages. It is our hope that this collection will serve as a vital resource in unlocking the full potential of nature’s chemistry for human well-being.

Editors

Prof. Dr. Ayşegül ÇEBİ

Assoc. Prof. Dr. E. Gülçeri GÜLEÇ PEKER

Giresun-2025

CONTENTS

Chapter 1	Flavonoids: General Characteristics and Classification	1
	<i>Ayşegül ÇEBİ</i>	
Chapter 2	Quercetin: From Molecular Structure to Therapeutic Horizons.....	11
	<i>Fatih Çağlar ÇELİKEZEN</i>	
Chapter 3	Naringenin	23
	<i>Bahar BİLGİN SÖKMEN</i>	
Chapter 4	Isoflavonoids: Molecular Regulation, Ecological Functions, and Health Applications.....	31
	<i>Ulku BAYKAL</i>	
Chapter 5	Apigenin	51
	<i>Ozlem AYDIN BERKTAS</i>	
Chapter 6	Kaempferol: Phytochemical Structure and Biological Activities.....	59
	<i>Tuba ACET</i>	
Chapter 7	Catechin and its Derivatives.....	69
	<i>Sinem AYDIN</i>	
Chapter 8	Flavanols	83
	<i>Funda DEMİRTAŞ KORKMAZ</i>	
Chapter 9	Resveratrol.....	95
	<i>Zeliha KAYA</i>	
Chapter 10	Cyanidin and Human Health: Mechanisms and Applications	107
	<i>Sevim İpek ACAR CÖMERT</i>	
Chapter 11	Rutin	121
	<i>Bengü TEMİZEL</i>	
Chapter 12	Anthocyanin.....	143
	<i>Yasemin SEVİM</i>	

AUTHORS

Doç. Dr. Tuba ACET

Gümüşhane University, Faculty of Health Sciences, Department of Occupational Health and Safety

Doç. Dr. Sinem AYDIN

Giresun University, Faculty of Science and Arts, Department of Biology

Assoc. Prof. Dr. Ulku BAYKAL

Giresun University, Faculty of Engineering, Department of Genetics and Bioengineering

Assoc. Prof. Ozlem AYDIN BERKTAS

Giresun University, Faculty of Healthy Sciences, Department of Nursing

Asst. Prof. Dr. Sevim İpek ACAR CÖMERT

Giresun University, Faculty of Medicine, Department of Physiology

Prof. Dr. Ayşegül ÇEBİ

Giresun University, Faculty of Health Sciences

Prof. Dr. Fatih Çağlar ÇELİKEZEN

Bitlis Eren University

Prof. Dr. Bahar BİLGİN SÖKMEN

Giresun University, Faculty of Science and Literature, Department of Chemistry

Assist. Prof. Zeliha KAYA

Giresun University, Faculty of Tourism

Asst. Prof. Funda DEMİRTAŞ KORKMAZ

Giresun University, Faculty of Medicine, Department of Medicinal Biology

Dr. Yasemin SEVİM

Giresun University, Faculty of Medicine, Department of Medicinal Biology

Dr. Bengü TEMİZEL

Giresun University

CHAPTER 1

FLAVONOIDS: GENERAL CHARACTERISTICS AND CLASSIFICATION

Ayşegül ÇEBİ¹

Introduction

Flavonoids are a large class of plant **polyphenolic secondary metabolites** widely distributed in fruits, vegetables, grains, bark, roots, stems, and flowers. Chemically they share a C6-C3-C6 skeleton (two benzene rings connected by a three-carbon bridge, typically forming a heterocyclic pyran ring) [1]. The term *flavonoid* derives from the Latin *flavus* (yellow), reflecting that many flavonoids are yellow pigments in plants. Historically, flavonoids entered scientific awareness in the 1930s: Albert Szent-Györgyi observed that crude citrus extracts contained unknown yellow compounds that enhanced vitamin C's effect on capillary strength, tentatively calling these substances “vitamin P” [2,3]. Later work revealed that “vitamin P” was not a true vitamin but a mixture of flavonoids, and no deficiency syndrome exists for it.

Flavonoids play critical roles in plant physiology and human nutrition. In plants, they contribute to flower and fruit pigmentation, protect against UV radiation, and mediate signaling in growth and defense [4]. In human health and nutrition, dietary flavonoids are studied for antioxidant, anti-inflammatory, cardioprotective, and anticancer activities (e.g. scavenging free radicals, inhibiting enzymes like xanthine oxidase or COX-2, modulating signaling pathways) [5]. Indeed, flavonoids have been associated with reduced risk of chronic diseases such as cardiovascular disease, diabetes, and cancer, and are regarded as important

¹ Prof. Dr., Giresun University, Faculty of Health Sciences, aysegul.cebi@giresun.edu.tr,
ORCID iD: 0000-0003-3804-7966

REFERENCES

1. Terahara N. 2015. Flavonoids in foods: A review. *Nat. Prod. Commun.* 10: 5221-528.
2. D.M. Kopustinskiene, V. Jakstas, A. Savickas, J. Bernatoniene, Flavonoids as anticancer agents
3. Wujun Chen, Shuai Wang, Yudong Wu, Xin Shen, Shutan Xu, Zhu Guo, Renshuai Zhang, Dongming Xing, The Physiologic Activity and Mechanism of Quercetin-Like Natural Plant Flavonoids, *Current Pharmaceutical Biotechnology*; Volume 21, Issue 8, Year 2020.
4. Jomova, K., Alomar, S. Y., Valko, R., Liska, J., Nepovimova, E., Kuca, K., & Valko, M. (2025). Flavonoids and their role in oxidative stress, inflammation, and human diseases. *Chemico-biological interactions*, 413, 111489. <https://doi.org/10.1016/j.cbi.2025.111489>
5. A. Bitto, D. Giuliani, G. Pallio, N. Irrera, E. Vandini, F. Canalini, D. Zaffe, A. Ottani, L. Minutoli, M. Rinaldi, S. Guarini, F. Squadrito, D. Altavilla, Effects of COX1-2/5-LOX blockade in Alzheimer transgenic 3xTg-AD mice, *Inflamm. Res.* 66 (2017) 389–398, <https://doi.org/10.1007/s00011-017-1022-x>
6. M.A. Incalza, R. D'Oria, A. Natalicchio, S. Perrini, L. Laviola, F. Giorgino, Oxidative stress and reactive oxygen species in endothelial dysfunction associated with cardiovascular and metabolic diseases, *Vasc. Pharmacol.* 100 (2018) 1–19, <https://doi.org/10.1016/j.vph.2017.05.005>.
7. S.M. Crusz, F.R. Balkwill, Inflammation and cancer: advances and new agents, *Nat. Rev. Clin. Oncol.* 12 (2015) 584–596, <https://doi.org/10.1038/nrclinonc.2015.105>.
8. Kim JS. Flavonoids, an Overview: Chemical Structures, Dietary Sources, and Biological Properties. *Food Eng Prog* 2020;24(3):151-163. <https://doi.org/10.13050/foodengprog.2020.24.3.151>
9. Zhao, C. L., Chen, Z. J., Bai, X. S., Ding, C., Long, T. J., Wei, F. G., & Miao, K. R. (2014). Structure-activity relationships of anthocyanidin glycosylation. *Molecular diversity*, 18(3), 687–700. <https://doi.org/10.1007/s11030-014-9520-z>
10. Ferreyra, M. L. F., Serra, P., & Casati, P. (2021). Recent advances on the roles of flavonoids as plant protective molecules after UV and high light exposure. *Physiologia plantarum*, 173(3), 736–749. <https://doi.org/10.1111/ppl.13543>
11. Cetinkaya, A., YAYLA, Ş., HÜRKÜL, M. M., & ÖZKAN, S. A., (2025). Comprehensive review on chromatographic analysis of flavonoids in fruits. *Journal of Chromatography Open* , vol.7.
12. Wang, Y., Chen, J., He, G., Yin, L., & Liao, Y. (2025). Unlocking the potential of flavonoid biosynthesis through integrated metabolic engineering. *Frontiers in plant science*, 16, 1597007. <https://doi.org/10.3389/fpls.2025.1597007>
13. Tapas AR, Sakarkar DM, Kakde RB. 2008. Flavonoids as nutraceuticals: a review. *Trop. J. Pharm. Res.* 7: 1089-1099.
14. Karak P. 2019. Biological activities of flavonoids: An overview. *Int. J. Pharm. Sci.* 10: 1567-1574.
15. Reis JF, Monteiro VV, de Souza Gomes R, do Carmo MM, da Costa GV, Ribera PC, Monteiro MC. 2016. Action mechanism and cardiovascular effect of anthocyanins: a systematic review of animal and human studies. *J. Transl. Med.* 14: 315-331.
16. Sung B, Chung HY, Kim ND. 2016. Role of apigenin in cancer prevention via the induction of apoptosis and autophagy. *J. Cancer Prev.* 21: 216-226.
17. Justesen U, Knuthsen P. 2001. Composition of flavonoids in fresh herbs and calculation of flavonoid intake by use of herbs in traditional Danish dishes. *Food Chem.* 73: 245-250.
18. Day AJ, Mellon F, Barron D, Sarrazin G, Morgan MR, Williamson G. 2001. Human metabolism of dietary flavonoids: Identification of plasma metabolites of quercetin. *Free Radic. Res.* 35: 941-952.
19. Ho PC, Saville DJ, Coville PF, Wanwimolruk S. 2000. Content of CYP3A4 inhibitors, naringin, naringenin and bergapten in grapefruit and grapefruit juice products. *Pharmaceutica. Acta. Helvetiae.* 74: 379-385.
20. Arts ICW, Van De PB, Hollman PC. 2000. Catechin content of foods commonly consumed in the Netherlands. *J. Agric. Food Chem.* 48: 1752-1757.

21. Szkudelska K, Nogowski L. 2007. Genistein-a dietary compound inducing hormonal and metabolic changes. *J. Steroid Biochem. Mol. Biol.* 105: 37-45.
22. Laurenz R, Tumbalam P, Naeve S, Thelen KD. 2017. Determination of isoflavone (genistein and daidzein) concentration of soybean seed as affected by environment and management inputs. *J. Sci. Food Agric.* 97: 3342-3347
23. Ding M, Feng R, Wang SY, Bowman L, Lu Y, Qian Y, Castranova V, Jiang BH, Shi X. 2006. Cyanidin-3-glucoside, a natural product derived from blackberry, exhibits chemopreventive and chemotherapeutic activity. *J. Biol. Chem.* 281: 17359-17368.
24. P. Jones, B. Messner, J. Nakajima, A.R. Schaffner, K. Saito, UGT73C6 and UGT78D1, glycosyltransferases involved in flavonol glycoside biosynthesis in *Arabidopsis thaliana*, *Journal of Biological Chemistry* 278 (2003) 43910e 43918.
25. Panche AN, Diwan AD, Chandra SR. 2016. Flavonoids: an overview. *J. Nutr. Sci.* 5: 1-15.
26. Wang TY, Li Q, Bi KS. 2018. Bioactive flavonoids in medicinal plants: structure, activity and biological fate. *Asian J. Pharm. Sci.* 13: 12-23.
27. Iwashina T. 2013. Flavonoid properties of five families newly incorporated into the order Caryophyllales. *Bull. Natl. Mus. Nat. Sci.* 39: 25-51.
28. Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. 2004. Polyphenolics: food sources and bioavailability. *Am. J. Clin. Nutr.* 79: 727-747.
29. Dixon R, Ferreira D. 2002. Molecules of interest: genistein. *Phytochemistry.* 60: 205-211.

CHAPTER 2

QUERCETIN: FROM MOLECULAR STRUCTURE TO THERAPEUTIC HORIZONS

Fatih Çağlar ÇELİKEZEN¹

Chemical Structure, Physicochemical Properties and Natural Foundations

Quercetin is a naturally occurring flavonol that possesses polyphenolic structure and is categorised within the subclass of flavonoids. Its International Union of Pure and Applied Chemistry (IUPAC) name is 3,3',4',5,7-pentahydroxyflavone with a molecular formula of $C_{15}H_{10}O_7$ and molecular weight of 302.24 g/mol. Structurally, quercetin consists of a three-ring structure (A,B,C rings) bearing five hydroxyl groups at the C3, C5, C7, C3' and C4' positions (1). These groups have been identified as the fundamental determinants of both the biological and chemical activities of the quercetin (2,3). The chemical reactivity capacities of these hydroxyl groups are subject to variation and can be categorised as follows: the sequence of numbers is as follows: $3 > 7 > 3' > 4' > 5$. The 5-hydroxyl group in the A ring structure exhibits significantly lower reactivity compared to the other hydroxyl groups and also shows resistance to glucuronidation (4). The catechol structure in ring B is defined by the presence of two hydroxyl groups located in adjacent positions at the 3' and 4' positions. This feature signifies a critical structural attribute that confers augmented electron-donating capacity and antioxidant potential in comparison to alternative hydroxylation models (4). It is evident that additional structural elements contribute to the biochemical properties of quercetin, including the C2-C3 double bond (unsaturating in the C ring), the 4 oxo group (carbonyl functionally at the 4 position), and the strategically

¹ Prof.Dr., Bitlis Eren University, ccelikezen@beu.edu.tr, ORCID iD: 0000-0001-5489-7384

mechanisms stop it from being harmful when it is eaten (76). People generally tolerate quercetin well, and researchers have extensively studied its safety in humans (27,77). Quercetin supplementation has been shown to be generally safe, with only a small number of mild side effect being reported in studies. The most common of these are gastrointestinal disturbances, headaches and tingling sensations, particularly at high doses (27,77). The best amount of quercetin depends on how it is used and how its made. Researchers have administrated doses ranging from 500 to 2000 mg/day in clinical studies (78). Conventional quercetin supplements generally provide daily dose of up to 1000 miligrams (77). It is generally safe for short-term use at common supplemental doses, but not for specific risk groups. Further long-term studies are neccessary to assess its safety for prolonged, high dose consumption (77).

REFERENCES

1. Sobhy R, Khalifa I, Rahaman A, Zeng XA, Nawaz A, Walayat N. Quercetin: The Biological Effects, Chemical Steadiness, Metabolism, and Delivery Systems. İçinde: Xiao J, (Eds). *Hand-book of Dietary Flavonoids*. Cham: Springer International Publishing; 2023. 1-33. https://doi.org/10.1007/978-3-030-94753-8_12-1
2. Mutha RE, Tatiya AU, Surana SJ. Flavonoids as natural phenolic compounds and their role in therapeutics: an overview. *Future Journal of Pharmaceutical Sciences*. 2021;7(1):25.
3. Roy A, Khan A, Ahmad I, Alghamdi S, Rajab BS, Babalghith AO, vd. Flavonoids a Bioactive Compound from Medicinal Plants and Its Therapeutic Applications. Ullah R, editör. *BioMed Research International*. 2022;2022(1):5445291.
4. Carrillo-Martinez EJ, Flores-Hernández FY, Salazar-Montes AM, Nario-Chaidez HF, Hernández-Ortega LD. Quercetin, a Flavonoid with Great Pharmacological Capacity. *Molecules*. 25 2024;29(5):1000.
5. Ulusoy HG, Sanlier N. A minireview of quercetin: from its metabolism to possible mechanisms of its biological activities. *Critical Reviews in Food Science and Nutrition*. 2020;60(19):3290-303.
6. Terao J. Potential Role of Quercetin Glycosides as Anti-Atherosclerotic Food-Derived Factors for Human Health. *Antioxidants*. 2023;12(2):258.
7. Xiao L, Luo G, Tang Y, Yao P. Quercetin and iron metabolism: What we know and what we need to know. *Food and Chemical Toxicology*. 2018;114:190-203.
8. Häkkinen SH, Kärenlampi SO, Heinonen IM, Mykkänen HM, Törrönen AR. Content of the Flavonols Quercetin, Myricetin, and Kaempferol in 25 Edible Berries. *Journal of Agriculture and Food Chemistry*. 1999;47(6):2274-9.
9. Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. *The American Journal of Clinical Nutrition*. 2005;81(1):243S-255S.
10. Wiczowski W, Romaszko J, Bucinski A, Szawara-Nowak D, Honke J, Zielinski H, vd. Quercetin from Shallots (*Allium cepa* L. var. aggregatum) Is More Bioavailable Than Its Glucosides , ,3. *The Journal of Nutrition*. 2008;138(5):885-8.
11. Crystal S, Lombard K, Peffley EB, Weixin L. Genetic analysis of quercetin in onion (*Allium cepa* L.) 'Lady Raider'. *Texas Journal of Agriculture and Natural Resources*. 2003;16:24-8.
12. Mitchell AE, Hong YJ, Koh E, Barrett DM, Bryant DE, Denison RF, vd. Ten-Year Comparison of the Influence of Organic and Conventional Crop Management Practices on the Content of Flavonoids in Tomatoes. *Journal of Agriculture and Food Chemistry*. 2007;55(15):6154-9.

13. Hăncianu M, Gîrd CE. Farmacognozie: produse vegetale cu substanțe bioactive. Iași: Polirom; 2020.
14. Rauf A, Imran M, Khan IA, ur-Rehman M, Gilani SA, Mehmood Z, vd. Anticancer potential of quercetin: A comprehensive review. *Phytotherapy Research*. 2018;32(11):2109-30.
15. Sharma A, Kashyap D, Sak K, Tuli HS, Sharma AK. Therapeutic Charm of Quercetin and Its Derivatives: a Review of Research and Patents. *Pharmaceutical Patent Analyst*. 2018;7(1):15-32.
16. Kumar R, Vijayalakshmi S, Nadanasabapathi S. Health Benefits of Quercetin. *Def Life Sc JI*. 3 2017;2(2):142.
17. Mirza MA, Mahmood S, Hilles AR, Ali A, Khan MZ, Zaidi SAA, vd. Quercetin as a Therapeutic Product: Evaluation of Its Pharmacological Action and Clinical Applications—A Review. *Pharmaceuticals*. 2023;16(11):1631.
18. Faridi Esfanjani A, Assadpour E, Jafari SM. Improving the bioavailability of phenolic compounds by loading them within lipid-based nanocarriers. *Trends in Food Science & Technology*. 2018;76:56-66.
19. Jafari SM, McClements DJ. Nanotechnology Approaches for Increasing Nutrient Bioavailability. In: *Advances in Food and Nutrition Research* Elsevier; 2017 p. 1-30.
20. Hai Y, Zhang Y, Liang Y, Ma X, Qi X, Xiao J, vd. Advance on the absorption, metabolism, and efficacy exertion of quercetin and its important derivatives: Absorption, metabolism and function of quercetin. *Food Frontiers*. 2020;1(4):420-34.
21. Graf BA, Ameho C, Dolnikowski GG, Milbury PE, Chen CY, Blumberg JB. Rat Gastrointestinal Tissues Metabolize Quercetin1, 2. *The Journal of Nutrition*. 2006;136(1):39-44.
22. Nielsen ILF, Chee WSS, Poulsen L, Offord-Cavin E, Rasmussen SE, Frederiksen H, vd. Bioavailability Is Improved by Enzymatic Modification of the Citrus Flavonoid Hesperidin in Humans: A Randomized, Double-Blind, Crossover Trial. *The Journal of Nutrition*. 2006;136(2):404-8.
23. Graefe EU, Wittig J, Mueller S, Riethling A, Uehleke B, Drewelow B, vd. Pharmacokinetics and Bioavailability of Quercetin Glycosides in Humans. *The Journal of Clinical Pharmacology*. 2001;41(5):492-9.
24. Wolfram S, Blöck M, Ader P. Quercetin-3-Glucoside Is Transported by the Glucose Carrier SGLT1 across the Brush Border Membrane of Rat Small Intestine. *The Journal of Nutrition*. 2002;132(4):630-5.
25. Gee JM, DuPont MS, Rhodes MJC, Johnson IT. Quercetin Glucosides Interact With the Intestinal Glucose Transport Pathway 11This work was supported by a UK Biotechnology and Biological Sciences Research Council Competitive Strategic Grant. *Free Radical Biology and Medicine*. 1998;25(1):19-25.
26. Walgren RA, Karnaky KJ, Lindenmayer GE, Walle T. Efflux of Dietary Flavonoid Quercetin 4'- β -Glucoside across Human Intestinal Caco-2 Cell Monolayers by Apical Multidrug Resistance-Associated Protein-2. *The Journal of Pharmacology and Experimental Therapeutics*. 2000;294(3):830-6.
27. Aghababaei F, Hadidi M. Recent Advances in Potential Health Benefits of Quercetin. *Pharmaceuticals*. 2023;16(7):1020.
28. Anand David A, Arulmoli R, Parasuraman S. Overviews of biological importance of quercetin: A bioactive flavonoid. *Pharmacognosy Reviews*. 2016;10(20):84.
29. Hadidi M, Orellana-Palacios JC, Aghababaei F, Gonzalez-Serrano DJ, Moreno A, Lorenzo JM. Plant by-product antioxidants: Control of protein-lipid oxidation in meat and meat products. *LWT*. 2022;169:114003.
30. Kalantari H, Foruozandeh H, Khodayar MJ, Siahpoosh A, Saki N, Kheradmand P. Antioxidant and hepatoprotective effects of Capparis spinosa L. fractions and Quercetin on tert-butyl hydroperoxide- induced acute liver damage in mice. *Journal of Traditional and Complementary Medicine*. 2018;8(1):120-7.
31. Xu D, Hu MJ, Wang YQ, Cui YL. Antioxidant Activities of Quercetin and Its Complexes for Medicinal Application. *Molecules*. 21 Mart 2019;24(6):1123.

32. Wang G, Wang Y, Yao L, Gu W, Zhao S, Shen Z, vd. Pharmacological Activity of Quercetin: An Updated Review. Cohen G, editor. *Evidence-Based Complementary and Alternative Medicine*. 2022;2022:1-12.
33. Haenen GRMM, Paquay JBG, Korthouwer REM, Bast A. Peroxynitrite Scavenging by Flavonoids. *Biochemical and Biophysical Research Communications*. 1997;236(3):591-3.
34. Comalada M, Camuesco D, Sierra S, Ballester I, Xaus J, Gálvez J, vd. *In vivo* quercitrin anti-inflammatory effect involves release of quercetin, which inhibits inflammation through down-regulation of the NF- κ B pathway. *European Journal of Immunology*. 2005;35(2):584-92.
35. García-Mediavilla V, Crespo I, Collado PS, Esteller A, Sánchez-Campos S, Tuñón MJ, vd. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. *European Journal of Pharmacology*. 2007;557(2-3):221-9.
36. Yuan K, Zhu Q, Lu Q, Jiang H, Zhu M, Li X, vd. Quercetin alleviates rheumatoid arthritis by inhibiting neutrophil inflammatory activities. *The Journal of Nutritional Biochemistry*. 2020;84:108454.
37. Saeedi-Boroujeni A, Mahmoudian-Sani MR. Anti-inflammatory potential of Quercetin in COVID-19 treatment. *Journal Inflammation*. 2021;18(1):3.
38. Lee KM, Hwang MK, Lee DE, Lee KW, Lee HJ. Protective Effect of Quercetin against Arsenite-Induced COX-2 Expression by Targeting PI3K in Rat Liver Epithelial Cells. *Journal of Agriculture and Food Chemistry*. 2010;58(9):5815-20.
39. Wang S, Yao J, Zhou B, Yang J, Chaudry MT, Wang M, vd. Bacteriostatic Effect of Quercetin as an Antibiotic Alternative In Vivo and Its Antibacterial Mechanism In Vitro. *Journal of Food Protection*. 2018;81(1):68-78.
40. Wang L, Li B, Si X, Liu X, Deng X, Niu X, vd. Quercetin protects rats from catheter-related *Staphylococcus aureus* infections by inhibiting coagulase activity. *Journal Cellular Molecular Medicine*. 2019;23(7):4808-18.
41. Qi W, Qi W, Xiong D, Long M. Quercetin: Its Antioxidant Mechanism, Antibacterial Properties and Potential Application in Prevention and Control of Toxipathy. *Molecules*. 2022;27(19):6545.
42. Khachatoorian R, Arumugaswami V, Raychaudhuri S, Yeh GK, Maloney EM, Wang J, vd. Divergent antiviral effects of bioflavonoids on the hepatitis C virus life cycle. *Virology*. 2012;433(2):346-55.
43. Rojas Á, Del Campo JA, Clement S, Lemasson M, García-Valdecasas M, Gil-Gómez A, vd. Effect of Quercetin on Hepatitis C Virus Life Cycle: From Viral to Host Targets. *Scientific Reports*. 2016;6(1):31777.
44. Cotin S, Calliste CA, Mazon MC, Hantz S, Duroux JL, Rawlinson WD, vd. Eight flavonoids and their potential as inhibitors of human cytomegalovirus replication. *Antiviral Research*. 2012;96(2):181-6.
45. Kim CH, Kim JE, Song YJ. Antiviral Activities of Quercetin and Isoquercitrin Against Human Herpesviruses. *Molecules*. 2020;25(10):2379.
46. Gansukh E, Nile A, Kim DH, Oh JW, Nile SH. New insights into antiviral and cytotoxic potential of quercetin and its derivatives – A biochemical perspective. *Food Chemistry*. 2021;334:127508.
47. Liu Z, Zhao J, Li W, Shen L, Huang S, Tang J, vd. Computational screen and experimental validation of anti-influenza effects of quercetin and chlorogenic acid from traditional Chinese medicine. *Scientific Reports*. 2016;6(1):19095.
48. Smith M, Smith JC. Repurposing Therapeutics for COVID-19: Supercomputer-Based Docking to the SARS-CoV-2 Viral Spike Protein and Viral Spike Protein-Human ACE2 Interface Chemistry; 2020
49. Jo S, Kim S, Shin DH, Kim MS. Inhibition of SARS-CoV 3CL protease by flavonoids. *Journal of Enzyme Inhibition and Medicinal Chemistry*. 2020;35(1):145-51.
50. Cheng Z, Sun G, Guo W, Huang Y, Sun W, Zhao F, vd. Inhibition of hepatitis B virus replication by quercetin in human hepatoma cell lines. *Virologica Sinica*. 2015;30(4):261-8.

51. Alam P, Parvez MK, Arbab AH, Al-Dosari MS. Quantitative analysis of rutin, quercetin, naringenin, and gallic acid by validated RP- and NP-HPTLC methods for quality control of anti-HBV active extract of *Guiera senegalensis*. *Pharmaceutical Biology*. 2017;55(1):1317-23.
52. Yang X, Zhu X, Ji H, Deng J, Lu P, Jiang Z, et al. Quercetin synergistically reactivates human immunodeficiency virus type 1 latency by activating nuclear factor- κ B. *Molecular Medicine Reports*. 17(2):2501-2508.
53. Chaniad P, Wattanapiromsakul C, Pianwanit S, Tewtrakul S. Anti-HIV-1 integrase compounds from *Dioscorea bulbifera* and molecular docking study. *Pharmaceutical Biology*. 2016;54(6):1077-85.
54. Vásquez-Garzón VR, Arellanes-Robledo J, García-Román R, Aparicio-Rautista DI, Villa-Treviño S. Inhibition of reactive oxygen species and pre-neoplastic lesions by quercetin through an antioxidant defense mechanism. *Free Radical Research*. 2009;43(2):128-37.
55. Akan Z, Garip AI. Antioxidants May Protect Cancer Cells from Apoptosis Signals and Enhance Cell Viability. *Asian Pacific Journal of Cancer Prevention*. 2013;14(8):4611-4.
56. Shan BE, Wang MX, Li R qing. Quercetin Inhibit Human SW480 Colon Cancer Growth in Association with Inhibition of Cyclin D₁ and Survivin Expression through Wnt/ β -Catenin Signaling Pathway. *Cancer Investigation*. 2009;27(6):604-12.
57. Pahlke G, Ngiewih Y, Kern M, Jakobs S, Marko D, Eisenbrand G. Impact of Quercetin and EGCG on Key Elements of the Wnt Pathway in Human Colon Carcinoma Cells. *Journal of Agriculture and Food Chemistry*. 2006;54(19):7075-82.
58. Xiang T, Fang Y, Wang S xuan. Quercetin suppresses HeLa cells by blocking PI3K/Akt pathway. *Journal of Huazhong University of Science and Technology [Medical Sciences]*. 2014;34(5):740-4.
59. Gulati N, Laudet B, Zohrabian VM, Murali R, Jhanwar-Uniyal M. The antiproliferative effect of Quercetin in cancer cells is mediated via inhibition of the PI3K-Akt/PKB pathway. *Anticancer Research*. 2006;26(2A):1177-81.
60. Qin Y, He L ya, Chen Y, Wang W yi, Zhao X hua, Wu M yun. [Quercetin affects leptin and its receptor in human gastric cancer MGC-803 cells and JAK-STAT pathway]. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*. 2012;28(1):12-6.
61. Michaud-Levesque J, Bousquet-Gagnon N, Béliveau R. Quercetin abrogates IL-6/STAT3 signaling and inhibits glioblastoma cell line growth and migration. *Experimental Cell Research*. 2012;318(8):925-35.
62. Mukherjee A, Khuda-Bukhsh AR. Quercetin Down-regulates IL-6/STAT-3 Signals to Induce Mitochondrial-mediated Apoptosis in a Non-small- cell Lung-cancer Cell Line, A549. *Journal of Pharmacopuncture*. 2015;18(1):19-26.
63. Asgharian P, Tazekand AP, Hosseini K, Forouhandeh H, Ghasemnejad T, Ranjbar M, vd. Potential mechanisms of quercetin in cancer prevention: focus on cellular and molecular targets. *Cancer Cell International*. 2022;22(1):257.
64. Maggiolini M, Vivacqua A, Fasanella G, Recchia AG, Sisci D, Pezzi V, vd. The G Protein-coupled Receptor GPR30 Mediates c-fos Up-regulation by 17 β -Estradiol and Phytoestrogens in Breast Cancer Cells. *Journal of Biological Chemistry*. 2004;279(26):27008-16.
65. Edwards RL, Lyon T, Litwin SE, Rabovsky A, Symons JD, Jalili T. Quercetin Reduces Blood Pressure in Hypertensive Subjects1,. *The Journal of Nutrition*. 2007;137(11):2405-11.
66. Eger S, Bosy-Westphal A, Seiberl J, Kürbitz C, Settler U, Plachta-Danielzik S, vd. Quercetin reduces systolic blood pressure and plasma oxidised low-density lipoprotein concentrations in overweight subjects with a high-cardiovascular disease risk phenotype: a double-blinded, placebo-controlled cross-over study. *British Journal Nutrition*. 2009;102(7):1065-74.
67. Park HJ, Yang JY, Ambati S, Della-Fera MA, Hausman DB, Rayalam S, vd. Combined Effects of Genistein, Quercetin, and Resveratrol in Human and 3T3-L1 Adipocytes. *Journal of Medicinal Food*. 2008;11(4):773-83.

68. Ahn J, Lee H, Kim S, Park J, Ha T. The anti-obesity effect of quercetin is mediated by the AMPK and MAPK signaling pathways. *Biochemical and Biophysical Research Communications*. 2008;373(4):545-9.
69. Sabogal-Guáqueta AM, Muñoz-Manco JI, Ramírez-Pineda JR, Lamprea-Rodriguez M, Osorio E, Cardona-Gómez GP. The flavonoid quercetin ameliorates Alzheimer's disease pathology and protects cognitive and emotional function in aged triple transgenic Alzheimer's disease model mice. *Neuropharmacology*. 2015;93:134-45.
70. Lakhanpal P, Rai DK. Quercetin: A Versatile Flavonoid. *Internet Journal of Medical Update - EJOURNAL* [Online]. 2007;2(2).
71. Tavakoli Z, Tahmasebi Dehkordi H, Lorigooini Z, Rahimi-Madiseh M, Korani MS, Amini-Khoei H. Anticonvulsant effect of quercetin in pentylenetetrazole (PTZ)-induced seizures in male mice: The role of anti-neuroinflammatory and anti-oxidative stress. *International Immunopharmacology*. 2023;116:109772.
72. Kobori M, Masumoto S, Akimoto Y, Takahashi Y. Dietary quercetin alleviates diabetic symptoms and reduces streptozotocin-induced disturbance of hepatic gene expression in mice. *Molecular Nutrition and Food Research*. 2009;53(7):859-68.
73. Kim JH, Kang MJ, Choi HN, Jeong SM, Lee YM, Kim JI. Quercetin attenuates fasting and post-prandial hyperglycemia in animal models of diabetes mellitus. *Nutrition Research and Practice*. 2011;5(2):107.
74. Haddad P, Eid H. The Antidiabetic Potential of Quercetin: Underlying Mechanisms. *CMC*. 2017;24(4):355-64.
75. Tadera K, Minami Y, Takamatsu K, Matsuoka T. Inhibition of .ALPHA.-Glucosidase and .ALPHA.-Amylase by Flavonoids. *Journal of Nutritional Science and Vitaminology*. 2006;52(2):149-53.
76. Harwood M, Danielewska-Nikiel B, Borzelleca JF, Flamm GW, Williams GM, Lines TC. A critical review of the data related to the safety of quercetin and lack of evidence of in vivo toxicity, including lack of genotoxic/carcinogenic properties. *Food and Chemical Toxicology*. 2007;45(11):2179-205.
77. Andres S, Pevny S, Ziegenhagen R, Bakhiya N, Schäfer B, Hirsch-Ernst KI, vd. Safety aspects of the use of quercetin as a dietary supplement. *Molecular Nutrition and Food Research*. 2018;62(1):1700447.
78. Devi V, Deswal G, Dass R, Chopra B, Kriplani P, Grewal AS, vd. Therapeutic Potential and clinical effectiveness of quercetin: a dietary supplement. *RAFNA*. 2024;15(1):13-32.

CHAPTER 3

NARINGENIN

Bahar BİLGİN SÖKMEN¹

Giriş

Flavonoids are polyphenolic compounds naturally found in plants and known to have multifaceted effects on human health. Plant polyphenols are increasingly being investigated for their potential to prevent chronic diseases and maintain health. Naringenin (4',5,7-trihydroxyflavanone) is found in particularly high concentrations in citrus fruits such as grapefruit, oranges, and tangerines. Its high antioxidant capacity and versatile biological effects have made this compound an important candidate for functional foods and pharmaceuticals.

Structural Properties of Naringenin

The disaccharide derivative naringin has a 2-O-(α -L-rhamnopyranosyl)- α -D-glucopyranosyl group replaced by an α -L-rhamnopyranosyl group at position 7 through a glycosidic bond. The melting point of naringin is reported as 83°C. Its solubility in a given solvent is 1 mg/mL at a temperature of 40°C (1).

Chemically, it is a glycoside made up of the disaccharide neohesperidone and the flavone naringenin. The structure of naringenin features a flavonoid core, a heterocyclic pyran ring, and two phenolic rings. The compound has a molecular weight of 580.54 g/mol and a molecular formula of $C_{27}H_{32}O_{14}$ (2).

The molecular structure of naringin (4',5,7-trihydroxyflavanone) is shown in Figure 1. With rising temperature, the solubility of naringin and its aglycone

¹ Prof. Dr., Giresun University, Faculty of Science and Literature, Department of Chemistry, bahar.sokmen@giresun.edu.tr, ORCID iD: 0000-0002-0981-9413

promote synaptic plasticity, nerve cell survival, and prevent neuronal damage, reducing and promoting neuroinflammation (25).

Antidiabetic Effects

Naringin has been shown to increase insulin sensitivity and may improve glucose uptake by cells (26). Naringin has the ability to inhibit certain enzymes involved in carbohydrate metabolism, such as glucosidase. By blocking this enzyme, naringin can lower postprandial glucose levels, which in turn slows the digestion and absorption of carbohydrates.

Naringin is a potent biomolecule with promising potential to improve outcomes in individuals with diabetes. (27). Naringin modulates multiple metabolic processes by limiting insulin secretion and sensitivity, regulating glucose transporters, enhancing peripheral glucose uptake, reducing hepatic glucose production and intestinal glucose absorption, and decreasing cholesterol synthesis, blood lipid levels, oxidative stress, and inflammation (28).

Conclusion

Naringenin, a natural flavanone with broad-spectrum biological activities, holds promise for the prevention and treatment of various chronic diseases. However, its low bioavailability and potential for drug interactions are important issues to address before clinical application. More comprehensive clinical studies will provide clearer information regarding therapeutic doses and the long-term safety profile of naringenin. Naringenin, with its potent antioxidant and anti-inflammatory properties, has significant potential in the prevention and treatment of metabolic, cardiovascular, and neurological diseases. Its multifaceted biological effects and ease of availability from natural sources make it a valuable bioactive compound at the heart of modern health research.

REFERENCES

1. Aherne SA, O'Brien NM. Dietary flavonols: Chemistry, food content, and metabolism. *Nutrition Journal*. 2002; 18(1): 75–81. doi: 10.1016/s0899-9007(01)00695-5.
2. Alam MA, Subhan N, Rahman MM, et al., Effect of citrus flavonoids, naringin and naringenin, on metabolic syndrome and their mechanisms of action. *Advances in Nutrition*. 2014; 5(4): 404–417. doi: 10.3945/an.113.005603
3. Zhang L, Song L, Zhang P, Liu T, Zhou L, Yang G, Lin R, Zhang J. Solubilities of naringin and naringenin in different solvents and dissociation constants of naringenin. *Journal of Chemical & Engineering Data*. 2015; 60(3): 932–940. doi: 10.1021/je501004g
4. Aron PM, Kennedy JA. Flavan-3-ols: Nature, occurrence and biological activity. *Molecular Nutrition & Food Research*. 2008; 52(1): 79–104. doi: 10.1002/mnfr.200700137

5. Wang S, Yang C, Tu H, et al., Characterization and metabolic diversity of flavonoids in Citrus species. *Scientific Reports*. 2017; 7(1): 10549. doi: 10.1038/s41598-017-10970-2
6. Gaitan A, Ravetti S, Garro AG, et al., Preformulation studies and *in vitro* cytotoxicity of naringin. *Drug Development and Industrial Pharmacy*. 2025; 51 (4): 344–353. doi: 10.1080/03639045.2025.2471912.
7. Memariani Z, Abbas SQ, Ul H.S. et al., Naringin and naringenin as anticancer agents and adjuvants in cancer combination therapy: Efficacy and molecular mechanisms of action, a comprehensive narrative review. *Pharmacological Research*. 2021; 171: 105264. doi: 10.1016/j.phrs.2020.105264
8. Stabrauskienė J, Dalia M, Kopustinskiene M, et al. Naringin and naringenin: Their mechanisms of action and potential anticancer activities. *Biomedicines*. 2022; 10(7): 1686 doi: 10.3390/biomedicines10071686
9. Rebello, C. J., Robbie AB, Juan JLL, et al. Safety and pharmacokinetics of naringenin: A randomized, controlled, single-ascending-dose clinical trial. *Diabetes Obesity and Metabolism*, 2019; 22(1): 91–98. doi: 10.1111/dom.13868.
10. Kawaguchi K, Maruyama H, Hasunuma et al., Suppression of inflammatory responses after onset of collagen induced arthritis in mice by oral administration of the Citrus flavanone naringin. *Immunopharmacology and Immunotoxicology*. 2011; 33(4): 723–729. doi: 10.3109/08923973.2011.564186.
11. Bailey, D.G.; Dresser, G.K. Interactions between grape fruit juice and cardiovascular drugs. *American Journal of Cardiology*. 2004; 4(5): 281–297. doi: 1175-3277/04/0005-0281/\$31.00/0
12. Hsiu SL, Huang TY, Hou, YC, et al., Comparison of metabolic pharmacokinetics of naringin and naringenin in rabbits. *Life Sciences*. 2002; 70(13): 1481–1489. doi: 10.1016/s0024-3205(01)01491-6
13. Tsai YJ, Tsai T.H. Mesenteric lymphatic absorption and the pharmacokinetics of naringin and naringenin in rats. *Journal of Agricultural and Food Chemistry*. 2012; 60(51): 12435–12442. doi:10.1021/jf301962g.
14. Tesseromatis C, Alevizou A. The role of the protein-binding on the mode of drug action as well as the interactions with other drugs. *European Journal of Drug Metabolism and Pharmacokinetics*. 2008; 33(4): 225–230. doi: 10.1007/BF03190876
15. Najmanová I, Vopršalová M, Saso L, et al., The pharmacokinetics of flavanones. *Critical Reviews in Food Science and Nutrition*. 2020; 60(18): 3155–3171. doi: 10.1080/10408398.2019.1679085.
16. Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: Chemistry, metabolism and structure activity relationships, *Journal of Nutritional Biochemistry*, 2002; 13: 572–584. PMID: 12550068 doi: 10.1016/S0955-2863(02)00208-5.
17. Hernández-Aquino E, Muriel P. Beneficial effects of naringenin in liver diseases: Molecular Mechanisms, *World Journal of Gastroenterology*. 2018; 24(16): 1679–1707. doi: 10.3748/wjg.v24.i16.1679.
18. Joshi R, Kulkarni YA, Wairkar S. Pharmacokinetic, pharmacodynamic and formulations aspects of Naringenin: An update, *Life Sciences*. 2018; 215: 43–56. doi:10.1016/j.lfs.2018.10.066.
19. Rahman J, Tareq AM, Hossain M, et al. Biological evaluation, DFT calculations and molecular docking studies on the antidepressant and cytotoxicity activities of Cynaspectinata Buch. - Ham. Compounds. *Pharmaceuticals*. 2020; 13(9): 232. doi: 10.3390/ph13090232.
20. Arafah A, Rehman MU, Mir TM, Wal, AF, et al. Multi-Therapeutic Potential of Naringenin (40,5,7-Trihydroxyflavone): Experimental Evidence and Mechanisms. *Plants*. 2020; 9(12): 1784. doi: 10.3390/plants9121784
21. Ginwala R, Bhavsar R, Chigbu DI, et al. Potential Role of Flavonoids in Treating Chronic Inflammatory Diseases with a Special Focus on the Anti-Inflammatory Activity of Apigenin. *Antioxidants*. 2019; 8: 35. doi: 10.3390/antiox8020035

22. Su W, Wang Y, Li P, et al., The potential application of the traditional Chinese herb *Exocarpium Citrigrandis* in the prevention and treatment of COVID-19. *Traditional Chinese Medicine*. 2020; 5(3): 160–166. doi: 10.12032/TMR20200406172
23. Yan Y, Zhou H, Wu, C, et al., Ultrasound-assisted aqueous two-phase extraction of synephrine, naringin, and neohesperidin from *Citrus aurantium* L. *Fruit lets. Preparative Biochemistry & Biotechnology*. 2021; 51(8): 780–791. doi:10.1080/10826068.2020.1858427
24. Yuan X, Wu H, Xu H, et al., Notch signaling: An emerging therapeutic target for cancer treatment. *Cancer Letters*. 2015; 369(1): 20–27. doi:10.1016/j.canlet.2015.07.048
25. Sun R, Liu C, Liu J, et al. Integrated network pharmacology and experimental verification reveal protective effect of naringenin in chronic wounds. *Scientific Reports*. 2023; 13(1): 132. doi:10.1038/s41598-022-26043-y
26. Ahmed S, Khan H, Aschner M, et al., Therapeutic potential of naringin in neurological disorders. *Food and Chemical Toxicology*. 2019; 132: 110646. doi: 10.1016/j.fct.2019.110646
27. Mahmoud AM, Hussein OE. Anti-diabetic effect of naringin: Insights into the molecular mechanism. *Diabetes & Obesity International Journal*. 2016; 1(5): 1-3. doi:10.23880/DOIJ-16000128
28. Pandey VK, Shams R, Singh R, et al., Comprehensive review on clove (*Caryophyllus aromaticus* L.) essential oil and its significance in the formulation of edible coatings for potential food applications. *Frontiers in Nutrition*. 2022; 9: 987674. doi:10.3389/fnut.2022.987674

CHAPTER 4

ISOFLAVONOIDS: MOLECULAR REGULATION, ECOLOGICAL FUNCTIONS, AND HEALTH APPLICATIONS

Ulku BAYKAL¹

INTRODUCTION

Flavonoids represent one of the most diverse groups of plant specialized metabolites, with more than 10,000 known structures contributing to pigmentation, UV protection, and ecological interactions (2). Among flavonoid subclasses, isoflavonoids occupy a unique position because their biosynthesis depends on the formation of the isoflavone skeleton, catalyzed by a branch-point enzyme, isoflavone synthase (IFS). This structural innovation, found primarily in the Fabaceae (legume) family, underpins both the chemical diversity and the broad functional roles of isoflavonoids (2,3). Isoflavonoids are distinguished not only by their structural diversity but also by the breadth of their ecological and physiological functions: they act as phytoalexins in disease resistance, as signaling molecules that initiate symbiotic nitrogen fixation with rhizobia, and as bioactive phytoestrogens in human consumption (6, 7, 9, 10, 12, 17, 19, 20).

Advances in genomics, transcriptomics, metabolomics, and epigenomics have transformed our understanding of how plants control isoflavonoid biosynthesis (4, 16). Regulatory networks in legumes integrate multiple layers of control, including transcription factors (R2R3-MYB, bHLH, bZIP, NAC, WRKY), epigenetic modifications (DNA methylation, histone acetylation, H3K4me3 enrichment), and post-transcriptional regulators (microRNAs, small interfering RNAs) (18, 21, 24, 25, 27, 29). These regulatory components interact with developmental signals

¹ Assoc. Prof. Dr., Giresun University, Faculty of Engineering, Department of Genetics and Bioengineering, ulku.baykal@giresun.edu.tr, ORCID iD: 0000-0003-2441-0592

In summary, isoflavonoids stand at the intersection of plant defense, ecological communication, metabolic regulation, and human health (1, 4, 8). Continued interdisciplinary research will expand their applications in biotechnology, agriculture, and personalized medicine, while deepening our understanding of the evolutionary and ecological forces that shaped this remarkable class of specialized metabolites.

REFERENCES

1. Geng D, Shen X, Xie Y, Yang Y, Bian R, Gao Y, Li P, Sun L, Feng H, Ma F, Guan Q. Regulation of phenylpropanoid biosynthesis by MdMYB88 and MdMYB124 contributes to pathogen and drought resistance in apple. *Horticulture Research*. 2020;7: 102.
2. Vogt T. Phenylpropanoid biosynthesis. *Molecular Plant*. 2010;3(1): 2–20.
3. Shimada N, Akashi T, Aoki T, Ayabe S. Induction of isoflavonoid pathway in the model legume *Lotus japonicus*: molecular characterization of enzymes involved in phytoalexin biosynthesis. *Plant Science*. 2000; 160(1): 37–47.
4. Yu O, Jez JM. Nature's assembly line: biosynthesis of simple phenylpropanoids and polyketides. *The Plant Journal*. 2008;54(4): 750–762.
5. Lee SG, Krishnan HB, Jez JM. Structural basis for regulation of rhizobial nodulation and symbiosis gene expression by the regulatory protein NodR. *PNAS*. 2014 Apr 29;111(17): 6509–14.
6. Yang Q, Wang G. Isoflavonoid metabolism in leguminous plants: an update and perspectives. *Frontiers in Plant Science*. 2024;15: 1368870.
7. Lygin AV, Zernova OV, Hill CB, et al. Glyceollin is an important component of soybean plant defense against *Phytophthora sojae* and *Macrophomina phaseolina*. *Phytopathology*. 2013;103(10): 984–94.
8. Sajid M, Stone SR, Kaur P. Recent Advances in Heterologous Synthesis Paving Way for Future Green-Modular Bioindustries: A Review With Special Reference to Isoflavonoids. *Frontiers in Bioengineering and Biotechnology*. 2021;9: 673270.
9. Xu W, Dubos C, Lepiniec L. Transcriptional control of flavonoid biosynthesis by MYB-bHLH-WDR complexes. *Trends in Plant Science*. 2015;20(3): 176–185.
10. Chezem WR, Clay NK. Regulation of plant secondary metabolism and associated specialized cell development by MYBs and bHLHs. *Phytochemistry*. 2016;131: 26–43.
11. Yi J, Derynck MR, Li X, et al. single-repeat MYB transcription factor, GmMYB176, regulates CHS8 gene expression and affects isoflavonoid biosynthesis in soybean. *The Plant Journal*. 2010;62(6): 1019–1034.
12. Hayashi K, Alseekh S, Fernie AR. Genetic and epigenetic control of the plant metabolome. *Proteomics*. 2023;23(13–14): e2200104.
13. Nakayama T, Takahashi S, Waki T. Formation of Flavonoid Metabolons: Functional Significance of Protein-Protein Interactions and Impact on Flavonoid Chemodiversity. *Frontiers in Plant Science*. 2019;10: 821
14. Sharma D, Tiwari M, Pandey A, Bhatia C, Sharma A, Trivedi PK. MicroRNA858 Is a Potential Regulator of Phenylpropanoid Pathway and Plant Development. *Plant Physiology*. 2016;171(2): 944–59.

15. Ražná K, Harenčár Ľ, Kučka M. The Involvement of microRNAs in Plant Lignan Biosynthesis-Current View. *Cells*. 2022;11(14): 2151.
16. Cui Q, Li X, Hu S, Yang D, Abozeid A, Yang Z, Jiang J, Ren Z, Li D, Li D, Zheng L, Qin A. The Critical Role of Phenylpropanoid Biosynthesis Pathway in Lily Resistance Against Gray Mold. *International Journal of Molecular Sciences*. 2024;25(20): 11068.
17. Weston LA, Mathesius U. Flavonoids: their structure, biosynthesis and role in the rhizosphere, including allelopathy. *Journal of Chemical Ecology* 2013;39(2): 283–297.
18. Hassan S, Mathesius U. The role of flavonoids in root-rhizosphere signalling: opportunities and challenges for improving plant-microbe interactions. *The Journal of Experimental Botany*. 2012;63(9): 3429–44.
19. Hu L, Robert CAM, Cadot S, et al. Root exudate metabolites drive plant-soil feedbacks on growth and defense by shaping the rhizosphere microbiota. *Nature Communications*. 2018;9: 2738.
20. Zhalnina, K., Louie, K.B., Hao, Z. et al. Dynamic root exudate chemistry and microbial substrate preferences drive patterns in rhizosphere microbial community assembly. *Nature Microbiology* 2018;3: 470–480.
21. Yasmin F, Zhang H, Leamy L, Wang B, Winnike J, Reid RW, Brouwer CR, Song BH. Genetic basis and selection of glyceollin elicitation in wild soybean. *Frontiers in Plant Science*. 2024;15: 1240981.
22. Dao TT, Linthorst HJ, Verpoorte R. Chalcone synthase and its functions in plant resistance. *Phytochem Rev*. 2011;10(3): 397–412.
23. Wasternack C, Hause B. Jasmonate–salicylate crosstalk. *Annals of Botany*. 2013;111(6): 1021–1058.
24. Polturak G, Misra RC, El-Demerdash A, et al. Discovery of isoflavone phytoalexins in wheat reveals an alternative route to isoflavonoid biosynthesis. *Nature Communications* 2023;14(1): 6977.
25. Hideg E, Jansen MA, Strid A. UV-B exposure, ROS, and stress: inseparable companions or loosely linked associates? *Trends in Plant Science*. 2013;18(2): 107–15.
26. Jenkins GI. Signal transduction in responses to UV-B radiation. *Annual Review of Plant Biology*. 2009;60: 407–431.
27. Dissanayake BM, Staudinger C, Ranathunge K, et al. Metabolic adaptations leading to an enhanced lignification in wheat roots under salinity stress. *The Plant Journal*. 2024;119(4): 1800–1815.
28. Nakabayashi R, Yonekura-Sakakibara K, Urano K. et al. Enhancement of oxidative and drought tolerance in Arabidopsis by overaccumulation of antioxidant flavonoids. *The Plant Journal*. 2014;77(3): 367–79.
29. Lämke J, Bäurle I. Epigenetic and chromatin-based mechanisms in environmental stress adaptation and stress memory in plants. *Genome Biology* 2017;18(1): 124.
30. Saito K, Matsuda F. Metabolomics for functional genomics, systems biology, and biotechnology. *Annual Review of Plant Biology*. 2010;61: 463–489.
31. Goh YX, Jalil J, Lam KW, Husain K, Premakumar CM. Genistein: A Review on its Anti-Inflammatory Properties. *Frontiers in Pharmacology*. 2022;13: 820969.
32. Sharifi-Rad J, Quispe C, Imran M, et al. Genistein: An integrative overview of its mode of action, pharmacological properties, and health benefits. *Oxidative Medicine and Cellular Longevity*. 2021;2021: 3268136.

33. Shirvanian K, Vali R, Farkhondeh T, et al. Genistein Effects on Various Human Disorders Mediated via Nrf2 Signaling. *Current Molecular Medicine*. 2024;24(1): 40-50.
34. Lampe JW. Isoflavonoid metabolism and health outcomes. *Annual Review of Nutrition*. 2007;27: 505–531.
35. Ionescu VS, Popa A, Alexandru A, Manole E, Neagu M, Pop S. Dietary Phytoestrogens and Their Metabolites as Epigenetic Modulators with Impact on Human Health. *Antioxidants (Basel)*. 2021;10(12): 1893.
36. Rafii F. The role of colonic bacteria in the metabolism of the natural isoflavone daidzin to equol. *Metabolites*. 2015;5(1): 56-73.
37. Yoshikata R., Myint KZY, Taguchi J. Comparison of blood and urine concentrations of equol by LC-MS/MS method and factors associated with equol production in 466 Japanese men and women. *PLoS ONE*. 2024;19(3): e0288946.
38. Rodríguez-Morató J, Farré M, Pérez-Mañá C, et al. Pharmacokinetic Comparison of Soy Isoflavone Extracts in Human Plasma. *Journal of Agricultural and Food Chemistry*. 2015;63(31): 6946-53.
39. Taku K, Melby MK, Kronenberg F, et al. Extracted or synthesized soybean isoflavones reduce menopausal hot flash frequency and severity: systematic review and meta-analysis of randomized controlled trials. *Menopause*. 2012;19(7): 776-90.
40. Barańska A, Kanadys W, Bogdan M, Stępień E, Barczyński B, Kłak A, Augustynowicz A, Szajnik M, Religioni U. The Role of Soy Isoflavones in the Prevention of Bone Loss in Postmenopausal Women: A Systematic Review with Meta-Analysis of Randomized Controlled Trials. *Journal of Clinical Medicine*. 2022;11(16): 4676.
41. Chen LR, Ko NY, Chen KH. Isoflavone Supplements for Menopausal Women: A Systematic Review. *Nutrients*. 2019;11(11): 2649.
42. Zhang P, Du H, Wang J, Pu Y, Yang C, Yan R, Yang H, Cheng H, Yu D. Multiplex CRISPR/Cas9-mediated metabolic engineering increases soya bean isoflavone content and resistance to soya bean mosaic virus. *Plant Biotechnology Journal*. 2020;18(6): 1384-1395.
43. Liu Q, Liu Y, Li G, Savolainen O, Chen Y, Nielsen J. De novo biosynthesis of bioactive isoflavonoids by engineered yeast cell factories. *Nature Communications*. 2021;12(1): 6085.
44. Alexandrov T. Spatial metabolomics and imaging mass spectrometry in the age of artificial intelligence. *Annual Review of Biomedical Data Science*. 2020;3: 61–87.
45. Setchell KDR, Clerici C. Equol: history, chemistry, and health effects. *The Journal of Nutrition*. 2010;140(7): 1355S–1362S.

CHAPTER 5

APIGENIN

Ozlem AYDIN BERKTAS¹

Apigenin

Flavonoids are phenolic compounds naturally found in plants that have multifaceted biological effects on human health. They contribute to plant color, aroma, and flavor, as well as playing a crucial role in their defense against environmental stresses. Within this broad biochemical class, apigenin (4',5,7-trihydroxyflavone) is one of the key flavones of note. (1).

Chemical Structure of Apigenin

Apigenin is a yellow compound belonging to the flavone group and has the chemical formula $C_{15}H_{10}O_5$ (2). Under normal conditions, it exists as yellow crystalline structures and has very limited solubility in water (1.35 $\mu\text{g/mL}$). However, it exhibits high solubility in organic solvents such as ethanol and DMSO, which is one of the primary factors reducing its bioavailability. It has a melting point between 345–350°C, and storage at –20°C is generally recommended (3-5). The structure of apigenin contains three hydroxyl groups (at positions 4', 5, and 7). Its basic skeleton consists of two aromatic rings (A and B) and one heterocyclic pyrone ring (C) (Figure 1). This configuration provides apigenin with high stability and strong free radical scavenging capacity (6). Chemically, apigenin differs from other flavones such as luteolin by the absence of a methoxy or additional hydroxyl group. This structural difference enhances the lipophilic properties of apigenin,

¹ Assoc. Prof., Giresun University, Faculty of Healthy Sciences, Department of Nursing, ozlem.berktas@giresun.edu.tr, ORCID iD: 0000-0002-7235-4890

Studies further indicate that apigenin reduces NF- κ B activation by preventing p65 phosphorylation and by downregulating Akt-mediated signaling (13).

Conclusion

Considering all these findings, apigenin—owing to its chemical stability, strong antioxidant capacity, and lipophilic characteristics—effectively neutralizes free radicals and mitigates cellular damage caused by oxidative stress. In addition, its anti-inflammatory, anti-proliferative, anti-fibrotic, and anticancer properties make apigenin a remarkable natural therapeutic agent in modern pharmacology. Preclinical and clinical studies have demonstrated that apigenin regulates and protects cellular signaling pathways, particularly in cancer, neurodegenerative diseases, diabetes, and autoimmune disorders. With its low toxicity profile and high biological efficacy, apigenin is regarded as a promising phytochemical candidate for use in future pharmaceutical formulations and complementary therapeutic approaches. However, to achieve effective clinical applications, it is crucial to address the low bioavailability of apigenin through pharmacokinetic optimization strategies. Enhancing its absorption and stability will be a key step in fully realizing apigenin's therapeutic potential in human health applications.

REFERENCES

1. Panche AN, Diwan AD, Chandra SR. Flavonoids: An overview. *Journal of Nutritional Science*. 2016;5:e47. doi: 10.1017/jns.2016.41.
2. Nabavi SF, Khan H, D'Onofrio G, Šamec D, Shirooie S, Dehpour AR, et al. Apigenin as neuroprotective agent: Of mice and men. *Pharmacological Research*. 2018;128:359–365. doi: 10.1016/j.phrs.2017.10.018.
3. Nurgali K, Jagoe RT, Abalo R. Adverse effects of cancer chemotherapy: Anything new to improve tolerance and reduce sequelae? *Frontiers Media SA*. 2018. doi: 10.3389/978-2-88945-245-3
4. Zhang J, Liu D, Huang Y, Gao Y, Qian S. Biopharmaceutics classification and intestinal absorption study of apigenin. *International Journal of Pharmaceutics*. 2012;436(1–2):311–317. doi: 10.1016/j.ijpharm.2012.07.032
5. Shukla S, Gupta S. Apigenin: A promising molecule for cancer prevention. *Pharmaceutical Research*. 2010;27(6):962–978. doi: 10.1007/s11095-010-0089-7
6. Miesan KH, Mohamed S. Flavonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) content of edible tropical plants. *Journal of Agricultural and Food Chemistry*. 2001;49(6):3106–3112. doi: 10.1021/jf000892m
7. Hostetler GL, Ralston RA, Schwartz SJ. Flavones: Food sources, bioavailability, metabolism, and bioactivity. *Advances in Nutrition*. 2017;8(3):423–435. doi: 10.3945/an.116.012948
8. Martens S, Mithöfer A. Flavones and flavone synthases. *Phytochemistry*. 2005;66(20):2399–2407. doi: 10.1016/j.phytochem.2005.07.013

9. Mushtaq Z, Sadeer NB, Hussain M, Alsagaby SA, Imran M, Mumtaz T, et al. Therapeutical properties of apigenin: A review on the experimental evidence and basic mechanisms. *International Journal of Food Properties*. 2023;26(1):1914–1939. doi: 10.1080/10942912.2023.2236329
10. Ali F, Rahul, Naz F, Jyoti S, Siddique YH. Health functionality of apigenin: A review. *International Journal of Food Properties*. 2017;20(6):1197–1238. doi: 10.1080/10942912.2017.1327335
11. Yoon JH, Kim MY, Cho JY. Apigenin: A therapeutic agent for treatment of skin inflammatory diseases and cancer. *International Journal of Molecular Sciences*. 2023;24(2):1498. doi: 10.3390/ijms24021498
12. Barky A, Ezz A, Mohammed T. The potential role of apigenin in diabetes mellitus. *International Journal of Clinical Case Reports and Reviews*. 2020;3(1):32–40.
13. Imran M, Gondal TA, Atif M, Shahbaz M, Qaisarani TB, Mughal MH, Salehi B, Martorell M, Sharifi-Rad J. Apigenin as an anticancer agent. *Phytotherapy Research*. 2020;34(8):1812–1828. doi: 10.1002/ptr.6647
14. Kilit AC, Aydemir D. Apiin'in sitotoksik etkisi. *Bilim Armonisi*. 2021;4(2):64–70.
15. Moslehi M, Rezaei S, Talebzadeh P, Ansari MJ, Jawad MA, Jalil AT, et al. Apigenin in cancer therapy: Prevention of genomic instability and anticancer mechanisms. *Clinical and Experimental Pharmacology and Physiology*. 2023;50(1):3–18. doi: 10.1111/1440-1681.13725
16. Susam S, Çıkım G. Apigenin'in potansiyel farmakolojik etkileri üzerine bir derleme. *Arşiv Kaynak Tarama Dergisi*. 2023;32(2):134–150.
17. Gupta S, Afaq F, Mukhtar H. Selective growth-inhibitory, cell-cycle deregulatory and apoptotic response of apigenin in normal versus human prostate carcinoma cells. *Biochemical and Biophysical Research Communications*. 2001;287(4):914–920. doi: 10.1006/bbrc.2001.5669
18. Bi CC, Han WW, Yu JR, Zhang HF, Xing GY, Liu Z. Insights into the pharmacological and therapeutic effects of apigenin in liver injuries and diseases. *Heliyon*. 2023;9(5):e15609. doi: 10.1016/j.heliyon.2023.e15609
19. Harborne JB, Williams CA. Advances in flavonoid research since 1992. *Phytochemistry*. 2000;55(6):481–504. doi: 10.1016/S0031-9422(00)00235-1
20. Shukla S, MacLennan GT, Fu P, Gupta S. Apigenin attenuates insulin-like growth factor-I signaling in an autochthonous mouse prostate cancer model. *Pharmaceutical Research*. 2012;29(6):1506–1517. doi: 10.1007/s11095-012-0675-1
21. Chiang LC, Ng LT, Lin IC, Kuo PL, Lin CC. Anti-proliferative effect of apigenin and its apoptotic induction in human Hep G2 cells. *Cancer Letters*. 2006;237(2):207–214. doi: 10.1016/j.canlet.2005.06.018
22. Hicks DE, Goossens N, Blas-García A, Tsuchida T, Wooden B, Wallace MC, Friedman SL. Transcriptome-based repurposing of apigenin as a potential anti-fibrotic agent targeting hepatic stellate cells. *Scientific Reports*. 2017;7(1):42563. doi: 10.1038/srep42563
23. Malik S, Suchal K, Khan SI, Bhatia J, Kishore K, Dinda AK, Arya DS. Apigenin ameliorates streptozotocin-induced diabetic nephropathy in rats via MAPK-NF- κ B-TNF- α and TGF- β 1-MAPK-fibronectin pathways. *American Journal of Physiology–Renal Physiology*. 2017;313(2):F414–F422. doi: 10.1152/ajprenal.00439.2016

24. Shin GC, Kim C, Lee JM, Cho WS, Lee SG, Jeong M, et al. Apigenin-induced apoptosis is mediated by reactive oxygen species and activation of ERK1/2 in rheumatoid fibroblast-like synoviocytes. *Chemico-Biological Interactions*. 2009;182(1):29–36. doi: 10.1016/j.cbi.2009.08.012
25. Zhang S, Liu X, Sun C, Yang J, Wang L, Liu J, et al. Apigenin attenuates experimental autoimmune myocarditis by modulating Th1/Th2 cytokine balance in mice. *Inflammation*. 2016;39(2):678–686. doi: 10.1007/s10753-015-0288-6
26. Zhao L, Wang JL, Liu R, Li XX, Li JF, Zhang L. Neuroprotective, anti-amyloidogenic and neurotrophic effects of apigenin in an Alzheimer's disease mouse model. *Molecules*. 2013;18(8):9949–9965. doi: 10.3390/molecules18089949
27. Salehi B, Venditti A, Sharifi-Rad M, Kregiel D, Sharifi-Rad J, Durazzo A, et al. The therapeutic potential of apigenin. *International Journal of Molecular Sciences*. 2019;20(6):1305. doi: 10.3390/ijms20061305.

CHAPTER 6

KAEMPFEROL: PHYTOCHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITIES

Tuba ACET¹

Introduction

Kaempferol is a naturally occurring polyphenolic compound belonging to the flavonol subclass of flavonoids and is widely distributed throughout the plant kingdom. With the growing interest in the bioactivity of phenolic compounds, kaempferol has emerged as a prominent phytochemical with potential roles in antioxidant defense, anti-inflammatory responses, neuroprotection, and cardiometabolic regulation (1). In plants, it predominantly occurs as glycosylated derivatives, and its physiological effects in humans are shaped by both its inherent chemical features and metabolic transformation processes.

Kaempferol is abundant in many fruits and vegetables (2,3) and possesses a wide range of therapeutic attributes (figure 1). Numerous studies indicate that it mitigates oxidative stress—one of the underlying drivers of metabolic disorders—by reducing the formation (4) of reactive oxygen species (ROS) (2). Its antitumor and anti-inflammatory activities have also been demonstrated extensively (5). Furthermore, kaempferol is reported to enhance biological performance through hormetic mechanisms, particularly in aging-associated processes (6). The major biological functions attributed to kaempferol are discussed in the following sections.

¹ Doç. Dr., Gümüşhane University, Faculty of Health Sciences, Department of Occupational Health and Safety, tubaacet@gumushane.edu.tr, ORCID iD: 0000-0002-0981-9413

concentrations in cell-culture models (48). Due to insufficient clinical evidence, recommended therapeutic doses are still based solely on experimental data.

Conclusion and Future Perspectives

Kaempferol is a multifunctional flavonoid with protective effects in diverse pathological contexts, including diabetes, obesity, arthritis, glaucoma, osteoporosis, and dermatological disorders. It exerts organ-protective effects on the liver, kidneys, heart, nervous system, and gastrointestinal tract primarily through antioxidant and anti-inflammatory mechanisms. Its anticancer potential is mediated by the modulation of multiple signaling pathways. Nevertheless, limited water solubility, rapid metabolism, and poor oral bioavailability hinder its clinical utility. Novel delivery systems—such as nanocarriers and encapsulation technologies—may help overcome these limitations. However, comprehensive clinical trials are required to confirm its therapeutic efficacy and safety.

REFERENCES

1. Calderon-Montano J, Burgos-Moron E, Perez-Guerrero C, Lopez-Lazaro M. A Review on the Dietary Flavonoid Kaempferol. *Mini-Reviews Med Chem*. 2011;11(4):298–344. doi: 10.2174/138955711795305335.
2. Imran M, Salehi B, Sharifi-Rad J, Gondal TA, Saeed F, Imran A, et al. Kaempferol: A key emphasis to its anticancer potential. *Molecules*. MDPI. 2019;24(12):2277. doi: 10.3390/molecules24122277.
3. Periferakis A, Periferakis K, Badarau IA, Petran EM, Popa DC, Caruntu A, et al. Kaempferol: Antimicrobial Properties, Sources, Clinical, and Traditional Applications. Vol. 23, *International Journal of Molecular Sciences*. MDPI; 23(23): 15054. <https://doi.org/10.3390/ijms232315054>.
4. Chen AY, Chen YC. A review of the dietary flavonoid, kaempferol on human health and cancer chemoprevention. *Food Chemistry*. 2013;138: 2099–107.
5. Devi KP, Malar DS, Nabavi SF, Sureda A, Xiao J, Nabavi SM, et al. Kaempferol and inflammation: From chemistry to medicine. *Pharmacological Research. Academic Press*. 2015;99: 1–10.
6. Calabrese EJ, Pressman P, Hayes AW, Baldwin L, Agathokleous E, Kapoor H, et al. Kaempferol, a widely ingested dietary flavonoid and supplement, enhances biological performance via hormesis, especially for ageing-related processes. *Mech Ageing Dev*. 2025;1:225.
7. Yang L, Gao Y, Bajpai VK, El-Kammar HA, Simal-Gandara J, Cao H, et al. Advance toward isolation, extraction, metabolism and health benefits of kaempferol, a major dietary flavonoid with future perspectives. *Critical Reviews in Food Science and Nutrition*. Taylor and Francis Ltd.; 2023;63: 2773–89.
8. Ren J, Lu Y, Qian Y, Chen B, Wu T, Ji G. Recent progress regarding kaempferol for the treatment of various diseases (Review). *Exp Ther Med*. 2019;18(4):2759–2776. doi: 10.3892/etm.2019.7886.
9. Chen J, Zhong H, Huang Z, Chen X, You J, Zou T. A Critical Review of Kaempferol in Intestinal Health and Diseases. *Antioxidants*. Multidisciplinary Digital Publishing Institute (MDPI); 2023;12(8): 1642. <https://doi.org/10.3390/antiox12081642>
10. Speisky H, Arias-Santé MF, Fuentes J. Oxidation of Quercetin and Kaempferol Markedly Amplifies Their Antioxidant, Cytoprotective, and Anti-Inflammatory Properties. *Antioxidants*. 2023;9;12(1):155. doi: 10.3390/antiox12010155.

11. Dabeek WM, Marra MV. Dietary quercetin and kaempferol: Bioavailability and potential cardiovascular-related bioactivity in humans. *Nutrients*. MDPI. 2019;25;11(10):2288. doi: 10.3390/nu11102288.
12. García-Mediavilla V, Crespo I, Collado PS, Esteller A, Sánchez-Campos S, Tuñón MJ, et al. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. *Eur J Pharmacol*. 2007;28;557(2–3):221–9.
13. Alam W, Khan H, Shah MA, Cauli O, Saso L. Kaempferol as a Dietary Anti-Inflammatory Agent: Current Therapeutic Standing. *Molecules*. 2020;7;25(18):4073. doi: 10.3390/molecules25184073.
14. Cannataro R, Fazio A, Torre C La, Caroleo MC, Cione E. Polyphenols in the mediterranean diet: From dietary sources to microrna modulation. *Antioxidants*. MDPI. 2021;23;10(2):328. doi: 10.3390/antiox10020328.
15. Ferrer MD, Sureda A, Mestre A, Tur JA, Pons A. The double edge of reactive oxygen species as damaging and signaling molecules in HL60 cell culture. *Cell Physiol Biochem*. 2010;25(2–3):241–52. doi: 10.1159/000276558.
16. Almatroodi SA, Almatroudi A, Alsahli MA, Khan AA, Rahmani AH. Thymoquinone, an Active Compound of *Nigella sativa*: Role in Prevention and Treatment of Cancer. *Curr Pharm Biotechnol*. 2020;21(11):1028–1041. doi: 10.2174/1389201021666200416092743.
17. Alzohairy MA, Khan AA, Alsahli MA, Almatroodi SA, Rahmani AH. Protective effects of thymoquinone, an active compound of *nigella sativa*, on rats with benzo(A)pyrene-induced lung injury through regulation of oxidative stress and inflammation. *Molecules*. 2021; 27;26(11):3218. doi: 10.3390/molecules26113218.
18. Procházková D, Boušová I, Wilhelmová N. Antioxidant and prooxidant properties of flavonoids. *Fitoterapia*. 2011;82(4):513–23. doi: 10.1016/j.fitote.2011.01.018.
19. Wang Y, Chen C, Li Y, Li R, Wang J, Wu C, et al. Kaempferol inhibits oxidative stress and reduces macrophage pyroptosis by activating the NRF2 signaling pathway. *PLoS One*. 2025;20(6): e0325189. <https://doi.org/10.1371/journal.pone.0325189>.
20. Biswas SK. Does the Interdependence between Oxidative Stress and Inflammation Explain the Antioxidant Paradox? *Oxidative Medicine and Cellular Longevity*. Hindawi Publishing Corporation. 2016;2016:5698931. doi: 10.1155/2016/5698931.
21. Hofer S, Geisler S, Lisandrelli R, Ngoc HN, Ganzera M, Schennach H, et al. Pharmacological targets of kaempferol within inflammatory pathways—a hint towards the central role of tryptophan metabolism. *Antioxidants*. 2020; 21;9(2):180. doi: 10.3390/antiox9020180.
22. Herrera TES, Tello IPS, Mustafa MA, Jamil NY, Alaraj M, Atiyah Altameem KK, et al. Kaempferol: Unveiling its anti-inflammatory properties for therapeutic innovation. *Cytokine*. 2025;186:156846. doi: 10.1016/j.cyto.2024.156846.
23. Alkhalidy H, Moore W, Wang Y, Luo J, McMillan RP, Zhen W, et al. The flavonoid kaempferol ameliorates streptozotocin-induced diabetes by suppressing hepatic glucose production. *Molecules*. 2018;13;23(9):2338. doi: 10.3390/molecules23092338.
24. Muriel P. Liver pathophysiology : therapies and antioxidants. Chapter 1 - The Liver: General Aspects and Epidemiology, Academic Press is an imprint of Elsevier; 2017. p.3–22. <https://doi.org/10.1016/B978-0-12-804274-8.00001-1>.
25. Almatroodi SA, Alsahli MA, Alharbi HM, Khan AA, Rahmani AH. Epigallocatechin-3-gallate (EGCG), an active constituent of green tea: Implications in the prevention of liver injury induced by diethylnitrosamine (DEN) in rats. *Appl Sci*. 2019;1;9(22):4821. <https://doi.org/10.3390/app9224821>.
26. Rahmani AH, Almatroudi A, Allemailem KS, Alharbi HOA, Babiker AY, Althwab SA, Alsuhaymi N, Alsugoor MH, Khan AA, Al-Megrin WAI. Oleuropein, a phenolic component of *Olea europaea* L. ameliorates CCl4-induced liver injury in rats through the regulation of oxidative

- stress and inflammation. *Eur Rev Med Pharmacol Sci.* 2024;28(4):1259–1271. doi: 10.26355/eurrev_202402_35447.
27. Przemyslaw N, Krajewska A, Oniszczyk T, Polak B, Oniszczyk A. Hepatoprotective Effect of Kaempferol — A Review. *Molecule.* 2025; 30(9): 1913. <https://doi.org/10.3390/molecules30091913>.
28. Li Q, Hu X, Xuan Y, Ying J, Fei Y, Rong J, et al. Kaempferol protects ethanol-induced gastric ulcers in mice via pro-inflammatory cytokines and NO. *Acta Biochim Biophys Sin (Shanghai).* 2018; 1;50(3):246–53. doi: 10.1093/abbs/gmy002. PMID: 29415150.
29. Alrumaihi F, Almatroodi SA, Alharbi HOA, Alwanian WM, Alharbi FA, Almatroudi A, et al. Pharmacological Potential of Kaempferol, a Flavonoid in the Management of Pathogenesis via Modulation of Inflammation and Other Biological Activities. *Molecules.* Multidisciplinary Digital Publishing Institute (MDPI); 2024;26;29(9):2007. doi: 10.3390/molecules29092007.
30. Sharma D, Kumar Tekade R, Kalia K. Kaempferol in ameliorating diabetes-induced fibrosis and renal damage: An in vitro and in vivo study in diabetic nephropathy mice model. *Phytomedicine.* 2020;1;76:153235. doi: 10.1016/j.phymed.2020.153235.
31. Babaei P, Eyvani K, Kouhestani S. Sex-Independent Cognition Improvement in Response to Kaempferol in the Model of Sporadic Alzheimer's Disease. *Neurochem Res.* 202;1;46(6):1480–1486. doi: 10.1007/s11064-021-03289-y.
32. Pate KM, Rogers M, Reed JW, van der Munnik N, Vance SZ, Moss MA. Anthoxanthin Polyphenols Attenuate A β Oligomer-induced Neuronal Responses Associated with Alzheimer's Disease. *CNS Neurosci Ther.* 2017;1;23(2):135–44. doi: 10.1111/cns.12659.
33. Beg T, Jyoti S, Naz F, Rahul, Ali F, Ali SK, et al. Protective Effect of Kaempferol on the Transgenic Drosophila Model of Alzheimer's Disease. *CNS Neurol Disord - Drug Targets.* 2018;10;17(6):421–9. doi: 10.2174/1871527317666180508123050.
34. Feng W, Kita D, Peaucelle A, Cartwright HN, Doan V, Duan Q, et al. The FERONIA Receptor Kinase Maintains Cell-Wall Integrity during Salt Stress through Ca²⁺ Signaling. *Curr Biol.* 2018;28(5):666–75e5. doi: 10.1016/j.cub.2018.01.023.
35. Hasanpourghadi M, Looi CY, Pandurangan AK, Sethi G, Wong WF, Mustafa MR. Phytometabolites Targeting the Warburg Effect in Cancer Cells: A Mechanistic Review. *Curr Drug Targets.* 2017;18(9):1086–1094. doi: 10.2174/1389450117666160401124842.
36. Turek M, Krzyczmonik M, Balczewski P. New Hopes in Cancer Battle - A Review of New Molecules and Treatment Strategies. *Med Chem (Los Angeles).* 2016;17;12(8):700–19. doi: 10.2174/1573406412666160502153700.
37. Almatroodi SA, Almatroudi A, Alsahli MA, Khan AA, Rahmani AH. Thymoquinone, an Active Compound of Nigella sativa: Role in Prevention and Treatment of Cancer. *Curr Pharm Biotechnol.* 2020;16;21(11):1028–41. doi: 10.2174/1389201021666200416092743.
38. Almatroudi A, Allemailem KS, Alwanian WM, Alharbi BF, Alrumaihi F, Khan AA, et al. Effects and Mechanisms of Kaempferol in the Management of Cancers through Modulation of Inflammation and Signal Transduction Pathways. Vol. 24, *International Journal of Molecular Sciences.* Multidisciplinary Digital Publishing Institute (MDPI). 2023;11;24(10):8630. doi: 10.3390/ijms24108630.
39. Rahmani AH, Almatroudi A, Allemailem KS, Alwanian WM, Alharbi BF, Alrumaihi F, et al. Myricetin: A Significant Emphasis on Its Anticancer Potential via the Modulation of Inflammation and Signal Transduction Pathways. Vol. 24, *International Journal of Molecular Sciences.* Multidisciplinary Digital Publishing Institute (MDPI). 2023 :2;24(11):9665. doi: 10.3390/ijms24119665.
40. Gonelimali FD, Lin J, Miao W, Xuan J, Charles F, Chen M, et al. Antimicrobial properties and mechanism of action of some plant extracts against food pathogens and spoilage microorganisms. *Front Microbiol.* 2018 :9;1639. doi: 10.3389/fmicb.2018.01639.
41. Liu J, Wen J, Lu J, Zhou H, Wang G. Kaempferol and Kaempferin Alleviate MRSA Virulence by Suppressing β -Lactamase and Inflammation. *Molecules,* 30(20), 4132. <https://doi.org/10.3390/molecules30204132>.

42. Patil SV, Borase HP, Patil CD. et al. Biosynthesis of Silver Nanoparticles Using Latex from Few Euphorbian Plants and Their Antimicrobial Potential. *Appl Biochem Biotechnol*;2012; 167, 776-790. <https://doi.org/10.1007/s12010-012-9710-z>
43. Namini MS, Mohandesnezhad S, Mohandesnezhad S, Mansouri V, Tayebi L. Enhancing bone regeneration using kaempferol as an osteoprotective compound : signaling mechanisms, delivery strategies, and potential applications. *J Biol Eng.* 19,74 (2025). <https://doi.org/10.1186/s13036-025-00545-5> 2025;5.
44. Silva dos Santos J, Gonçalves Cirino JP, de Oliveira Carvalho P, Ortega MM. The Pharmacological Action of Kaempferol in Central Nervous System Diseases: A Review. *Front Pharmacol.* 2021;11.
45. Zhou H, Xu M, Guo W, Yao Z, Du X, Chen L, et al. The Antibacterial Activity of Kaempferol Combined with Colistin against Colistin-Resistant Gram-Negative Bacteria. *Microbiol Spectr.* 2022; 21;10(6).
46. Subbaraj GK, Masoodi T, Yasam SK, Chandrashekar K, Kulanthaivel L, Shaik NA, et al. Anti-angiogenic effect of nano-formulated water soluble kaempferol and combretastatin in an in vivo chick chorioallantoic membrane model and HUVEC cells. *Biomed Pharmacother.* 2023; 1;163.
47. Aghazadeh T, Bakhtiari N, Rad IA, Ramezani F. Formulation of kaempferol in nanostructured lipid carriers (Nlcs): A delivery platform to sensitization of mda-mb468 breast cancer cells to paclitaxel. *Biointerface Res Appl Chem.* 2021;11(6):14601–14591.
48. Hussain MS, Altamimi ASA, Afzal M, Almalki WH, Kazmi I, Alzarea SI, et al. Kaempferol: Paving the path for advanced treatments in aging-related diseases. *Experimental Gerontology*; 2024;188:112389. doi: 10.1016/j.exger.2024.112389.

CHAPTER 7

CATECHIN AND ITS DERIVATIVES

Sinem AYDIN¹

Catechins are categorized under flavanols (1). The catechin term originally comes from the word ‘catechu’, which is the boiled extract (tannic juices) of the *Mimosa catechu* (2). Fruits and vegetables have catechins at high amounts (1).

They are an important component of tea, making up approximately one-third of the dry weight of tea (3). Non-esterified catechins are catechin, gallic catechin, epicatechin, epigallocatechin and esterified catechins are epigallocatechin gallate, epicatechin gallate, gallic catechin gallate and catechin gallate (1). Catechin derivatives are given in **Fig 1**.

Catechins react easily with oxygen in the air by chemical and enzymatic ways to form condensed tannins (proanthocyanidins). The astringency or bitterness of these compounds is determined by their molecular weights. While proanthocyanidins with sixty-eight monomers create an astringent taste, those with thirty-five monomers create a bitter taste. While short chain molecules are colorless, their color changes from yellow to brown as the degree of polymerization increases (4).

Catechin and type B procyanidin found in legumes. Catechin, epicatechin and epicatechin gallate are found in fruit wine (1). *Camellia sinensis* teas were also studied for their catechins (5).

¹ Doç. Dr., Giresun University, Faculty of Science and Arts, Department of Biology, sinem.aydin@giresun.edu.tr, ORCID iD: 0000-0002-0484-7191

in green tea has been shown in various studies to prevent obesity by reducing adipocyte proliferation, lipogenesis, fat absorption, and food intake (22).

L. Antidiabetic Effect

Tea catechins increase insulin activity by protecting pancreatic β cells. Tea polyphenols protect against insulin resistance, the primary cause of Type II diabetes, by improving insulin sensitivity. Jing et al. (19) revealed that tea has a preventing impact towards diabetes, with a modest reduction in Type II diabetes in individuals who drank more than four cups of green tea every day.

A study by Iso et al. (19) declared that consuming more than six cups of green tea per day was inversely linked with incidence of diabetes, while the same effect was not observed for black and oolong teas. Tea exerts a preventive influence to diabetes and its complications by increasing insulin effect, improving insulin resistance, activating insulin signaling and defending β cells. Researchs from different countries demonstrated that tea increases fasting insulin levels and that consuming more than four cups every day reduces the risk of Type II diabetes (19).

REFERENCES

1. Gadkari PV, Balaraman M. Catechins: sources, extraction and encapsulation: a review. *Food and Bioproducts Processing*. 2015; 93: 122-138. doi.org/10.1016/j.fbp.2013.12.004
2. What are Catechins? [Online] Available from: <https://www.news-medical.net/life-sciences/What-are-Catechins.aspx> (Accessed: 15 May 2025).
3. Yalçın AS, Yılmaz AM, Altundağ EM, et al. Anti-cancer effects of curcumin, quercetin and tea catechins. *Marmara Pharmaceutical Journal*. 2017; 21: 19-29. doi.org/10.12991/marupj.259877.
4. Gökçen İS, Keskin N, Kunter B, et al. Üzüm fitokimyasalları ve Türkiye’de yetiştirilen üzüm çeşitleri üzerindeki araştırmalar. *Turkish Journal of Forest Science*. 2017; 1(1):93-111. doi.org/10.32328/turkjforsci.285695.
5. Pedro AC, Maciel GM, Ribeiro VR, et al. Fundamental and applied aspects of catechins from different sources: a review. *International Journal of Food Science and Technology*. 2019; 55(2): 429-442. doi.org/10.1111/ijfs.14371.
6. Bae J, Kim N, Shin Y, et al. Activity of catechins and their applications. *Biomedical Dermatology*. 2020; 4 (8): 1-10. doi.org/10.1186/s41702-020-0057-8.
7. Coşarcă S, Tanase C, Muntean DL. Therapeutic aspects of catechin and its derivatives – an update. *Acta Biologica Marisensis*. 2019; 2(1): 21-29. Doi: 10.2478/abmj-2019-0003.
8. Capasso L, De Masi L, Sirignano C, et al. Epigallocatechin gallate (EGCG): pharmacological properties, biological activities and therapeutic potential. *Molecules*. 2025; 30(3): 654-688. doi: 10.3390/molecules30030654.
9. Sheng YY, Xiang J, Lu JL, et al. Protective effects of gallic catechin gallate against ultraviolet B induced skin damages in hairless mice. *Scientific Reports*. 2022; 12, 1310-1321. doi.org/10.1038/s41598-022-05305-9.
10. Çayda Bulunan Kateşinler [Online] Available from: <https://wikifarmer.com/library/tr/article/%C3%A7ayda-bulunan-kate%C5%9Finler> (Accessed: 05 June 2025).
11. Zengin A. 2007. Sıçan frenik sinir-hemidiyafram preparatına antioksidan resveratrol, kateşin

- ve epikateşinin etkileri. Master Thesis, Istanbul University Health Sciences Institute, Department of Neuroscience.
12. Sarıca Ş, Karataş Ü, Diktaş M. Çay (*Camellia sinensis*); içeriği, metabolizma ve sağlık üzerine etkileri, antioksidan aktivitesi ve etlik piliç karma yemlerinde kullanımı. GaziOsmanpaşa Üniversitesi Ziraat Fakültesi Dergisi. 2008; 25(2), 79-85.
 13. Ünal A. 2016. Deneysel olarak oluşturulmuş meme tümöründe epigallokateşin-3-gallat'ın olası tedavi edici rolü. Master Thesis, Trakya University Health Sciences Institute Department of Medical Biochemistry.
 14. Mita SR, Husni P, Putriana NA, et al. A recent update on the potential use of catechins in cosmetics. *Cosmetics*. 2024; 11(1):23-39. doi.org/10.3390/cosmetics11010023.
 15. Bernatoniene J, Kopustinskiene DM. The role of catechins in cellular responses to oxidative stress. *Molecules*. 2018; 23(4):965-976. Doi: 10.3390/molecules23040965.
 16. Akbulut A. 2019. Siyah, yeşil ve beyaz çayların kalite kriterleri, mineral içerikleri, antioksidan ve antimikrobiyal aktivite yönünden karşılaştırılması. Master Thesis, Ordu University, Institute of Science, Department of Field Crops.
 17. Yılmaz Y. Novel uses of catechins in foods. *Trends in Food Science & Technology*. 2006; 17(2): 64- 71. doi.org/10.1016/j.tifs.2005.10.005.
 18. Reygaert WC. Green tea catechins: their use in treating and preventing infectious diseases. *Biomed Research International*. 2018; 1-9. doi: 10.1155/2018/9105261.
 19. Akbay GD. Çayın sağlığa etkilerinin belirlenmesi. *Bayburt Üniversitesi Fen Bilimleri Dergisi*. 2021; 4(1): 91-97.
 20. Kontaş Aşkar T, Aşkar Ş, Er E, et al. Anti-aging etkili nutrasötikler. Hişmioğulları ŞE, editör. *Nutrasötikler*. 1. Baskı. Ankara: Türkiye Klinikleri; 2021. p.50-4.
 21. Yapar EA, Tanrıverdi ST. Yaşlanma karşıtı kozmetik yaklaşımlar ve ürün bileşenleri. *Balıkesir Sağlık Bilimleri Dergisi*. 2016; 5(2): 99-109. Doi: 10.5505/bsbd.2016.04696.
 22. Yılmaz FT, Demirel G, Kumsar AK. Çay, obezite ve kadın. *Journal Of Contemporary Medicine*. 2016; 6(2): 137-146. doi.org/10.16899/ctd.90351.

CHAPTER 8

FLAVANOLS

Funda DEMİRTAŞ KORKMAZ¹

Introduction

Flavanols represent a significant subgroup of flavonoids, which are naturally occurring polyphenolic molecules broadly found throughout the plant kingdom (1,2). They are found abundantly in foods such as cocoa, tea, apples, grapes, and various berries (3). In recent decades, flavanols have attracted significant scientific interest because of their possible positive impacts on human health, especially in relation to cardiovascular, neurological, and metabolic systems (4–7). Their biological activity stems not only from their direct antioxidant properties but also from their ability to modulate cellular signaling pathways, gene expression, and enzymatic activity (8–10).

The health-promoting potential of flavanols is closely linked to their structure and metabolism. While early research emphasized their role as antioxidants that neutralize reactive oxygen species, recent findings suggest that their physiological effects are more complex and involve interactions with endothelial cells, the gut microbiota, and the immune system (11–13). This chapter reviews the essential chemical structures, sources, bioavailability, biological activities, and clinical evidence surrounding flavanols, along with an outlook on future research directions.

¹ Asst. Prof. Giresun University, Faculty of Medicine, Department of Medicinal Biology, demirtas.korkmaz@giresun.edu.tr, ORCID iD: 0000-0003-3978-9427

REFERENCES

1. Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources and bio-availability. *Am J Clin Nutr.* 2004;79(5):727–47.
2. Luo Y, Jian Y, Liu Y, Jiang S, Muhammad D, Wang W. Flavanols from Nature: A Phytochemistry and Biological Activity Review. *Molecules.* 2022;27(3):719.
3. Fraga CG, Oteiza PI. Dietary flavonoids: Role of (–)-epicatechin and related procyanidins in cell signaling. *Free Radic Biol Med.* 2011;51(4):813–23.
4. Garcia JP, Santana A, Baruqui DL, Suraci N. The cardiovascular effects of chocolate. Vol. 19, *Reviews in Cardiovascular Medicine.* 2018;19:123–7.
5. Vlachojannis J, Erne P, Zimmermann B, Chrubasik-Hausmann S. The Impact of Cocoa Flavanols on Cardiovascular Health. *Phyther Res.* 2016;30(10):1641–57.
6. Mahdipour R, Ebrahimzadeh-Bideskan A, Hosseini M, Shahba S, Lombardi G, Malvandi AM, et al. The benefits of grape seed extract in neurological disorders and brain aging. *Nutritional Neuroscience.* 2023;26(5):369–83.
7. Salaish Kumar S, Mhd Jalil AM, Hussin N, Mat Daud ZA, Ismail A. Effects of flavanols and procyanidins-rich cocoa consumption on metabolic syndrome: an update review (2013–2023). Vol. 88, *Bioscience, Biotechnology and Biochemistry.* 2024;88(4):352–360.
8. Wang J, Yu Z, Wu W, He S, Xie B, Wu M, et al. Molecular mechanism of epicatechin gallate binding with carboxymethyl β -glucan and its effect on antibacterial activity. *Carbohydr Polym.* 2022;298:120105.
9. Anitha S, Saranya V, Shankar R, Sasirekha V. Structural exploration of interactions of (+) catechin and (–) epicatechin with bovine serum albumin: Insights from molecular dynamics and spectroscopic methods. *J Mol Liq.* 2022;348:118026.
10. Lee K, Bharadwaj S, Yadava U, Kang S. Computational and In Vitro Investigation of (–)-Epicatechin and Proanthocyanidin B2 as Inhibitors of Human Matrix Metalloproteinase 1. *Biomolecules.* 2020;10(10):1379.
11. Ramirez-Sanchez I, Nogueira L, Moreno A, Murphy A, Taub P, Perkins G, et al. Stimulatory effects of the flavanol (–)-epicatechin on cardiac angiogenesis: Additive effects with exercise. *J Cardiovasc Pharmacol.* 2012;60(5):429–38.
12. Sorrenti V, Ali S, Mancin L, Davinelli S, Paoli A, Scapagnini G. Cocoa Polyphenols and Gut Microbiota Interplay: Bioavailability, Prebiotic Effect, and Impact on Human Health. *Nutrients.* 2020;12(7):1908.
13. Azam S, Jakaria M, Kim IS, Kim J, Haque ME, Choi DK. Regulation of Toll-Like Receptor (TLR) Signaling Pathway by Polyphenols in the Treatment of Age-Linked Neurodegenerative Diseases: Focus on TLR4 Signaling. *Front Immunol.* 2019;10(MAY):1000.
14. Flores MEJ. Cocoa flavanols: Natural agents with attenuating effects on metabolic syndrome risk factors. *Nutrients.* 2019;11(4):751.
15. Amoah I, Lim JJ, Osei EO, Arthur M, Tawiah P, Oduro IN, et al. Effect of Cocoa Beverage and Dark Chocolate Consumption on Blood Pressure in Those with Normal and Elevated Blood Pressure: A Systematic Review and Meta-Analysis. *Foods.* 2022; 11(13): 1962.
16. Chen TS, Liao WY, Huang CW, Chang CH. Adipose-Derived Stem Cells Preincubated with Green Tea EGCG Enhance Pancreatic Tissue Regeneration in Rats with Type 1 Diabetes through ROS/Sirt1 Signaling Regulation. *Int J Mol Sci.* 2022 Mar 15;23(6):3165.
17. Wang YQ, Li QS, Zheng XQ, Lu JL, Liang YR. Antiviral Effects of Green Tea EGCG and Its Potential Application against COVID-19. *Molecules.* 2021;26(13):3962.
18. Tang G, Xu Y, Zhang C, Wang N, Li H, Feng Y. Green tea and epigallocatechin gallate (Egcg) for the management of nonalcoholic fatty liver diseases (nafld): Insights into the role of oxidative stress and antioxidant mechanism. *Antioxidants.* 2021;10(7):1076.
19. Hackman RM, Polagruto JA, Zhu QY, Sun B, Fujii H, Keen CL. Flavanols: digestion, absorption and bioactivity. *Phytochem Rev.* 2007;7(1):195–208.

20. Taubert D, Roesen R, Lehmann C, Jung N, Schömig E. Effects of Low Habitual Cocoa Intake on Blood Pressure and Bioactive Nitric Oxide. *JAMA*. 2007;298(1):49.
21. Manach C, Williamson G, Morand C, Scalbert A, Rémésy C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am J Clin Nutr*. 2005;81(1):230S-242S.
22. Mena P, Bresciani L, Brindani N, Ludwig IA, Pereira-Caro G, Angelino D, et al. Phenyl- γ -valerolactones and phenylvaleric acids, the main colonic metabolites of flavan-3-ols: synthesis, analysis, bioavailability, and bioactivity. *Nat Prod Rep*. 2019;36(5):714–52.
23. Fernandes I, Nave F, Gonçalves R, de Freitas V, Mateus N. On the bioavailability of flavanols and anthocyanins: Flavanol–anthocyanin dimers. *Food Chem*. 2012;135(2):812–8.
24. Di Lorenzo C, Colombo F, Biella S, Stockley C, Restani P. Polyphenols and Human Health: The Role of Bioavailability. *Nutrients*. 2021;13(1):273.
25. Payne A, Taka E, Adinew GM, Soliman KFA. Molecular Mechanisms of the Anti-Inflammatory Effects of Epigallocatechin 3-Gallate (EGCG) in LPS-Activated BV-2 Microglia Cells. *Brain Sci*. 2023;13(4):632.
26. Cheng Z, Zhang Z, Han Y, Wang J, Wang Y, Chen X, et al. A review on anti-cancer effect of green tea catechins. *Journal of Functional Foods*. 2020;74:104172.
27. Perron NR, Brumaghim JL. A review of the antioxidant mechanisms of polyphenol compounds related to iron binding. *Cell Biochem Biophys*. 2009;53(2):75–100.
28. Su J, Liao T, Ren Z, Kuang Y, Yu W, Qiao Q, et al. Polydopamine nanoparticles coated with a metal-polyphenol network for enhanced photothermal/chemodynamic cancer combination therapy. *Int J Biol Macromol*. 2023;238:124088.
29. Han J, Wang M, Jing X, Shi H, Ren M, Lou H. (-)-Epigallocatechin gallate protects against cerebral ischemia-induced oxidative stress via Nrf2/ARE signaling. *Neurochem Res*. 2014;39(7):1292–9.
30. Sun W, Liu X, Zhang H, Song Y, Li T, Liu X, et al. Epigallocatechin gallate upregulates NRF2 to prevent diabetic nephropathy via disabling KEAP1. *Free Radic Biol Med*. 2017;108:840–57.
31. Heiss C, Keen CL, Kelm M. Flavanols and cardiovascular disease prevention. *European Heart Journal*. 2010; 31(21):2583-2592.
32. Steffen Y, Gruber C, Schewe T, Sies H. Mono-O-methylated flavanols and other flavonoids as inhibitors of endothelial NADPH oxidase. *Arch Biochem Biophys*. 2008;469(2):209–19.
33. Actis-Goretta L, Ottaviani JI, Keen CL, Fraga CG. Inhibition of angiotensin converting enzyme (ACE) activity by flavan-3-ols and procyanidins. *FEBS Lett*. 2003;555(3):597–600.
34. Wolfram S. Effects of green tea and egcg on cardiovascular and metabolic health. *J Am Coll Nutr*. 2007;26(4):373S-388S.
35. Ottaviani JI, Heiss C, Spencer JPE, Kelm M, Schroeter H. Recommending flavanols and procyanidins for cardiovascular health: Revisited. *Mol Aspects Med*. 2018;61:63–75.
36. Sharifi-Rad M, Pezzani R, Redaelli M, Zorzan M, Imran M, Ahmed Khalil A, et al. Pre-clinical Activities of Epigallocatechin Gallate in Signaling Pathways in Cancer. *Molecules*. 2020;25(3):467.
37. Xu T, Liu R, Zhu H, Zhou Y, Pei T, Yang Z. The Inhibition of LPS-Induced Oxidative Stress and Inflammatory Responses Is Associated with the Protective Effect of (-)-Epigallocatechin-3-Gallate on Bovine Hepatocytes and Murine Liver. *Antioxidants*. 2022;11(5):914.
38. Kim JS, Kim JM, O JJ, Jeon BS. Inhibition of inducible nitric oxide synthase expression and cell death by (-)-epigallocatechin-3-gallate, a green tea catechin, in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *J Clin Neurosci*. 2010;17(9):1165–8.
39. Yoshida S, Inaba H, Nomura R, Nakano K, Matsumoto-Nakano M. Green tea catechins inhibit *Porphyromonas gingivalis* LPS-induced inflammatory responses in human gingival epithelial cells. *J Oral Biosci*. 2022;64(3):352–8.

40. Han Y, Pei D, Li W, Luo B, Jiang Q. Epigallocatechin gallate attenuates tumor necrosis factor (TNF)- α -induced inhibition of osteoblastic differentiation by up-regulating lncRNA TUG1 in osteoporosis. *Bioengineered*. 2022;13(4):8950–61.
41. Ranjith-Kumar CT, Lai Y, Sarisky RT, Kao CC. Green tea catechin, epigallocatechin gallate, suppresses signaling by the dsRNA innate immune receptor RIG-I. Cheriya V, editor. *PLoS One*. 2010;5(9):1–11.
42. Narita K, Hisamoto M, Okuda T, Takeda S. Differential neuroprotective activity of two different grape seed extracts. Ikeda K, editor. *PLoS One*. 2011;6(1):e14575.
43. Ratnawati R, Arofah AN, Novitasari A, Utami SD, Hariningsih MA, Rahayu M, et al. Catechins decrease neurological severity score through apoptosis and neurotropic factor pathway in rat traumatic brain injury. *Universa Med*. 2017;36(2):110–22.
44. Pervin M, Unno K, Takagaki A, Isemura M, Nakamura Y. Function of Green Tea Catechins in the Brain: Epigallocatechin Gallate and its Metabolites. *Int J Mol Sci*. 2019;20(15):3630.
45. Özduran G, Becer E, Vatansever HS. The Role and Mechanisms of Action of Catechins in Neurodegenerative Diseases. Vol. 42, *Journal of the American Nutrition Association*. 2023;42(1):67–74.
46. Fernandes L, Cardim-Pires TR, Foguel D, Palhano FL. Green Tea Polyphenol Epigallocatechin-Gallate in Amyloid Aggregation and Neurodegenerative Diseases. *Frontiers in Neuroscience*. 2021;15: 718188.
47. Ding ML, Ma H, Man YG, Lv HY. Protective effects of a green tea polyphenol, epigallocatechin-3-gallate, against sevoflurane-induced neuronal apoptosis involve regulation of CREB/BDNF/TrkB and PI3K/Akt/mTOR signalling pathways in neonatal mice. *Can J Physiol Pharmacol*. 2017;95(12):1396–405.
48. Almaguer G, Almaguer-Vargas G, Molina-Trinidad EM, Becerril-Flores MA, Montejano B, Madrigal-Santillan E, et al. Antitumor Effect of Epigallocatechin Gallate and Vincristine in Mice with L5178Y Lymphoma. *Plants*. 2023;12(21):3757.
49. Zhang D, Al-Hendy M, Richard-Davis G, Montgomery-Rice V, Rajaratnam V, Al-Hendy A. Antiproliferative and proapoptotic effects of epigallocatechin gallate on human leiomyoma cells. *Fertil Steril*. 2010;94(5):1887–93.
50. Gu JW, Makey KL, Tucker KB, Chinchar E, Mao X, Pei I, et al. EGCG, a major green tea catechin suppresses breast tumor angiogenesis and growth via inhibiting the activation of HIF-1 α and NF κ B, and VEGF expression. *Vasc Cell*. 2013;5(1):9.
51. Shimizu M, Shirakami Y, Sakai H, Yasuda Y, Kubota M, Adachi S, et al. (-)-Epigallocatechin gallate inhibits growth and activation of the VEGF/VEGFR axis in human colorectal cancer cells. *Chem Biol Interact*. 2010;185(3):247–52.
52. García-Cordero J, Martínez A, Blanco-Valverde C, Pino A, Puertas-Martín V, San Román R, et al. Regular Consumption of Cocoa and Red Berries as a Strategy to Improve Cardiovascular Biomarkers via Modulation of Microbiota Metabolism in Healthy Aging Adults. *Nutrients*. 2023;15(10).
53. Goya L, Martín M, Sarriá B, Ramos S, Mateos R, Bravo L. Effect of Cocoa and Its Flavonoids on Biomarkers of Inflammation: Studies of Cell Culture, Animals and Humans. *Nutrients*. 2016;8(4):212.
54. Li F, Gao C, Yan P, Zhang M, Wang Y, Hu Y, et al. EGCG reduces obesity and white adipose tissue gain partly through AMPK activation in mice. *Front Pharmacol*. 2018;9:389872.
55. Sampath C, Rashid MR, Sang S, Ahmedna M. Green tea epigallocatechin 3-gallate alleviates hyperglycemia and reduces advanced glycation end products via nrf2 pathway in mice with high fat diet-induced obesity. *Biomed Pharmacother*. 2017;87:73–81.
56. Heiss C. Vascular Effects of Cocoa Rich in Flavan-3-ols. *JAMA J Am Med Assoc*. 2003;290(8):1030–1.

57. Grosso G, Godos J, Currenti W, Micek A, Falzone L, Libra M, et al. The Effect of Dietary Polyphenols on Vascular Health and Hypertension: Current Evidence and Mechanisms of Action. *Nutrients*. 2022;14(3):545.
58. Akhlaghi M, Ghobadi S, Mohammad Hosseini M, Gholami Z, Mohammadian F. Flavanols are potential anti-obesity agents, a systematic review and meta-analysis of controlled clinical trials. *Nutr Metab Cardiovasc Dis*. 2018;28(7):675–90.
59. Bazzyar H, Hosseini SA, Saradar S, Mombaini D, Allivand M, Labibzadeh M, et al. Effects of epigallocatechin-3-gallate of *Camellia sinensis* leaves on blood pressure, lipid profile, atherogenic index of plasma and some inflammatory and antioxidant markers in type 2 diabetes mellitus patients: a clinical trial. *J Complement Integr Med*. 2021;18(2):405–11.
60. Gratton G, Weaver SR, Burley C V., Low KA, Maclin EL, Johns PW, et al. Dietary flavanols improve cerebral cortical oxygenation and cognition in healthy adults. *Sci Rep*. 2020;10(1):19409.
61. Vyas CM, Manson JAE, Sesso HD, Rist PM, Weinberg A, Kim E, et al. Effect of cocoa extract supplementation on cognitive function: results from the clinic subcohort of the COSMOS trial. *Am J Clin Nutr*. 2024;119(1).
62. Brickman AM, Yeung LK, Alschuler DM, Ottaviani JI, Kuhnle GGC, Sloan RP, et al. Dietary flavanols restore hippocampal-dependent memory in older adults with lower diet quality and lower habitual flavanol consumption. *Proc Natl Acad Sci*. 2023;120(23).
63. Akyürek EG, Altınok A, Karabay A. Concurrent consumption of cocoa flavanols and caffeine does not acutely modulate working memory and attention. *Eur J Nutr*. 2025;64(1):1–17.

CHAPTER 9

RESVERATROL

Zeliha KAYA¹

1. Introduction

Free radicals are harmful compounds that can lead to oxidative damage and cause chronic degenerative diseases. Substances that prevent the formation of these radicals or neutralize them are referred to as “antioxidants.” Resveratrol, one of these antioxidants, is a phytoalexin found in more than 70 plant species, particularly in grapes and grape-derived products (1). On an industrial scale, it is predominantly obtained from the roots of *Polygonum cuspidatum* (Japanese knotweed)(2). Owing to its beneficial effects on human health, resveratrol has attracted significant global attention in recent years. Numerous experimental systems have been used to thoroughly examine its *in vitro* and *in vivo* biological activity (3). Resveratrol has been reported to exhibit numerous therapeutic properties, including anti-inflammatory, antioxidant, antidiabetic, antiaggregatory, antihyperlipidemic, immunomodulatory, cardioprotective, vasorelaxant, and neuroprotective effects (4,5). However, clinical outcomes remain inconsistent due to factors such as low bioavailability, rapid metabolism, and dosage variations (6–8). These pharmacokinetic limitations have recently prompted a shift toward innovative approaches, including nanoformulations, polymeric and bioceramic hybrid systems, and green extraction technologies (9,10).

¹ Assist. Prof., Giresun University, Faculty of Tourism, zeliha.kaya@giresun.edu.tr ,
ORCID iD: 0000-0002-3285-9659

REFERENCES

1. Sarman E, Gülle K. Resveratrol Güçlü Bir Antioksidan Mı? Bingöl Üniversitesi Sağlık Derg. 2021 June 30;2(1):57–63.
2. Tian B, Liu J. Resveratrol: a review of plant sources, synthesis, stability, modification and food application. J Sci Food Agric. 2020 Mar 15;100(4):1392–404.
3. Göldağ R, Doğan M. Resveratrolün Antioksidan Etkisi, Farmakolojik Özellikleri ve Bazı Sağlık Faydaları. Int Conf Recent Innov Results Eng Technol. 2023 Aug 29;40–5.
4. Öztürk E, Arslan AKK, Yerer MB, Bishayee A. Resveratrol and diabetes: A critical review of clinical studies. Biomed Pharmacother. 2017 Nov;95:230–4.
5. Singh AP, Singh R, Verma SS, Rai V, Kaschula CH, Maiti P, et al. Health benefits of resveratrol: Evidence from clinical studies. Med Res Rev. 2019 Sept;39(5):1851–91.
6. Berman AY, Motechin RA, Wiesenfeld MY, Holz MK. The therapeutic potential of resveratrol: a review of clinical trials. Npj Precis Oncol. 2017 Sept 25;1(1):35.
7. Springer M, Moco S. Resveratrol and Its Human Metabolites—Effects on Metabolic Health and Obesity. Nutrients. 2019 Jan 11;11(1):143.
8. Yu X, Jia Y, Ren F. Multidimensional biological activities of resveratrol and its prospects and challenges in the health field. Front Nutr. 2024 June 12;11:1408651.
9. Ben Dassi R, Ibidhi S, Jemai H, Cherif A, Driouich Chaouachi R. Resveratrol: Challenges and prospects in extraction and hybridization with nanoparticles, polymers, and bio-ceramics. Phytother Res. 2024 Nov;38(11):5309–22.
10. Sharifi-Rad J, Quispe C, Durazzo A, Lucarini M, Souto EB, Santini A, et al. Resveratrol' biotechnological applications: Enlightening its antimicrobial and antioxidant properties. J Herb Med. 2022 Mar;32:100550.
11. Meng T, Xiao D, Muhammed A, Deng J, Chen L, He J. Anti-Inflammatory Action and Mechanisms of Resveratrol. Molecules. 2021 Jan 5;26(1):229.
12. Ren Z, Zheng S, Sun Z, Luo Y, Wang Y, Yi P, et al. Resveratrol: Molecular Mechanisms, Health Benefits, and Potential Adverse Effects. MedComm. 2025 June;6(6):e70252.
13. Pannu N, Bhatnagar A. Resveratrol: from enhanced biosynthesis and bioavailability to multi-targeting chronic diseases. Biomed Pharmacother. 2019 Jan;109:2237–51.
14. Salehi B, Mishra AP, Nigam M, Sener B, Kilic M, Sharifi-Rad M, et al. Resveratrol: A Double-Edged Sword in Health Benefits. Biomedicines. 2018 Sept 9;6(3):91.
15. Meng Q, Li J, Wang C, Shan A. Biological function of resveratrol and its application in animal production: a review. J Anim Sci Biotechnol. 2023 Feb 11;14(1):25.
16. Fiod Riccio BV, Fonseca-Santos B, Colerato Ferrari P, Chorilli M. Characteristics, Biological Properties and Analytical Methods of *Trans* -Resveratrol: A Review. Crit Rev Anal Chem. 2020 July 3;50(4):339–58.
17. Santos AC, Pereira I, Pereira-Silva M, Ferreira L, Caldas M, Magalhães M, et al. Nanocarriers for resveratrol delivery: Impact on stability and solubility concerns. Trends Food Sci Technol. 2019 Sept;91:483–97.
18. Jang JY, Im E, Kim ND. Mechanism of Resveratrol-Induced Programmed Cell Death and New Drug Discovery against Cancer: A Review. Int J Mol Sci. 2022 Nov 8;23(22):13689.
19. Valletta A, Iozia LM, Leonelli F. Impact of Environmental Factors on Stilbene Biosynthesis. Plants. 2021 Jan 4;10(1):90.
20. Dubrovina AS, Kiselev KV. Regulation of stilbene biosynthesis in plants. Planta. 2017 Oct;246(4):597–623.
21. Kumar S, Chang YC, Lai KH. Resveratrol, a Molecule with Anti-Inflammatory and Anti-Cancer Activities: Natural Product to Chemical Synthesis. Curr Med Chem. 2021;28(19):3773–86.
22. Yang Y, Sun Y, Gu T, Yan Y, Guo J, Zhang X, et al. The Metabolic Characteristics and Bioavailability of Resveratrol Based on Metabolic Enzymes. Nutr Rev. 2024 Nov 9;83(4):749–70.

23. Ramírez-Garza S, Laveriano-Santos E, Marhuenda-Muñoz M, Storniolo C, Tresserra-Rimbau A, Vallverdú-Queralt A, et al. Health Effects of Resveratrol: Results from Human Intervention Trials. *Nutrients*. 2018 Dec 3;10(12):1892.
24. Favari C, Rinaldi De Alvarenga JF, Sánchez-Martínez L, Tosi N, Mignogna C, Cremonini E, et al. Factors driving the inter-individual variability in the metabolism and bioavailability of (poly) phenolic metabolites: A systematic review of human studies. *Redox Biol*. 2024 May;71:103095.
25. Barber TM, Kabisch S, Randeva HS, Pfeiffer AFH, Weickert MO. Implications of Resveratrol in Obesity and Insulin Resistance: A State-of-the-Art Review. *Nutrients*. 2022 July 13;14(14):2870.
26. Luca SV, Macovei I, Bujor A, Miron A, Skalicka-Woźniak K, Aprutosaie AC, et al. Bioactivity of dietary polyphenols: The role of metabolites. *Crit Rev Food Sci Nutr*. 2020 Feb 21;60(4):626–59.
27. Chimento A, De Amicis F, Sirianni R, Sinicropi MS, Puoci F, Casaburi I, et al. Progress to Improve Oral Bioavailability and Beneficial Effects of Resveratrol. *Int J Mol Sci*. 2019 Mar 19;20(6):1381.
28. Islam SU, Muhammad B. A, Islam MU, Shehzad A, Young Sup L. Switching from Conventional to Nano-natural Phytochemicals to Prevent and Treat Cancers: Special Emphasis on Resveratrol. *Curr Pharm Des*. 2019 Sept 1;25(34):3620–32.
29. Su CF, Jiang L, Zhang XW, Iyaswamy A, Li M. Resveratrol in Rodent Models of Parkinson's Disease: A Systematic Review of Experimental Studies. *Front Pharmacol*. 2021 Apr 22;12:644219.
30. Wang Q, Yu Q, Wu M. Antioxidant and neuroprotective actions of resveratrol in cerebrovascular diseases. *Front Pharmacol*. 2022 Sept 5;13:948889.
31. Thiruvengadam M, Venkidasamy B, Subramanian U, Samynathan R, Ali Shariati M, Rebezov M, et al. Bioactive Compounds in Oxidative Stress-Mediated Diseases: Targeting the NRF2/ARE Signaling Pathway and Epigenetic Regulation. *Antioxidants*. 2021 Nov 23;10(12):1859.
32. Alavi M, Farkhondeh T, Aschner M, Samarghandian S. Resveratrol mediates its anti-cancer effects by Nrf2 signaling pathway activation. *Cancer Cell Int*. 2021 Dec;21(1):579.
33. Pourbagher-Shahri AM, Farkhondeh T, Jafari-Nozad AM, Darroudi M, Naseri K, Amirian M, et al. Nrf2 Mediates Effect of Resveratrol in Ischemia-reperfusion Injury. *Curr Mol Pharmacol*. 2024 Jan 30;17:e18761429246578.
34. Gaggini M, Fenizia S, Vassalle C. Sphingolipid Levels and Signaling via Resveratrol and Antioxidant Actions in Cardiometabolic Risk and Disease. *Antioxidants*. 2023 May 16;12(5):1102.
35. Arinno A, Apaijai N, Chattipakorn SC, Chattipakorn N. The roles of resveratrol on cardiac mitochondrial function in cardiac diseases. *Eur J Nutr*. 2021 Feb;60(1):29–44.
36. Ashrafzadeh M, Javanmardi S, Moradi-Ozarlou M, Mohammadinejad R, Farkhondeh T, Samarghandian S, et al. Natural products and phytochemical nanoformulations targeting mitochondria in oncotherapy: an updated review on resveratrol. *Biosci Rep*. 2020 Apr 30;40(4):BSR20200257.
37. Griñán-Ferré C, Bellver-Sanchis A, Izquierdo V, Corpas R, Roig-Soriano J, Chillón M, et al. The pleiotropic neuroprotective effects of resveratrol in cognitive decline and Alzheimer's disease pathology: From antioxidant to epigenetic therapy. *Ageing Res Rev*. 2021 May;67:101271.
38. Meng X, Zhou J, Zhao CN, Gan RY, Li HB. Health Benefits and Molecular Mechanisms of Resveratrol: A Narrative Review. *Foods*. 2020 Mar 14;9(3):340.
39. Vikal A, Maurya R, Bhowmik S, Khare S, Raikwar S, Patel P, et al. Resveratrol: A comprehensive review of its multifaceted health benefits, mechanisms of action, and potential therapeutic applications in chronic disease. *Pharmacol Res - Nat Prod*. 2024 June;3:100047.
40. Gal R, Deres L, Toth K, Halmosi R, Habon T. The Effect of Resveratrol on the Cardiovascular System from Molecular Mechanisms to Clinical Results. *Int J Mol Sci*. 2021 Sept 21;22(18):10152.
41. Godos J, Romano GL, Gozzo L, Laudani S, Paladino N, Dominguez Azpiroz I, et al. Resveratrol and vascular health: evidence from clinical studies and mechanisms of actions related to its metabolites produced by gut microbiota. *Front Pharmacol*. 2024 Mar 18;15:1368949.

42. Banez MJ, Geluz MI, Chandra A, Hamdan T, Biswas OS, Bryan NS, et al. A systemic review on the antioxidant and anti-inflammatory effects of resveratrol, curcumin, and dietary nitric oxide supplementation on human cardiovascular health. *Nutr Res.* 2020 June;78:11–26.
43. Xiao Q, Zhu W, Feng W, Lee SS, Leung AW, Shen J, et al. A Review of Resveratrol as a Potent Chemoprotective and Synergistic Agent in Cancer Chemotherapy. *Front Pharmacol.* 2019 Jan 9;9:1534.
44. Wu SX, Xiong RG, Huang SY, Zhou DD, Saimaiti A, Zhao CN, et al. Effects and mechanisms of resveratrol for prevention and management of cancers: An updated review. *Crit Rev Food Sci Nutr.* 2023 Dec 20;63(33):12422–40.
45. Novelle MG, Wahl D, Diéguez C, Bernier M, De Cabo R. Resveratrol supplementation: Where are we now and where should we go? *Ageing Res Rev.* 2015 May;21:1–15.
46. Malaguarnera L. Influence of Resveratrol on the Immune Response. *Nutrients.* 2019 Apr 26;11(5):946.
47. Inchingolo AD, Malcangi G, Inchingolo AM, Piras F, Settanni V, Garofoli G, et al. Benefits and Implications of Resveratrol Supplementation on Microbiota Modulations: A Systematic Review of the Literature. *Int J Mol Sci.* 2022 Apr 5;23(7):4027.
48. Shaito A, Posadino AM, Younes N, Hasan H, Halabi S, Alhababi D, et al. Potential Adverse Effects of Resveratrol: A Literature Review. *Int J Mol Sci.* 2020 Mar 18;21(6):2084.
49. Trombino S, Cassano R, Di Gioia ML, Aiello F. Emerging Trends in Green Extraction Techniques, Chemical Modifications, and Drug Delivery Systems for Resveratrol. *Antioxidants.* 2025 May 29;14(6):654.
50. Faisal Z, Mazhar A, Batool SA, Akram N, Hassan M, Khan MU, et al. Exploring the multimodal health-promoting properties of resveratrol: A comprehensive review. *Food Sci Nutr.* 2024 Apr;12(4):2240–58.
51. Prakash V, Bose C, Sunilkumar D, Cherian RM, Thomas SS, Nair BG. Resveratrol as a Promising Nutraceutical: Implications in Gut Microbiota Modulation, Inflammatory Disorders, and Colorectal Cancer. *Int J Mol Sci.* 2024 Mar 16;25(6):3370.
52. Ratz-Lyko A, Arct J. Resveratrol as an active ingredient for cosmetic and dermatological applications: a review. *J Cosmet Laser Ther.* 2019 Feb 17;21(2):84–90.
53. Farhan M, Rizvi A. The Pharmacological Properties of Red Grape Polyphenol Resveratrol: Clinical Trials and Obstacles in Drug Development. *Nutrients.* 2023 Oct 23;15(20):4486.
54. Brown K, Theofanous D, Britton RG, Aburido G, Pepper C, Sri Undru S, et al. Resveratrol for the Management of Human Health: How Far Have We Come? A Systematic Review of Resveratrol Clinical Trials to Highlight Gaps and Opportunities. *Int J Mol Sci.* 2024 Jan 6;25(2):747.

CHAPTER 10

CYANIDIN AND HUMAN HEALTH: MECHANISMS AND APPLICATIONS

Sevim İpek ACAR CÖMERT¹

Introduction

The Place of Cyanidin within the Flavonoid Family

Flavonoids represent a diverse family of plant-derived polyphenolic compounds with significant health implications. Within this family, anthocyanins constitute a pigmented subclass responsible for the characteristic red, purple, and blue hues observed in numerous fruits and vegetables. Six major anthocyanidin structures—pelargonidin, cyanidin, delphinidin, peonidin, petunidin, and malvidin—predominate in nature, with cyanidin exhibiting particularly pronounced biological activity. Contemporary investigations have characterized cyanidin's antioxidant, anti-inflammatory, and metabolic regulatory properties. Berry species including red raspberry (*Rubus idaeus* L.), blue honeysuckle (*Lonicera caerulea* L.), and mulberry have long traditions in folk medicine, and their extracts now attract scientific interest for potential applications in cardiovascular disease, obesity, neurodegeneration, hepatic dysfunction, and oncology. Cyanidin-3-glucoside predominates in fruits and edible flowers, while cyanidin-3-galactoside also occurs abundantly across fruit species. The aglycone form is principal in vegetables, whereas cyanidin-3-glucoside appears notably in certain cereal grains (1).

¹ Asst. Prof. Dr., Giresun University, Faculty of medicine, Department of Physiology, ipek.comert@giresun.edu.tr, ORCID iD: 0000-0003-3696-9473

REFERENCES

1. Tena, N., Martín, J., & Asuero, A. G. State of the art of anthocyanins: Antioxidant activity, sources, bioavailability, and therapeutic effect in human health. *Antioxidants*. 2020; 9(5), 451.
2. Ye, X., Chen, W., Yan, F., Zheng, X., & Tu, P. Cyanidin-3-O-glucoside enhances GLP-1 secretion via PPAR β / δ - β -catenin-TCF-4 pathway in type 2 diabetes mellitus. *npj Science of Food*. 2025; 9(1), 81.
3. Maslov, D. L., Ipatova, O. M., Tsvetkova, T. A., & Prozorovskii, V. N. Hypoglycemic effect of an extract from Aronia melanocarpa leaves. *Voprosy meditsinskoi khimii*. 2002; 48(3), 271-277.
4. Pilaczynska-Szczesniak, L., Skarpanska-Steinborn, A., Deskur, E., Basta, P., & Horoszkiewicz-Hasan, M. The influence of chokeberry juice supplementation on the reduction of oxidative stress resulting from an incremental rowing ergometer exercise. *International journal of sport nutrition and exercise metabolism*. 2005; 15(1), 48-58.
5. Broncel1ABDEFG, M., Koziróg1ABDEF, M., Duchnowicz2ABD, P., Koter-Michalak2ABD, M., Sikora3ACD, J., & Chojnowska-Jezierska1ABG, J. Aronia melanocarpa extract reduces blood pressure, serum endothelin, lipid, and oxidative stress marker levels in patients with metabolic syndrome. *Med Sci Monit*. 2010; 16(1), 34.
6. Qin, B., & Anderson, R. A. An extract of chokeberry attenuates weight gain and modulates insulin, adipogenic and inflammatory signalling pathways in epididymal adipose tissue of rats fed a fructose-rich diet. *British Journal of Nutrition*. 2012; 108(4), 581-587.
7. Shi, M., O'Keefe, L., Simcocks, A. C., Su, X. Q., & McAinch, A. J. The effect of cyanidin-3-O- β -glucoside and peptides extracted from yoghurt on glucose uptake and gene expression in human primary skeletal muscle myotubes from obese and obese diabetic participants. *Journal of Functional Foods*. 2018; 51, 55-64.
8. Shi, M., Mathai, M. L., Xu, G., McAinch, A. J., & Su, X. Q. The effects of supplementation with blueberry, cyanidin-3-O- β -glucoside, yoghurt and its peptides on obesity and related comorbidities in a diet-induced obese mouse model. *Journal of functional foods*. 2019; 56, 92-101.
9. Daveri, E., Cremonini, E., Mastaloudis, A., Hester, S. N., Wood, S. M., Waterhouse, A. L., & Oteiza, P. I. Cyanidin and delphinidin modulate inflammation and altered redox signaling improving insulin resistance in high fat-fed mice. *Redox biology*. 2018; 18, 16-24.
10. Tian, L., Ning, H., Shao, W., Song, Z., Badakhshi, Y., Ling, W., & Jin, T. Dietary cyanidin-3-glucoside attenuates high-fat-diet-induced body-weight gain and impairment of glucose tolerance in mice via effects on the hepatic hormone FGF21. *The Journal of nutrition*. 2020; 150(8), 2101-2111.
11. Zhao, R., Xiang, B., Dolinsky, V. W., Xia, M., & Shen, G. X. Saskatoon berry powder reduces hepatic steatosis and insulin resistance in high fat-high sucrose diet-induced obese mice. *The Journal of Nutritional Biochemistry*. 2021; 95, 108778.
12. Tsuda, T., Ueno, Y., Yoshikawa, T., Kojo, H., & Osawa, T. Microarray profiling of gene expression in human adipocytes in response to anthocyanins. *Biochemical pharmacology*. 2006; 71(8), 1184-1197.
13. Scazzocchio, B., Vari, R., Filesì, C., D'Archivio, M., Santangelo, C., Giovannini, C., & Masella, R. Cyanidin-3-O- β -glucoside and protocatechuic acid exert insulin-like effects by upregulating PPAR γ activity in human omental adipocytes. *Diabetes*. 2011; 60(9), 2234-2244.
14. Tsuda, T., Horio, F., Uchida, K., Aoki, H., & Osawa, T. Dietary cyanidin 3-O- β -D-glucoside-rich purple corn color prevents obesity and ameliorates hyperglycemia in mice. *The Journal of nutrition*. 2003; 133(7), 2125-2130.
15. Lim, S. M., Lee, H. S., Jung, J. I., Kim, S. M., Kim, N. Y., Seo, T. S., & Kim, E. J. Cyanidin-3-O-galactoside-enriched Aronia melanocarpa extract attenuates weight gain and adipogenic pathways in high-fat diet-induced obese C57BL/6 mice. *Nutrients*. 2019; 11(5), 1190.
16. Han, S., Yang, Y., Lu, Y., Guo, J., Han, X., Gao, Y., & Zhan, J. Cyanidin-3-O-glucoside regulates the expression of Ucp1 in brown adipose tissue by activating Prdm16 gene. *Antioxidants*. 2021; 10(12), 1986.

17. You, Y., Yuan, X., Liu, X., Liang, C., Meng, M., Huang, Y., & Zhan, J. Cyanidin-3-glucoside increases whole body energy metabolism by upregulating brown adipose tissue mitochondrial function. *Molecular Nutrition & Food Research*. 2017; 61(11), 1700261.
18. You, Y., Han, X., Guo, J., Guo, Y., Yin, M., Liu, G., & Zhan, J. Cyanidin-3-glucoside attenuates high-fat and high-fructose diet-induced obesity by promoting the thermogenic capacity of brown adipose tissue. *Journal of Functional Foods*. 2018; 41, 62-71.
19. Tomay, F., Marinelli, A., Leoni, V., Caccia, C., Matros, A., Mock, H. P., & Petroni, K. Purple corn extract induces long-lasting reprogramming and M2 phenotypic switch of adipose tissue macrophages in obese mice. *Journal of Translational Medicine*. 2019; 17(1), 237.
20. Jia, Y., Wu, C., Kim, Y. S., Yang, S. O., Kim, Y., Kim, J. S., & Lee, S. J. A dietary anthocyanin cyanidin-3-O-glucoside binds to PPARs to regulate glucose metabolism and insulin sensitivity in mice. *Communications Biology*. 2020; 3(1), 514.
21. Guo, H., Liu, G., Zhong, R., Wang, Y., Wang, D., & Xia, M. Cyanidin-3-O- β -glucoside regulates fatty acid metabolism via an AMP-activated protein kinase-dependent signaling pathway in human HepG2 cells. *Lipids in health and disease*. 2012; 11(1), 10.
22. Yu, R. Q., Wu, X. Y., Zhou, X., & Zhu, J. Cyanidin-3-glucoside attenuates body weight gain, serum lipid concentrations and insulin resistance in high-fat diet-induced obese rats. *Zhongguo dangdai erke zazhi*. 2014; 534-538.
23. Biswas, D., Sarkar, S., De Silva, A. B. K. H., D'Souza, K., Kienesberger, P., Rupasinghe, H. V., & Pulinilkunnil, T. Cyanidin-3-O-glucoside rich extract from haskap berry improves glucose homeostasis and insulin sensitivity in diet-induced obese mice. *Canadian Journal of Diabetes*. 2018; 42(5), S55.
24. Pei, L., Wan, T., Wang, S., Ye, M., Qiu, Y., Jiang, R., & Yang, L. Cyanidin-3-O- β -glucoside regulates the activation and the secretion of adipokines from brown adipose tissue and alleviates diet induced fatty liver. *Biomedicine & Pharmacotherapy*. 2018; 105, 625-632.
25. Sikora, J., Broncel, M., & Mikiciuk-Olasik, E. Aronia melanocarpa Elliot reduces the activity of angiotensin i-converting enzyme—in vitro and ex vivo studies. *Oxidative Medicine and Cellular Longevity*. 2014; 2014 (1), 739721.
26. Poreba, R., Skoczynska, A., Gac, P., Poreba, M., Jedrychowska, I., Affelska-Jercha, A., & Andrzejak, R. Drinking of chokeberry juice from the ecological farm Dzieciolowo and distensibility of brachial artery in men with mild hypercholesterolemia. *Annals of Agricultural and Environmental Medicine*. 2009; 16(2), 305-308.
27. Tan, J., Li, Y., Hou, D. X., & Wu, S. The effects and mechanisms of cyanidin-3-glucoside and its phenolic metabolites in maintaining intestinal integrity. *Antioxidants*. 2019; 8(10), 479.
28. Zhu, Y., Sun, H., He, S., Lou, Q., Yu, M., Tang, M., & Tu, L. Metabolism and prebiotics activity of anthocyanins from black rice (*Oryza sativa* L.) in vitro. *Plos one*. 2018; 13(4), e0195754.
29. Min, S. W., Ryu, S. N., & Kim, D. H. Anti-inflammatory effects of black rice, cyanidin-3-O- β -D-glucoside, and its metabolites, cyanidin and protocatechuic acid. *International immunopharmacology*. 2010; 10(8), 959-966.
30. Deepa, P., Hong, M., Sowndhararajan, K., & Kim, S. A Review of the Role of an Anthocyanin, Cyanidin-3-O- β -glucoside in Obesity-Related Complications. *Plants*. 2023; 12(22), 3889.
31. Rahman, S., Mathew, S., Nair, P., Ramadan, W. S., & Vazhappilly, C. G. Health benefits of cyanidin-3-glucoside as a potent modulator of Nrf2-mediated oxidative stress. *Inflammopharmacology*. 2021; 29(4), 907-923.
32. Tang, Z. Cyanidin-3-glucoside: Targeting Atherosclerosis through Gut Microbiota and Anti-Inflammation. *Frontiers in Nutrition*. 2025; 12, 1627868.
33. Jiao, X., Shen, Y., Deng, H., Zhang, Q., & Zhao, J. Cyanidin-3-O-galactoside from Aronia melanocarpa attenuates high-fat diet-induced obesity and inflammation via AMPK, STAT3, and NF- κ B p65 signaling pathways in Sprague-Dawley rats. *Journal of functional foods*. 2021; 85, 104616.

34. Yang HongPeng, Y. H., Jiang YuGang, J. Y., Pang Wei, P. W., Chen DaoMei, C. D., Lu Hao, L. H., & Lu ShiJun, L. S. Study on the effect of anthocyanin from blueberry on improving learning and memory in aged mice. 2009
35. Yang, H., Pang, W., Lu, H., Cheng, D., Yan, X., Cheng, Y., & Jiang, Y. Comparison of metabolic profiling of cyanidin-3-O-galactoside and extracts from blueberry in aged mice. *Journal of agricultural and food chemistry*. 2011; 59(5), 2069-2076.
36. Lee, H. Y., Weon, J. B., Jung, Y. S., Kim, N. Y., Kim, M. K., & Ma, C. J. Cognitive-Enhancing Effect of Aronia melanocarpa Extract against Memory Impairment Induced by Scopolamine in Mice. *Evidence-Based Complementary and Alternative Medicine*. 2016; 2016(1), 6145926.
37. Skemiene, K., Pampuscenko, K., Rekuviene, E., & Borutaite, V. (2020). Protective effects of anthocyanins against brain ischemic damage. *Journal of Bioenergetics and Biomembranes*, 52(2), 71-82.
38. Wen, H., Cui, H., Tian, H., Zhang, X., Ma, L., Ramassamy, C., & Li, J. Isolation of neuroprotective anthocyanins from black chokeberry (*Aronia melanocarpa*) against amyloid- β -induced cognitive impairment. *Foods*. 2020; 10(1), 63.
39. Shim, S. H., Kim, J. M., Choi, C. Y., Kim, C. Y., & Park, K. H. Ginkgo biloba extract and bilberry anthocyanins improve visual function in patients with normal tension glaucoma. *Journal of medicinal food*. 2012; 15(9), 818-823.
40. Miyake, S., Takahashi, N., Sasaki, M., Kobayashi, S., Tsubota, K., & Ozawa, Y. Vision preservation during retinal inflammation by anthocyanin-rich bilberry extract: cellular and molecular mechanism. *Laboratory investigation*. 2012; (1), 102-109.
41. Safdar, M. A., Aslam, R. M. N., Shakeel, A., Shiza, Waqar, M., Jmail, A., & Gul, H. (2023). Cyanidin as potential anticancer agent targeting various proliferative pathways. *Chemical Biology & Drug Design*, 101(2), 438-452.
42. Wallace, T. C., & Giusti, M. M. (Eds.) *Anthocyanins in health and disease*. CRC Press, Taylor & Francis. 2014.
43. Chen, P. N., Kuo, W. H., Chiang, C. L., Chiou, H. L., Hsieh, Y. S., & Chu, S. C. Black rice anthocyanins inhibit cancer cells invasion via repressions of MMPs and u-PA expression. *Chemico-biological interactions*. 2006; 163(3), 218-229.
44. Lala, G., Malik, M., Zhao, C., He, J., Kwon, Y., Giusti, M. M., & Magnuson, B. A. Anthocyanin-rich extracts inhibit multiple biomarkers of colon cancer in rats. *Nutrition and cancer*. 2006; 54(1), 84-93.
45. Ding, M., Feng, R., Wang, S. Y., Bowman, L., Lu, Y., Qian, Y., & Shi, X. Cyanidin-3-glucoside, a natural product derived from blackberry, exhibits chemopreventive and chemotherapeutic activity. *Journal of Biological Chemistry*. 2006; 281(25), 17359-17368.
46. Wu, C. F., Wu, C. Y., Lin, C. F., Liu, Y. W., Lin, T. C., Liao, H. J., & Chang, G. R. The anticancer effects of cyanidin 3-O-glucoside combined with 5-fluorouracil on lung large-cell carcinoma in nude mice. *Biomedicine & Pharmacotherapy*. 2022; 151, 113128.
47. Pratheeshkumar, P., Son, Y. O., Wang, X., Divya, S. P., Joseph, B., Hitron, J. A., & Shi, X. Cyanidin-3-glucoside inhibits UVB-induced oxidative damage and inflammation by regulating MAP kinase and NF- κ B signaling pathways in SKH-1 hairless mice skin. *Toxicology and applied pharmacology*. 2014; 280(1), 127-137.
48. Liu, M., Du, Y., Li, H., Wang, L., Ponikwicka-Tyszko, D., Lebedzinska, W., & Li, X. Cyanidin-3-o-glucoside pharmacologically inhibits tumorigenesis via estrogen receptor β in melanoma mice. *Frontiers in oncology*. 2019; 9, 1110.
49. Li, X., Mu, J., Lin, Y., Zhao, J., & Meng, X. Combination of cyanidin-3-O-glucoside and cisplatin induces oxidative stress and apoptosis in HeLa cells by reducing activity of endogenous antioxidants, increasing bax/bcl-2 mRNA expression ratio, and downregulating Nrf2 expression. *Journal of Food Biochemistry*. 2021; 45(7), e13806.
50. Majerik Behinska, K., Balkova, E., Mihal, M., Roychoudhury, S., & Kolesarova, A. The therapeutic potential of cyanidin-3-O-glucoside relating to female reproductive health. *Frontiers in Pharmacology*. 2025; 16, 1599688.

CHAPTER 11

RUTIN

Bengü TEMİZEL¹

Introduction

Natural products have been the primary source of many life-saving medicines, cosmetics and dietary supplements throughout history (1). Many health products, including the semi synthetic drugs used today, are either derived directly from natural sources or are inspired by the structures of natural products (2). Today, thousands of natural molecules, particularly those derived from plants, have been studied for their biological activity against various human diseases and a large number of them have been identified as having therapeutic potential (3).

Flavonoids, which have long attracted the attention of scientists due to their positive effects on human health, are particularly popular today thanks to their antioxidant and membrane-stabilizing properties (4). Rutin (quercetin-3-O-rutinoside), a prominent member of the flavonoid group, is a natural compound widely found in plants. It takes its name from the *Ruta graveolens* plant, which contains high amounts of rutin. It is also known by other names such as rutoside, quercetin-3-O-rutinoside and sophorin. Rutin, which plays a protective role in plants, is abundant in a wide variety of plant species, including buckwheat, apples, citrus fruits and berries.

Numerous studies have demonstrated the antioxidant, anti-inflammatory, anticancer, antimicrobial, cardioprotective and neuroprotective properties of rutin. These effects are underpinned by rutin's ability to neutralize free radicals, bind metal ions and regulate intracellular signaling pathways. Furthermore, rutin's

¹ Giresun University, bengu.temizel@giresun.edu.tr, ORCID iD: 0000-0002-5217-3013

REFERENCES

1. Li JWH, Vederas JC. Drug discovery and natural products: end of an era or an endless frontier. *Science*; 2009;325(5937): 161-165. doi:10.1126/science.1168243
2. Chaachouay N, Zidane L. Plant-derived natural products: a source for drug discovery and development. *Drugs and Drug Candidates*; 2024;3(1): 184-207. doi:10.3390/ddc301001
3. Pandey P, Khan F, Qari HA, Oves M. Rutin (Bioflavonoid) as cell signaling pathway modulator: Prospects in treatment and chemoprevention. *Pharmaceuticals*; 2021;14(11): 1069. doi:10.3390/ph14111069
4. Patel K, Patel DK. The beneficial role of rutin, a naturally occurring flavonoid in health promotion and disease prevention: A systematic review and update. *Bioactive food as dietary interventions for arthritis and related inflammatory diseases*; 2019;457-479. doi:10.1016/B978-0-12-813820-5.00026-X
5. Frutos MJ, Rincón-Frutos L, Valero-Cases, E. *Rutin*. In *Nonvitamin and nonmineral nutritional supplements*. Academic Press; 2019. p. 111-117.
6. Forouzanfar F, Sahranavard T, Tsatsakis A, Iranshahi M, Rezaee, R. Rutin: a pain-relieving flavonoid. *Inflammopharmacology*; 2025; 1-13. doi:10.1007/s10787-025-01671-8
7. National Center for Biotechnology Information. PubChem Compound Summary for CID 5280805, 2025; Rutin.
8. Semwal R, Joshi SK, Semwal RB, Semwal DK. Health benefits and limitations of rutin-A natural flavonoid with high nutraceutical value. *Phytochemistry Letters*; 2021;46: 119-128. doi: 10.1016/j.phytol.2021.10.006
9. Tobar-Delgado E, Mejía-España D, Osorio-Mora O, Serna-Cock L. Rutin: family farming products' extraction sources, industrial applications and current trends in biological activity protection. *Molecules*; 2023;28(15): 5864. doi:10.3390/molecules28155864
10. Sequeira IR, Poppitt SD. Unfolding novel mechanisms of polyphenol flavonoids for better glycaemic control: Targeting pancreatic islet amyloid polypeptide (IAPP). *Nutrients*; 2017;9(7): 788. doi:10.3390/nu9070788
11. Li B, Yang X. Rutin-loaded cellulose acetate/poly (ethylene oxide) fiber membrane fabricated by electrospinning: a bioactive material. *Materials Science and Engineering*; 2020;109: 110601. doi:10.1016/j.msec.2019.110601
12. Selvaraj G, Kaliamurthi S, Thirungnasambandam R, Vivekanandan L, Balasubramanian, T. Anti-nociceptive effect in mice of thillai flavonoid rutin. *Biomed Environ Sci.*; 2014;27(4): 295-299. doi:10.3967/bes2014.052
13. Gęgotek A, Bielawska K, Biernacki M, Dobrzyńska I, Skrzydlewska E. Time-dependent effect of rutin on skin fibroblasts membrane disruption following UV radiation. *Redox Biology*; 2017;12: 733-744. doi:10.1016/j.redox.2017.04.014
14. Kamel R, Mostafa DM. Rutin nanostructured lipid cosmeceutical preparation with sun protective potential. *Journal of photochemistry and photobiology B: biology*; 2015;153: 59-66. doi: 10.1016/j.jphotobiol.2015.09.002.
15. Park SN, Lee MH, Kim SJ, Yu ER. Preparation of quercetin and rutin-loaded ceramide liposomes and drug-releasing effect in liposome-in-hydrogel complex system. *Biochemical and biophysical research communications*; 2013;435(3): 361-366. doi:10.1016/j.bbrc.2013.04.093.
16. Cui Z, Kong X, Chen Y, Zhang C, Hua Y. Effects of rutin incorporation on the physical and oxidative stability of soy protein-stabilized emulsions. *Food Hydrocolloids*; 2014;41: 1-9. doi: 10.1016/j.foodhyd.2014.03.006
17. Gullón B, Lú-Chau, TA, Moreira MT, Lema JM, Eibes G. Rutin: A review on extraction, identification and purification methods, biological activities and approaches to enhance its bioavailability. *Trends in food science & technology*; 2017;67: 220-235. doi: 10.1016/j.tifs.2017.07.008
18. Yang N, Ren G. Application of near-infrared reflectance spectroscopy to the evaluation of rutin and d-chiro-inositol contents in tartary buckwheat. *Journal of Agricultural and Food Chemistry*; 2008; 56: 761e764. doi: 10.1021/jf072453u

19. Kamalakkannan N, Prince SMP. Rutin improves the antioxidant status in streptozotocin-induced diabetic rat tissues. *Molecular and Cellular Biochemistry*; 2006; 293: 211e219. doi: 10.1007/s11010-006-9244-1
20. Pulido R, Bravo L, Saura-Calixto F. Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry*; 2000; 48: 3396e3402. doi: 10.1021/jf9913458
21. Yang J, Guo J, Yuan J. In vitro antioxidant properties of rutin. *LWT-Food Science and Technology*; 2008; 41: 1060e1066. doi: 10.1016/j.lwt.2007.06.010
22. Sharma S, Ali A, Ali J, Sahni JK, Baboota S. Rutin: Therapeutic potential and recent advances in drug delivery. *Expert Opinion on Investigational Drugs*; 2013a; 22: 1063e1079. doi: 10.1517/13543784.2013.805744
23. Boyle SP, Dobson VL, Duthie SJ, Hinselwood DC, Kyle JA, Collins AR. Bioavailability and efficiency of rutin as an antioxidant: A human supplementation study. *European Journal of Clinical Nutrition*; 2000; 54: 774e782.
24. Chua LS. A review on plant-based rutin extraction methods and its pharmacological activities. *Journal of Ethnopharmacology*; 2013; 150: 805e817. doi: 10.1016/j.jep.2013.10.036
25. Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: impact on human health. *Pharmacogn. Rev.*; 2010; 4 (8): 118–126. doi: 10.4103/0973-7847.70902
26. Singh S, Singh DK, Meena A, Dubey V, Masood N, Luqman S. Rutin protects to butyl hydroperoxide-induced oxidative impairment via modulating the Nrf2 and iNOS activity. *Phytomedicine*; 2019;55: 92–104. doi: 10.1016/j.phymed.2018.07.009
27. Li S, Li J, Pan R, Cheng J, Cui Q, Chen J, Yuan Z. Sodium rutin extends lifespan and health span in mice including positive impacts on liver health. *British journal of pharmacology*; 2022; 179(9): 1825-1838. doi: 10.1111/bph.15410
28. Cushnie TT, Lamb AJ. Recent advances in understanding the antibacterial properties of flavonoids. *International Journal of Antimicrobial Agents*; 2011;38: 99e107. doi: 10.1016/j.ijantimicag.2011.02.014
29. Tsuchiya H, Iinuma M. Reduction of membrane fluidity by antibacterial sophoraflavanone G isolated from *Sophora exigua*. *Phytomedicine*; 2000;7: 161e165. doi: 10.1016/S0944-7113(00)80089-6
30. Plaper A, Golob M, Hafner I, Oblak M, Solmajer T, Jerala R. Characterization of quercetin binding site on DNA gyrase. *Biochemical and Biophysical Research Communications*; 2003;306: 530e536. doi: 10.1016/S0006-291X(03)01006-4
31. Haraguchi H, Tanimoto K, Tamura Y, Mizutani K, Kinoshita T. Mode of antibacterial action of retrochalcones from *Glycyrrhiza inflata*. *Phytochemistry*; 1998;48: 125e129. doi: 10.1016/S0031-9422(97)01105-9
32. Mazzeo MF, Lippolis R, Sorrentino A, Liberti S, Fragnito F, Siciliano RA. Lactobacillus acidophilus-rutin interplay investigated by proteomics. *PLoS One*; 2015;10: e0142376. doi: 10.1371/journal.pone.0142376
33. Rym KH, Eo SK, Kim YS, Lee CK, Han SS. Antimicrobial activity and acute toxicity of natural rutin. *Korean Journal of Pharmacognosy*; 1996; 27, 309e315.
34. Soni H Malik, J, Singhai AK, Sharma S. Antimicrobial and antiinflammatory activity of the hydrogels containing rutin delivery. *Asian Journal of Chemistry*; 2013;25: 8371e8373. doi: 10.14233/ajchem.2013.14912
35. Singh M, Govindarajan R, Rawat AK, Khare PB. Antimicrobial flavonoid rutin from *Pteris vittata* L. against pathogenic gastrointestinal microflora. *American Fern Journal*; 2008; 98: 98e103. doi: 10.1640/0002-8444(2008)98[98:AFRFPV]2.0.CO;2
36. Arima H, Ashida H, Danno GI. Rutin-enhanced antibacterial activities of flavonoids against *Bacillus cereus* and *Salmonella enteritidis*. *Bioscience, Biotechnology, and Biochemistry*; 2002;66: 1009e1014. doi: 10.1271/bbb.66.1009

37. Middleton JE, Kandaswami C. *The impact of plant flavonoids on mammalian biology: Implications for immunity, inflammation and cancer*. In J. B. Harborne (Ed.), *The Flavonoids: Advances in research since 1986*. London, UK: Chapman and Hall; 1994. p. 619e652.
38. Selway JWT. Antiviral activity of flavones and flavans. *Progress in Clinical and Biological Research*; 1986; 213, 521e536.
39. Stojković D, Petrović J, Soković M, Glamočlija J, Kukić-Marković J, Petrović S. In situ antioxidant and antimicrobial activities of naturally occurring caffeic acid, p-coumaric acid and rutin, using food systems. *Journal of the Science of Food and Agriculture*; 2013; 93(13): 3205-3208. doi: 10.1002/jsfa.6156
40. Orhan DD, Özçelik B, Özgen S, Ergun F. Antibacterial, antifungal, and antiviral activities of some flavonoids. *Microbiological Research*; 2010;165: 496e504. doi: 10.1016/j.micres.2009.09.002
41. Dubey S, Ganeshpurkar A, Bansal D, Dubey N. Experimental studies on bioactive potential of rutin. *Chronicles of Young Scientists*; 2013;4: 153.
42. Dong J, Zhang L, Xu N, Zhou S, Song Y, Yang Q, Liu Y, Yang Y, Ai X. Rutin reduces the pathogenicity of *Streptococcus agalactiae* to tilapia by inhibiting the activity of sortase A. *Aquaculture*; 2021;530: 735743. doi: 10.1016/j.aquaculture.2020.735743
43. Basile A, Sorbo, S, Giordano S, Ricciardi L, Ferrara S, Montesano D, Cobianchi RC, Vuotto ML, Ferrara L. Antibacterial and allelopathic activity of extract from *Castanea sativa* leaves. *Fitoterapia*; 2000;71: S110-S116. doi: 10.1016/S0367-326X(00)00185-4
44. An R, Shi C, Tang Y, Cui Z, Li Y, Chen Z, ... Xu L. Chitosan/rutin multifunctional hydrogel with tunable adhesion, anti-inflammatory and antibacterial properties for skin wound healing. *Carbohydrate Polymers*; 2024; 343: 122492. doi: 10.1016/j.carbpol.2024.122492
45. Amin MU, Khurram M, Khattak B, Khan J. Antibiotic additive and synergistic action of rutin, morin and quercetin against methicillin resistant *Staphylococcus aureus*. *BMC Complementary and Alternative Medicine*; 2015;15: 59-72. doi: 10.1186/s12906-015-0580-0
46. Wang S, Wang C, Gao L, Cai H, Zhou Y, Yang Y, Xu C, Ding W, Chen J, Muhammad I, Chen X, He X, Liu D, Li Y. Rutin inhibits *Streptococcus suis* biofilm formation by affecting CPS biosynthesis. *Frontiers in Pharmacology*; 2017; 8(379): 1-12. doi: 10.3389/fphar.2017.00379
47. Danciu C, Pinzaru IA, Dehelean CA, Hancianu M, Zupkó I, Navolan D, Licker M, Ghuilai RM, Șoica CM. Antiproliferative and antimicrobial properties of pure and encapsulated rutin. *Farmacia*; 2018; 66(2): 302-308.
48. Pimentel RBQ, Costa CA, Albuquerque PM, Junior SD. Antimicrobial activity and rutin identification of honey produced by the stingless bee *Melipona compressipes manausensis* and commercial honey. *BMC Complementary and Alternative Medicine*; 2013;13: 151-163. doi: 10.1186/1472-6882-13-151
49. Prasad R, Prasad SB. A review on the chemistry and biological properties of Rutin, a promising nutraceutical agent. *Asian J. Pharm. Pharmacol*; 2019;5(1): 1-20. doi:10.31024/ajpp.2019.5.s1.1
50. Oliveira VM, Carraro E, Aulerb ME, Khalil NM. Quercetin and rutin as potential agents antifungal against *Cryptococcus* spp. *Brazilian Journal of Biology*; 2016;76(4): 1029-1034. doi:10.1590/1519-6984.07415
51. Johann S, Mendes BG, Missau FC, de Resende MA, Pizzolatti MG. Antifungal activity of five species of *Polygala*. *Brazilian Journal of Microbiology*; 2011;42(3): 1065-1075. doi: 10.1590/S1517-83822011000300027
52. Han Y. Rutin has therapeutic effect on septic arthritis caused by *Candida albicans*. *International Immunopharmacology*; 2009;9: 207-211. doi: 10.1016/j.intimp.2008.11.002
53. Mane MD, Patole NS, Kodalkar VN, Metkari SA. An Overview of Antimicrobial Properties of Rutin. *International Journal of Research in Pharmacy and Allied Science*; 2024; 3(2): 17-23.
54. Pereira AP, Ferreira ICFR, Marcelino F, Valentão P, Andrade PB, Seabra R, Estevinho L, Bento A, Pereira JA. Phenolic compounds and antimicrobial activity of olive (*L. Cv. Cobrançosa*) *Olea europaea* leaves. *Molecules*; 2007;12: 1153-1162. doi: 10.3390/12051153

55. Rabišková M, Bautzová T, Gajdziok J, Dvořáčková K, Lamprecht A, Pellequer Y, Spilková J. Coated chitosan pellets containing rutin intended for the treatment of inflammatory bowel disease: in vitro characteristics and in vivo evaluation. *International journal of pharmaceutics*; 2012; 422(1-2): 151-159. doi: 10.1016/j.ijpharm.2011.10.045
56. Torres-Rêgo M, Furtado AA, Bitencourt MAO, Lima MCJDS, Andrade RCLCD, Azevedo EPD, ... Fernandes-Pedrosa MDF. Anti-inflammatory activity of aqueous extract and bioactive compounds identified from the fruits of *Hancornia speciosa* Gomes (Apocynaceae). *BMC Complementary and Alternative Medicine*; 2016;16(1): 275. doi: 10.1186/s12906-016-1259-x
57. Nafees S, Rashid S, Ali N, Hasan SK, Sultana S. Rutin ameliorates cyclophosphamide induced oxidative stress and inflammation in Wistar rats: Role of NFkB/MAPK pathway. *Chemico-biological Interactions*; 2015; 231: 98e107. doi: 10.1016/j.cbi.2015.02.021
58. Hao G, Dong Y, Huo R, Wen K, Zhang Y, Liang G. Rutin inhibits neuroinflammation and provides neuroprotection in an experimental rat model of subarachnoid hemorrhage, possibly through suppressing the RAGEeNF-kB inflammatory signaling pathway. *Neurochemical Research*; 2016; 41: 1496e1504. doi: 10.1007/s11064-016-1863-7
59. Yoo H, Ku SK, Baek YD, Bae JS. Anti-inflammatory effects of rutin on HMGB1-induced inflammatory responses in vitro and in vivo. *Inflammation Research*; 2014;63: 197e206. doi: 10.1007/s00011-013-0689-x
60. Jeong HJ, Yoo MS, Han NR, Hwang SY, Yoon KW, Kim HM. The new therapeutic herbal drug HM0601 and its bioactive compound rutin exert potent antiproliferative activities in mast cells. *Fund Clin Pharmacol*; 2018;32(3): 279-287. doi: 10.1111/fcp.12350
61. Tian C, Guo Y, Chang Y, Zhao J, Cui C, Liu M. Dose-effect relationship on anti-inflammatory activity on LPS induced RAW 264.7 cells and antioxidant activity of rutin in vitro. *Acta Pol. Pharm.*; 2019;76(3): 511-522. doi: 10.32383/appdr/102677
62. Huang FM, Chang YC, Lee MW, Su NY, Yang LC, Kuan YH. Rutin alleviates bisphenol A-glycidyl methacrylate-induced generation of proinflammatory mediators through the MAPK and NF-kB pathways in macrophages. *Environmental Toxicology*; 2023;38(3): 628-634. <https://doi.org/10.1002/tox.23711>
63. Madkour DA, Ahmed M, Elkirdasy AF, Orabi SH, Mousa AA. Rutin: Chemical properties, Pharmacokinetic properties and Biological activities. *Matrouh Journal of Veterinary Medicine*; 2024; 4(1): 26-34. doi:10.21608/mjvm.2024.341806
64. Ren W, Qiao Z, Wang H, Zhu L, Zhang L. Flavonoids: Promising anticancer agents. *Medicinal Research Reviews*; 2003;23(4): 519-534. doi: 10.1002/med.10033
65. Perk AA, Shatynska-Mytsyk I, Gerçek YC, Boztas K, Yazgan M, Fayyaz S., et al. Rutin mediated targeting of signaling machinery in cancer cells. *Cancer Cell International*; 2014; 14: 124e128.
66. Karakurt S. Modulatory effects of rutin on the expression of cytochrome P450s and antioxidant enzymes in human hepatoma cells. *Acta Pharmaceutica*; 2016;66: 491e502. doi: 10.1515/acph-2016-0046
67. Alonso-Castro AJ, Domínguez F, García-Carranca A. Rutin exerts antitumor effects on nude mice bearing SW480 tumor. *Archives of Medical Research*; 2013;44: 346e351. doi: 10.1016/j.arcmed.2013.06.002
68. Gonçalves CF, Santos MC, Ginabreda MG, Fortunato RS, Carvalho DP, Freitas-Ferreira AC. Flavonoid rutin increases thyroid iodide uptake in rats. *PLoS One*; 2013;8: e73908. doi:10.1371/journal.pone.0073908
69. Chen YS, Hu QH, Zhang X, Zhu Q, Kong LD. Beneficial effect of rutin on oxonate-induced hyperuricemia and renal dysfunction in mice. *Pharmacology*; 2013;92(1-2): 75-83. doi:10.1159/000351703
70. Kawaii S, Tomono Y, Katase E, Ogawa K., Yano M. Antiproliferative activity of flavonoids on several cancer cell lines. *Bioscience, Biotechnology and Biochemistry*; 1999;63: 896e899. doi: 10.1271/bbb.63.896

71. Pouget C, Lauthier F, Simon A, Fagnere C, Basly JP, Delage C, et al. Flavonoids: Structural requirements for antiproliferative activity on breast cancer cells. *Bioorganic & Medicinal Chemistry Letters*; 2001;11: 3095e3097. doi: 10.1016/S0960-894X(01)00617-5
72. Srinivasan R, Natarajan D, Shivakumar MS. In vitro evaluation of antioxidant, antiproliferative potentials of bioactive extract-cum-rutin compound isolated from *Memecylon edule* leaves and its molecular docking study. *Journal of Biologically Active Products from Nature*; 2016;6: 43e58. doi: 10.1080/22311866.2016.1173592
73. Ben Sghaier M, Pagano A, Mousslim M, Ammari Y, Kovacic H, Luis J. Rutin inhibits proliferation, attenuates superoxide production and decreases adhesion and migration of human cancerous cells. *Biomedicine & Pharmacotherapy*; 2016;84: 1972e1978. doi: 10.1016/j.biopha.2016.11.001
74. Nasiri F, Kismali G, Alpay M, Kosova F, Cakir DU, Sel T. Rutin enhances the antiproliferative effect of 5-FU and oxaliplatin in colon cancer cells. *Cancer Research*; 2016; 76: 2177. doi: 10.1158/1538-7445.AM2016-2177
75. Yang S, Zhang H, Yang X, Zhao H, Zhu Y. Evaluation of antiproliferative activities of rutin on human colon cancer lovo cells and breast cancer MCF-7 cells. *Anal. Quant. Cytol. Histol.*; 2017;39 (2): 99–107.
76. Caparica R, Júlio A, Araújo MEM, Baby AR, Fonte P, Costa JG, de Almeida TS. Anticancer activity of rutin and its combination with ionic liquids on renal cells. *Biomolecules*; 2020;10(2): 233. doi:10.3390/biom10020233
77. Alajmi, M.F., Alam, P., Rehman, M.T., Husain, F.M., Khan, A.A., Siddiqui, N.A., Hussain, A., Kalam, M.A., Parvez, M.K. Interspecies anticancer and antimicrobial activities of genus *solanum* and estimation of rutin by validated UPLC-PDA Method. *Evid. Complement. Alternat. Med.*; 2018; 6040815. doi:10.1155/2018/6040815
78. Imani A, Maleki N, Bohlouli S, Kouhsoltani M, Sharifi S, Maleki Dizaj S. Molecular mechanisms of anticancer effect of rutin. *Phytother. Res.*; 2021;35(5): 2500-2513. doi:10.1002/ptr.6977
79. Caparica R, Júlio A, Araújo MEM, Baby AR, Fonte P, Costa JG, Santos de Almeida T. Anticancer activity of rutin and its combination with ionic liquids on renal cells. *Biomolecules*; 2020; 10(2): 233. doi:10.3390/biom10020233
80. Goyal J, Verma PK. An overview of biosynthetic pathway and therapeutic potential of rutin. *Mini Reviews in Medicinal Chemistry*; 2023;23(14): 1451-1460. doi: 10.2174/1389557523666230125104101
81. Lee YJ, Jeune KH. The effect of rutin on antioxidant and antiinflammation in streptozotocin-induced diabetic rats. *Applied Microscopy*; 2013;43: 54e64.
82. Saklani R, Gupta SK, Mohanty IR, Kumar B, Srivastava S, Mathur R. Cardioprotective effects of rutin via alteration in TNF- α , CRP, and BNP levels coupled with antioxidant effect in STZ-induced diabetic rats. *Molecular and Cellular Biochemistry*; 2016; 420: 65e72. doi:10.1007/s11010-016-2767-1
83. Al-Ishaq RK, Abotaleb M, Kubatka P, Kajo K, Büsselberg D. Flavonoids and their anti-diabetic effects: cellular mechanisms and effects to improve blood sugar levels. *Biomolecules*; 2019;9(9): 430. doi:10.3390/biom9090430.
84. Calzada F, Solares-Pascasio JI, Ordoñez-Razo RM, Velazquez, C. Barbosa, E. García-Hernández N, Mendez-Luna D, Correa-Basurto J. Antihyperglycemic activity of the leaves from *Annona cherimola* miller and rutin on alloxan-induced diabetic rats. *Pharmacogn Res.*; 2017; 9(1): 1–6. doi:10.4103/0974-8490.199781
85. Chielle EO, Bonfanti G, De Bona KS, Cargnelutti LO, Bitencourt PER, Da Silva PS, Campos MMA, Moretto MB. Rutin restores adenosine deaminase activity in serum and the liver and improves biochemical parameters in streptozotocin-induced diabetic rats. *Rev Bras Plantas Med*; 2016; 18(1): 273–278. doi:10.1590/1983-084X/15_189
86. Chen SS, Gong J, Liu FT, Mohammed U. Naturally occurring polyphenolic antioxidants modulate IgE-mediated mast cell activation. *Immunology*; 2000; 100: 471e480. doi: 10.1046/j.1365-2567.2000.00045.x

87. Kim HY, Nam SY, Hong SW, Kim MJ, Jeong HJ, Kim HM. Protective effects of rutin through regulation of vascular endothelial growth factor in allergic rhinitis. *American Journal of Rhinology and Allergy*; 2015;29: e87e94. doi: 10.2500/ajra.2015.29.4195
88. Choi JK, Kim SH. Rutin suppresses atopic dermatitis and allergic contact dermatitis. *Experimental Biology and Medicine*; 2013;238: 410e417. doi:10.1177/1535370213477
89. Chuffa LGA, Fioruci-Fontanelli BA, Bordon JG, Pires RB, Braga CP, Seiva FRE, Fernandes AAH. Rutin ameliorates glycemic index, lipid profile and enzymatic activities in serum, heart and liver tissues of rats fed with a combination of hypercaloric diet and chronic ethanol consumption. *Indian J. Biochem. Biophys.*; 2014;51(3): 215–222.
90. Manzoni AG, Passos DF, da Silva JLG, Bernardes VM, Bremm JM, Jantsch MH, de Oliveira JS, Mann TR, de Andrade CM, Leal DBR. Rutin and curcumin reduce inflammation, triglyceride levels and ADA activity in serum and immune cells in a model of hyperlipidemia. *Blood Cells Mol. Dis.*; 2019; 76: 13–21. doi:10.1016/j.bcmd.2018.12.005
91. Panda S, Kar A. Antithyroid effects of naringin, hesperidin and rutin in l-T4 induced hyperthyroid rats: possible mediation through 5' DI activity. *Pharmacol. Rep.*; 2014;66 (6): 1092–1099. doi:10.1016/j.pharep.2014.07.002
92. Barrau E, Fabre N, Fouraste I, Hoste H. Effect of bioactive compounds from sainfoin (*Onobrychis viciifolia* Scop.) on the in vitro larval migration of *Haemonchus contortus*: Role of tannins and flavonol glycosides. *Parasitology*; 2005;131: 531–538. doi:10.1017/S0031182005008024
93. Giovanelli F, Mattellini M, Fichi G, Flamini G, Perrucci S. In vitro anthelmintic activity of four plant-derived compounds against sheep gastrointestinal nematodes. *Veterinary Sciences*; 2018; 5: 78–85. doi: 10.3390/vetsci5030078
94. Chauhan K, Kaur G, Kaur S. Activity of rutin, a potent flavonoid against SSG-sensitive and -resistant *Leishmania donovani* parasites in experimental leishmaniasis. *Int. Immunopharmacol.*; 2018; 64: 372–385. doi:10.1016/j.intimp.2018.09.026
95. Azam F, Abodabos HS, Taban IM, Rfieda AR, Mahmood D, Anwar MJ, ... Ali HI. Rutin as promising drug for the treatment of Parkinson's disease: An assessment of MAO-B inhibitory potential by docking, molecular dynamics and DFT studies. *Molecular Simulation*; 2019; 45(18): 1563–1571. doi: 10.1080/08927022.2019.1662003
96. Lai X, Zhang Y, Wu J, Shen M, Yin S, Yan J. Rutin attenuates oxidative stress via PHB2-mediated mitophagy in MPP+-Induced SH-SY5Y cells. *Neurotoxicity Research*; 2023;41(3): 242–255. doi: 10.1007/s12640-023-00636-5
97. Elmazoglu Z, Ergin V, Sahin E, Kayhan H, Karasu C. Oleuropein and rutin protect against 6-OHDA-induced neurotoxicity in PC12 cells through modulation of mitochondrial function and unfolded protein response. *Interdisciplinary Toxicology*; 2017;10(4):129. doi:10.1515/intox-2017-0019
98. Magalingam KB, Radhakrishnan A, Haleagrahara N. Rutin, a bioflavonoid antioxidant protects rat pheochromocytoma (PC-12) cells against 6-hydroxydopamine (6-OHDA)-induced neurotoxicity. *International journal of molecular medicine*; 2013;32(1): 235–240. doi:10.3892/ijmm.2013.1375.
99. Zhang P, Gou Y, Gao X, Bai R, Chen W, Sun B, Hu F, Zhao, W. The pharmacokinetic study of rutin in rat plasma based on an electrochemically reduced graphene oxide modified sensor. *J. Pharm. Anal.*; 2016; 6(2): 80–86. doi:10.1016/j.jpha.2015.12.003
100. Zhang Y, Yin S, Song R, Lai X, Shen M, Wu J, Yan J. A novel mechanism of PHB2-mediated mitophagy participating in the development of Parkinson's disease. *Neural Regeneration Research*; 2024; 19(8): 1828–1834. doi: 10.4103/1673-5374.389356
101. Sălcudean A, Bodo CR, Popovici RA, Cozma MM, Păcurar M, Crăciun RE, Strete EG. Neuroinflammation. A Crucial Factor in the Pathophysiology of Depression. A Comprehensive Review. *Biomolecules*; 2025; 15(4): 502. doi: <https://doi.org/10.3390/biom15040502>
102. Schloms L, Smith C, Storbeck KH, Marnewick JL, Swart P, Swart AC. Rooibos influences glucocorticoid levels and steroid ratios in vivo and in vitro: a natural approach in themanagement

- of stress and metabolic disorders? Mol. Nutr. Food Res.; 2014; 58(3): 537–549. doi:10.1002/mnfr.201300463
103. Tongjaroenbuangam W, Ruksee N, Chantiratikul P, Pakdeenarong N, Kongbuntad W, Govitrapong P. Neuroprotective effects of quercetin, rutin and okra (*Abelmoschus esculentus* Linn.) in dexamethasone-treated mice. Neurochemistry international; 2011;59(5): 677-685. doi:10.1016/j.neuint.2011.06.014
 104. Pandian SRK, Pavadai P, Vellaisamy S, Ravishankar V, Palanisamy P, Sundar LM, ... & Kunjiappan S. Formulation and evaluation of rutin-loaded solid lipid nanoparticles for the treatment of brain tumor. Naunyn-Schmiedeberg's Archives of Pharmacology; 2021;394(4): 735-749. doi:10.1007/s00210-020-02015-9
 105. Afsar T, Khan MR, Razak S, Ullah S, Mirza B. Antipyretic, antiinflammatory and analgesic activity of *Acacia hydaspica* R. Parker and its phytochemical analysis. BMC Complement. Altern. Med.; 2015;15: 136. doi:10.1186/s12906-015-0658-8
 106. Ferreira, RS, Teles-Souza J, Dos Santos Souza C, Pereira ÉPL, de Araújo FM, da Silva AB, et al. Rutin improves glutamate uptake and inhibits glutamate excitotoxicity in rat brain slices. Mol. Biol. Rep.; 2021;48(2): 1475–1483. doi:10.1007/s11033-021-06145-y
 107. Zapata-Morales JR, Alonso-Castro AJ, González-Rivera ML, González Prado HI, Barragán-Gálvez JC, Hernández-Flores A, et al. Synergistic interaction between Justicia spicigera extract and analgesics on the formalin test in rats. Pharm.; 2025;18(2): 187. doi:10.3390/ph18020187
 108. Su KY, Yu CY, Chen YW, Huang YT, Chen CT, Wu HF, et al. Rutin, a flavonoid and principal component of saussurea involucre, attenuates physical fatigue in a forced swimming mouse model. Int. J. Med. Sci.; 2014;11(5): 528–537. doi:10.7150/ijms.8220
 109. Tian R, Yang W, Xue Q, Gao L, Huo J, Ren D, et al. Rutin ameliorates diabetic neuropathy by lowering plasma glucose and decreasing oxidative stress via Nrf2 signaling pathway in rats. Eur. J. Pharmacol; 2016; 771, 84–92. doi:10.1016/j.ejphar.2015.12.02
 110. Wilson RH, Mortarotti TG, Doxtader, EK. Toxicity studies on rutin. Proceedings of the Society for Experimental Biology and Medicine; 1947;64(3): 324-327. doi:10.3181/00379727-64-15781

CHAPTER 12

ANTHOCYANIN

Yasemin SEVİM¹

Introduction

Growing recognition of the potential adverse health implications associated with synthetic colorants, coupled with an expanding interest in the functional properties of naturally occurring pigments, has markedly increased the value of plant-derived colorants within the food, nutraceutical, and pharmaceutical industries (1). The term *anthocyanin*, derived from the Greek words *anthos* (flower) and *kyanos* (blue), reflects the extensive chromatic diversity that these compounds impart to plant tissues. As members of the flavonoid subclass, anthocyanins represent one of the most abundant and functionally consequential categories of secondary metabolites in higher plants. Within the broader family of polyphenolic compounds, anthocyanins are of particular scientific interest due to their substantial structural heterogeneity and the correspondingly wide range of physiological and biological processes they influence.

Anthocyanins are water-soluble vacuolar pigments that accumulate predominantly in fruits and floral tissues, although they are also detected in leaves, stems, roots, and other vegetative organs. They are responsible for the characteristic red, purple, and blue coloration of numerous horticultural and agricultural commodities, thereby playing a critical role in defining the sensory attributes and market value of these products. From a biological perspective, anthocyanins contribute to plant reproductive success by attracting pollinators and exert protective functions through the absorption of ultraviolet (UV) radiation.

¹ Dr., Giresun University, Faculty of Medicine, Department of Medicinal Biology, sevimyasemin61@gmail.com, ORCID iD: 0000-0002-2736-8898

Conclusion

Anthocyanins, characterized by their extensive structural diversity and wide-ranging biological activities, constitute a class of natural compounds with considerable potential for both health promotion and industrial applications. Their capacity to modulate oxidative stress, attenuate inflammation, regulate apoptotic pathways, and influence critical cellular signaling cascades underscores their promise as therapeutic agents for the prevention and management of various chronic conditions, including cardiovascular, neurodegenerative, metabolic, and cancer-related disorders. Nonetheless, limited bioavailability and chemical instability under physiological and environmental conditions continue to pose significant obstacles to their effective clinical application.

Recent technological advancements, including nanoformulations, microencapsulation, and other innovative delivery systems, have markedly improved the stability and bioavailability of anthocyanins, addressing key limitations in their clinical application. Moreover, accumulating evidence indicates that interactions between anthocyanins and the gut microbiota, as well as the biological activity of their metabolites, are critical determinants of their systemic effects. Beyond their established role as natural colorants, anthocyanins are increasingly incorporated into functional foods and nutraceutical products, highlighting their relevance as bioactive dietary constituents.

Future research should aim to elucidate the pharmacokinetics, biotransformation pathways, and tissue-specific molecular targets of anthocyanins. The development of standardized formulations, coupled with long-term clinical investigations, will be essential to fully establish their safety, efficacy, and therapeutic potential. In conclusion, anthocyanins are more than mere plant pigments; they are potent phytochemicals with diverse biological activities. Continued scientific and technological exploration is expected to broaden their applications across the food, health, and pharmaceutical sectors, thereby contributing meaningfully to strategies for disease prevention and the promotion of human health.

REFERENCES

1. De Mejia EG, Zhang Q, Penta K, Eroglu A, Lila MA (2020) The colors of health: chemistry, bioactivity, and market demand for colorful foods and natural food sources of colorants. *Annu Rev Food Sci Technol* 11:145–182
2. Qiu Z., Wang X., Gao J., Guo Y., Huang Z., Du Y. The tomato huffman's anthocyaninless gene encodes a bHLH transcription factor involved in anthocyanin biosynthesis that is developmentally regulated and induced by low temperatures. *PLoS ONE*. 2016;11:e0151067. doi: 10.1371/journal.pone.0151067.

3. Khoo H.E., Azlan A., Tang S.T., Lim S.M. Anthocyanidins and anthocyanins: Colored pigments as food, pharmaceutical ingredients, and the potential health benefits. *Food Nutr. Res.* 2017;61:1361779. doi: 10.1080/16546628.2017.1361779.
4. He J., Giusti M.M. Anthocyanins: Natural colorants with health-promoting properties. *Annu. Rev. Food Sci. Technol.* 2010;1:163–187. doi: 10.1146/annurev.food.080708.100754.
5. Khoo H., Ng H., Yap W.-S., Goh H., Yim H. Nutrients for prevention of macular degeneration and eye-related diseases. *Antioxidants.* 2019;8:85. doi: 10.3390/antiox8040085.
6. Castañeda-Ovando A., de Lourdes Pacheco-Hernández M., Páez-Hernández M.E., Rodríguez J.A., Galán-Vidal C.A. Chemical studies of anthocyanins: A review. *Food Chem.* 2009;113:859–871.
7. Gollop R, Even S, ColovaTsolova V, Perl A (2002) Expression of the grape dihydroflavonol reductase gene and analysis of its promoter region. *J Exp Bot* 53: 1397-1409.
8. de Pascual-Teresa S, Sanchez-Ballesta MT (2008) Anthocyanins: from plant to health. *Phytochem Rev* 7: 281-299.
9. 3. Harborne JB, Grayer RJ (1988) The anthocyanins. In *The flavonoids*, Springer, pp. 1-20.
10. Bridle P, Timberlake C (1997) Anthocyanins as natural food colours— selected aspects. *Food Chem* 58: 103-109
11. Nagy I.Z. On the true role of oxygen free radicals in the living state, aging, and degenerative disorders. *Ann. N.Y. Acad. Sci.* 2001; 928: 187-199.
12. Dröge W. Free radicals in the physiological control of cell function. *Physiol. Rev.* 2002; 82: 47-95
13. ueno, J.M.; Sáez-Plaza, P; Ramos-Escudero, F; Jiménez, A.M.; Fett, R.; Asuero, A.G. Analysis and Antioxidant Capacity of Anthocyanin Pigments. Part II: Chemical Structure, Color, and Intake of Anthocyanins. *Crit. Rev. Anal. Chem.* 2012, 42, 126–151.
14. Fukumoto, L.R.; Mazza, G. Assessing antioxidant and prooxidant activities of phenolic compounds. *J. Agric. Food Chem.* 2000, 48, 3597–3604.
15. Kay C.C., Mazza G., Holub B.J., Wang J. Anthocyanin metabolites in human urine and serum. *Brit. J. Nutr.* 2004; 91: 933-942
16. Joseph, S.V.; Edirisinghe, I.; Burton-Freeman, B.M. Berries: Anti-inflammatory Effects in Humans. *J. Agric. Food Chem.* **2014**, 62, 3886–3903.
17. Vendrame, S.; Klimis-Zacas, D. Anti-inflammatory effect of anthocyanins via modulation of nuclear factor- B and mitogen-activated protein kinase signaling cascades. *Nutr. Rev.* **2015**, 73, 348–358.
18. Wang H., Nair M.G., Strasburg G.M., Chang Y., Booren A.M., Gray J.I., DeWitt D.L. Antioxidant and anti-inflammatory activities of anthocyanins and their aglycon, cyanidin, from tart cherries. *J. Nat. Prod.* 1999; 62: 294–296
19. Galvano F, La Fauci L, Lazzarino G, Fogliano V, Ritieni A, Ciappellano S, Battistini N.C., Tavazzi B, Galvano G. Cyanidins: metabolism and biological properties. *J. Nutr. Biochem.* 2004; 15: 2-11.
20. Pedersen C.B., Kyle J., Jenkinson A.M., Gardner P.T., McPhail D.B., Duthie G.G. Effects of blueberry and cranberry juice consumption on the plasma antioxidant capacity of healthy female volunteers. *Eur. J. Clin. Nutr.* 2000; 54: 405-408.
21. Hou, D.X.; Kai, K.; Li, J.J.; Lin, S.; Terahara, N.; Wakamatsu, M.; Fujii, M.; Young, M.R.; Colburn, N. Anthocyanidins inhibit activator protein 1 activity and cell transformation: Structure-activity relationship and molecular mechanisms. *Carcinogenesis* 2004, 25, 29–36.
22. Min, S.W.; Ryu, S.N.; Kim, D.H. Anti-inflammatory effects of black rice, cyanidin-3-O-beta-D-glycoside, and its metabolites, cyanidin and protocatechuic acid. *Int. Immunopharmacol.* 2010, 10, 959–966.
23. Hou, D.X.; Yanagita, T.; Uto, T.; Masuzaki, S.; Fujii, M. Anthocyanidins inhibit cyclooxygenase-2 expression in LPS-evoked macrophages: Structure-activity relationship and molecular mechanisms involved. *Biochem. Pharmacol.* 2005, 70, 417–425.

24. Chen, L.; Teng, H.; Fang, T.; Xiao, J. Agrimonolide from *Agrimonia pilosa* suppresses inflammatory responses through down-regulation of COX-2/iNOS and inactivation of NF-kappaB in lipopolysaccharide-stimulated macrophages. *Phytomedicine* 2016, 23, 846–855.
25. Duarte, L.J.; Chaves, V.C.; Nascimento, M.; Calvete, E.; Li, M.; Ciraolo, E.; Ghigo, A.; Hirsch, E.; Simoes, C.M.O.; Reginatto, F.H.; et al. Molecular mechanism of action of Pelargonidin-3-O-glucoside, the main anthocyanin responsible for the anti-inflammatory effect of strawberry fruits. *Food Chem.* 2018, 247, 56–65.
26. Karunarathne, W.; Lee, K.T.; Choi, Y.H.; Jin, C.Y.; Kim, G.Y. Anthocyanins isolated from *Hibiscus syriacus* L. attenuate lipopolysaccharide-induced inflammation and endotoxic shock by inhibiting the TLR4/MD2-mediated NF-kappaB signaling pathway. *Phytomedicine* 2020, 76, 153237.
27. Wang LS, Hecht SS, Carmella SG, et al. Anthocyanins in black raspberries prevent esophageal tumors in rats. *Cancer Prev Res.* 2009;2(1):84–93.
28. Faria A, Pestana D, Teixeira D, et al. Blueberry anthocyanins and pyruvic acid adducts: anticancer properties in breast cancer cell lines. *Phytother Res.* 2010;24(12):1862–1869.
29. Lala G, Malik M, Zhao C, et al. Anthocyanin-rich extracts inhibit multiple biomarkers of colon cancer in rats. *Nutr Cancer.* 2006;54(1):84–93.
30. Malik M, Zhao C, Schoene N, et al. Anthocyanin-rich extract from *Aronia meloncarpa* E. induces a cell cycle block in colon cancer but not normal colonic cells. *Nutr Cancer.* 2003;46(2):186–196.
31. Lim S, Xu J, Kim J, et al. Role of anthocyanin-enriched purple-fleshed sweet potato p40 in colorectal cancer prevention. *Mol Nutr Food Res.* 2013;57(11):1908–1917.
32. Jang H, Ha US, Kim SJ, et al. Anthocyanin extracted from black soybean reduces prostate weight and promotes apoptosis in the prostatic hyperplasia-induced rat model. *J Agric Food Chem.* 2010;58:12686–12691.
33. Shin DY, Lee WS, Kim SH, et al. Anti-invasive activity of anthocyanins isolated from *Vitis coignetiae* in human hepatocarcinoma cells. *J Med Food.* 2009;12(5):967–972.
34. Bontempo P, de Masi L, Carafa V, et al. Anticancer activities of anthocyanin extract from genotyped *Solanum tuberosum* L. “Vitelotte”. *J Funct Foods.* 2015;19:584–593.
35. Takikawa M, Inoue S, Horio F, et al. Dietary anthocyanin-rich bilberry extract ameliorates hyperglycemia and insulin sensitivity via activation of AMP-activated protein kinase in diabetic mice. *J Nutr.* 2010;140(3):527–533.
36. Li D, Zhang Y, Liu Y, et al. Purified anthocyanin supplementation reduces dyslipidemia, enhances antioxidant capacity, and prevents insulin resistance in diabetic patients. *J Nutr.* 2015;145(4):742–748.
37. Kang MK, Lim SS, Lee JY, et al. Anthocyanin-rich purple corn extract inhibit diabetes-associated glomerular angiogenesis. *Plos One.* 2013;8(11):e79823.
38. Koh ES, Lim JH, Kim MY, et al. Anthocyanin-rich *Seoritae* extract ameliorates renal lipotoxicity via activation of AMP-activated protein kinase in diabetic mice. *J Transl Med.* 2015;13:203.
39. Liu Y, Li D, Zhang Y, et al. Anthocyanin increases adiponectin secretion and protects against diabetes-related endothelial dysfunction. *Am J Physiol Endocrinol Metab.* 2014;306(8):E975–E988.