

Bölüm 16

TÜMÖR MİKROÇEVRESİ

Elif POLAT¹

GİRİŞ

Tümör mikroçevresi (tumor microenvironment; TME), kanser dışı hücreler ve bunların stromalarından oluşan, kanserin büyümesini etkileyen önemli bir faktör olarak kabul edilir. Mikroçevre ve orada yer alan hücreler arasındaki çift yönlü iletişim, hem tümör büyümesi hem de normal doku homeostazı için önem arz eder¹. Özellikle, tümör hücreleri ve tümörle ilişkili stroma arasındaki etkileşimler, hastalığın başlangıcını, ilerlemesini ve prognozunu etkileyen güçlü bir ilişki ile sonuçlanır². Kanser, önceleri tümör hücrelerinde anormal mutasyonlar içeren heterojen bir hastalık olarak görülmesine rağmen, daha sonraları tümörlerin aynı zamanda mikro-çevresel oluşumlarının doğası ve stromal hücre oranları veya aktivasyon durumları gereği çeşitlilik gösterdiği bilinir^{3,4}.

Tümör ve mikroçevre arasındaki etkileşimlerle ilgili ilk çalışmalardan biri, Rudolph Virchow tarafından 1863 yılında lökosit infiltrasyonunun solid tümörlerde karakterize edildiğini ortaya koymasıyla gerçekleştirildi⁵. Steven Paget⁶ 1889'da meme kanseri metastazının organ tercihlerini analiz ederken, metastatik kolonizasyonun organın özelliklerine bağlı olduğunu göstermiştir. Bu prensip ayrıca primer tümörler ve mikroçevre arasındaki ilişkinin temelini oluşturur ve tümör evrimini etkiler⁷. Paget, bu nedenle tümör mikroçevresi kavramının öncüsü olarak bilinir⁸. TME araştırmaları 1970'lerden sonra artık tümör hücrelerinden türetilen sinyallere yanıt olarak TME'nin kanserin ilerlemesini aktif olarak etkilediği kabul edilir.

Tümör stroması, kan ve lenfatik dolaşım sistemleri, perisitler, fibroblastlar/miyofibroblastlar, glial, epitel, adiposit, vasküler, düz kas, bağışıklık hücrele-

¹ Dr. Öğr. Üyesi, Namık Kemal Üniversitesi Tıp Fakültesi Histoloji ve Embriyoloji Anabilim Dalı, epolat@nku.edu.tr, elifpol@gmail.com

zal membran, kollajenler, glikoproteinler, hiyalüronan, glikozaminoglikanlar, lamininler, proteoglikanlar, büyüme faktörleri ve kemokinlerinden oluşmaktadır. ECM hücreyel yapılar destek sağlar. Ayrıca ECM bileşimindeki değişikliklerin metastazda da önemli bir rol oynadığı düşünülmektedir¹⁰⁰.

TME'DE TERAPÖTİK YAKLAŞIMLAR

Kansere karşı birçok terapötik yaklaşım, tümör hücrelerinin çeşitli yönlerini hedeflemeyi amaçlar. Ayrıca TME'de bulunan stromal hücreler tümör hücrelerine göre genetik olarak stabildir. Bu yüzden klasik terapötik direnç mekanizmalarına karşı daha az duyarlı oldukları bilinir. Çeşitli anjiyogenez inhibitörleri ve stroma hücrelerini tüketmeyi hedefleyen tedaviler⁸⁸, TME'nin protümörjenik etkilerini ortadan kaldırdıkları için yararlı olamamaktadır. TME-yönelimli tedavilerin ortaya çıkan diğer bir örneği, tümör ile ilişkili kronik inflamasyonu nötralize etmeye yöneliktir¹⁰¹. Bu amaçla hedeflenen hücre tipleri arasında, CSF-1R, CCR2 ve CXCR2 sayılabilirken sinyal moleküllerinden de bağışıklık hücresi fonksiyonunu sağlayan NF- κ B yolağı veya anahtar sitokin yollarının inhibisyonunu bloke eden stratejilerden söz edilebilir¹⁰².

SONUÇ

Hücreler, yapısal proteinler ve sinyal moleküllerinden oluşan tümör stromasının; tümör başlangıcı, ilerlemesi ve metastazında merkezi bir rol oynadığı kabul edilmektedir. Bu nedenle tümör mikroçevresindeki kanser hücreleri ve stromal hücreler arasındaki etkileşimin belirlenmesi gelecekte kanser tedavisine büyük katkılar sağlayabilir.

Anahtar Kelimeler: Tümör mikroçevresi, stroma hücreleri, kanser, hücre dışı matriks, metastaz

KAYNAKLAR

1. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nature medicine*, 2013;19(11):1423.
2. Joyce JA, Pollard JW. Microenvironmental regulation of metastasis. *Nature Reviews Cancer*, 2009;9(4):239-52.
3. Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer cell*, 2012;21(3):309-22.
4. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *cell*, 2011;144(5):646-74.
5. Schmidt A, Weber O. In memoriam of Rudolf Virchow: a historical retrospective including aspects of inflammation, infection and neoplasia. *Infection and inflammation: impacts on oncogenesis*. 13: Karger Publishers; 2006. p. 1-15.
6. Paget S. The distribution of secondary growths in cancer of the breast. *Cancer Metastasis Rev*, 1989;8:98-101.
7. Witz IP, Levy-Nissenbaum O. The tumor microenvironment in the post-PAGET era. *Cancer*

- letters, 2006;242(1):1-10.
8. Jodele S, Blavier L, Yoon JM, et al. Modifying the soil to affect the seed: role of stromal-derived matrix metalloproteinases in cancer progression. *Cancer and Metastasis Reviews*, 2006;25(1):35-43.
 9. Tlsty TD, Coussens LM. Tumor stroma and regulation of cancer development. *Annu Rev Pathol Mech Dis*, 2006;1:119-50.
 10. Li H, Fan X, Houghton J. Tumor microenvironment: the role of the tumor stroma in cancer. *Journal of cellular biochemistry*, 2007;101(4):805-15.
 11. Witz IP. Tumor-microenvironment interactions: dangerous liaisons. *Advances in cancer research*, 2008;100:203-29.
 12. Szatrowski TP, Nathan CF. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer research*, 1991;51(3):794-8.
 13. Micke P. Tumour-stroma interaction: cancer-associated fibroblasts as novel targets in anti-cancer therapy? *Lung cancer*, 2004;45:S163-S75.
 14. Tomasek JJ, Gabbiani G, Hinz B, et al. Myofibroblasts and mechano-regulation of connective tissue remodelling. *Nature reviews Molecular cell biology*, 2002;3(5):349-63.
 15. Kalluri R, Zeisberg M. Fibroblasts in cancer. *Nature Reviews Cancer*, 2006;6(5):392-401.
 16. Clayton A, Evans RA, Pettit E, et al. Cellular activation through the ligation of intercellular adhesion molecule-1. *Journal of cell science*, 1998;111(4):443-53.
 17. Zeisberg M, Strutz F, Müller GA. Role of fibroblast activation in inducing interstitial fibrosis. *Journal of nephrology*, 2000;13:S111-20.
 18. Kojima Y, Acar A, Eaton EN, et al. Autocrine TGF- β and stromal cell-derived factor-1 (SDF-1) signaling drives the evolution of tumor-promoting mammary stromal myofibroblasts. *Proceedings of the National Academy of Sciences*, 2010;107(46):20009-14.
 19. Moskovits N, Kalinkovich A, Bar J, et al. p53 Attenuates cancer cell migration and invasion through repression of SDF-1/CXCL12 expression in stromal fibroblasts. *Cancer research*, 2006;66(22):10671-6.
 20. Aboussekhra A. Role of cancer-associated fibroblasts in breast cancer development and prognosis. *International Journal of Developmental Biology*, 2011;55(7-8-9):841-9.
 21. Kiaris H, Chatzistamou I, Trimis G, et al. Evidence for nonautonomous effect of p53 tumor suppressor in carcinogenesis. *Cancer research*, 2005;65(5):1627-30.
 22. Trimmer C, Sotgia F, Whitaker-Menezes D, et al. Caveolin-1 and mitochondrial SOD2 (Mn-SOD) function as tumor suppressors in the stromal microenvironment: a new genetically tractable model for human cancer associated fibroblasts. *Cancer biology & therapy*, 2011;11(4):383-94.
 23. Spaeth EL, Dembinski JL, Sasser AK, et al. Mesenchymal stem cell transition to tumor-associated fibroblasts contributes to fibrovascular network expansion and tumor progression. *PLoS one*, 2009;4(4).
 24. Zeisberg EM, Potenta S, Xie L, et al. Discovery of endothelial to mesenchymal transition as a source for carcinoma-associated fibroblasts. *Cancer research*, 2007;67(21):10123-8.
 25. Houghton J, Stoicov C, Nomura S, et al. Gastric cancer originating from bone marrow-derived cells. *Science*, 2004;306(5701):1568-71.
 26. Aractingi S, Kanitakis J, Euvrard S, et al. Skin carcinoma arising from donor cells in a kidney transplant recipient. *Cancer Research*, 2005;65(5):1755-60.
 27. Peters BA, Diaz LA, Polyak K, et al. Contribution of bone marrow-derived endothelial cells to human tumor vasculature. *Nature medicine*, 2005;11(3):261-2.
 28. Direkze NC, Hodivala-Dilke K, Jeffery R, et al. Bone marrow contribution to tumor-associated myofibroblasts and fibroblasts. *Cancer research*, 2004;64(23):8492-5.
 29. Rubio D, Garcia-Castro J, Martín MC, et al. Spontaneous human adult stem cell transformation. *Cancer research*, 2005;65(8):3035-9.
 30. Trimboli AJ, Cantemir-Stone CZ, Li F, et al. Pten in stromal fibroblasts suppresses mammary epithelial tumours. *Nature*, 2009;461(7267):1084-91.

31. Uccelli A, Moretta L, Pistoia V. Mesenchymal stem cells in health and disease. *Nature reviews immunology*, 2008;8(9):726-36.
32. Shi Y, Su J, Roberts AI, et al. How mesenchymal stem cells interact with tissue immune responses. *Trends in immunology*, 2012;33(3):136-43.
33. Quante M, Tu SP, Tomita H, et al. Bone marrow-derived myofibroblasts contribute to the mesenchymal stem cell niche and promote tumor growth. *Cancer cell*, 2011;19(2):257-72.
34. Le Blanc K, Rasmusson I, Sundberg B, et al. Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells. *The Lancet*, 2004;363(9419):1439-41.
35. Sun L, Wang D, Liang J, et al. Umbilical cord mesenchymal stem cell transplantation in severe and refractory systemic lupus erythematosus. *Arthritis & Rheumatism*, 2010;62(8):2467-75.
36. Vojtaššák J, Danišovič L, Kubeš M, et al. Autologous biograft and mesenchymal stem cells in treatment of the diabetic foot. *Neuroendocrinology Letters*, 2006;27(supplement 2):134-7.
37. Le Blanc K, Frassoni F, Ball L, et al. Mesenchymal stem cells for treatment of steroid-resistant, severe, acute graft-versus-host disease: a phase II study. *The Lancet*, 2008;371(9624):1579-86.
38. Wang Y, Chen X, Cao W, et al. Plasticity of mesenchymal stem cells in immunomodulation: pathological and therapeutic implications. *Nature immunology*, 2014;15(11):1009.
39. Ren G, Zhao X, Wang Y, et al. CCR2-dependent recruitment of macrophages by tumor-educated mesenchymal stromal cells promotes tumor development and is mimicked by TNF α . *Cell stem cell*, 2012;11(6):812-24.
40. Shah K. Mesenchymal stem cells engineered for cancer therapy. *Advanced drug delivery reviews*, 2012;64(8):739-48.
41. Stagg J. Mesenchymal stem cells in cancer. *Stem cell reviews*, 2008;4(2):119-24.
42. Jung Y, Kim JK, Shiozawa Y, et al. Recruitment of mesenchymal stem cells into prostate tumours promotes metastasis. *Nature communications*, 2013;4(1):1-11.
43. Dwyer R, Potter-Beirne S, Harrington K, et al. Monocyte chemotactic protein-1 secreted by primary breast tumors stimulates migration of mesenchymal stem cells. *Clinical Cancer Research*, 2007;13(17):5020-7.
44. Gao H, Priebe W, Glod J, et al. Activation of signal transducers and activators of transcription 3 and focal adhesion kinase by stromal cell-derived factor 1 is required for migration of human mesenchymal stem cells in response to tumor cell-conditioned medium. *Stem cells*, 2009;27(4):857-65.
45. Spaeth E, Klopp A, Dembinski J, et al. Inflammation and tumor microenvironments: defining the migratory itinerary of mesenchymal stem cells. *Gene therapy*, 2008;15(10):730-8.
46. Grisendi G, Spano C, Rossignoli F, et al. Tumor stroma manipulation by MSC. *Current drug targets*, 2016;17(10):1111-26.
47. Brennen WN, Rosen DM, Wang H, et al. Targeting carcinoma-associated fibroblasts within the tumor stroma with a fibroblast activation protein-activated prodrug. *Journal of the National Cancer Institute*, 2012;104(17):1320-34.
48. Kidd S, Spaeth E, Watson K, et al. Origins of the tumor microenvironment: quantitative assessment of adipose-derived and bone marrow-derived stroma. *PloS one*, 2012;7(2).
49. Serafini P, De Santo C, Marigo I, et al. Derangement of immune responses by myeloid suppressor cells. *Cancer Immunology, Immunotherapy*, 2004;53(2):64-72.
50. Lin P, DeBusk L, Huang J, et al. Expansion of myeloid immune suppressor cells in Tumor-bearing host directly promotes tumor angiogenesis, tumor growth, and metastasis. *AACR*; 2007.
51. Coussens LM, Raymond WW, Bergers G, et al. Inflammatory mast cells up-regulate angiogenesis during squamous epithelial carcinogenesis. *Genes & development*, 1999;13(11):1382-97.
52. Cimpean AM, Tamma R, Ruggieri S, et al. Mast cells in breast cancer angiogenesis. *Critical reviews in oncology/hematology*, 2017;115:23-6.
53. Lewis CE, Pollard JW. Distinct role of macrophages in different tumor microenvironments. *Cancer research*, 2006;66(2):605-12.
54. Qian B-Z, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell*, 2010;141(1):39-51.

55. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. *Nature reviews immunology*, 2008;8(12):958-69.
56. Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nature immunology*, 2010;11(10):889.
57. Lewis C, Murdoch C. Macrophage responses to hypoxia: implications for tumor progression and anti-cancer therapies. *The American journal of pathology*, 2005;167(3):627-35.
58. Goede V, Brogelli L, Ziche M, et al. Induction of inflammatory angiogenesis by monocyte chemoattractant protein-1. *International Journal of Cancer*, 1999;82(5):765-70.
59. Kadhim SA, Rees RC. Enhancement of tumor growth in mice: evidence for the involvement of host macrophages. *Cellular immunology*, 1984;87(1):259-69.
60. Kreider JW, Bartlett GL, Butkiewicz BL. Relationship of tumor leucocytic infiltration to host defense mechanisms and prognosis. *Cancer and Metastasis Reviews*, 1984;3(1):53-74.
61. Mantovani A, Marchesi F, Malesci A, et al. Tumour-associated macrophages as treatment targets in oncology. *Nature reviews Clinical oncology*, 2017;14(7):399.
62. Gabrilovich DI, Bronte V, Chen S-H, et al. The terminology issue for myeloid-derived suppressor cells. *Cancer research*, 2007;67(1):425-.
63. Ostrand-Rosenberg S, Sinha P. Myeloid-derived suppressor cells: linking inflammation and cancer. *The Journal of Immunology*, 2009;182(8):4499-506.
64. Ségaliny AI, Mohamadi A, Dizier B, et al. Interleukin-34 promotes tumor progression and metastatic process in osteosarcoma through induction of angiogenesis and macrophage recruitment. *International journal of cancer*, 2015;137(1):73-85.
65. Marigo I, Dolcetti L, Serafini P, et al. Tumor-induced tolerance and immune suppression by myeloid derived suppressor cells. *Immunological reviews*, 2008;222(1):162-79.
66. Ostrand-Rosenberg S. Immune surveillance: a balance between protumor and antitumor immunity. *Current opinion in genetics & development*, 2008;18(1):11-8.
67. Gregory AD, Houghton AM. Tumor-associated neutrophils: new targets for cancer therapy. *Cancer research*, 2011;71(7):2411-6.
68. Wikberg ML, Ling A, Li X, et al. Neutrophil infiltration is a favorable prognostic factor in early stages of colon cancer. *Human pathology*, 2017;68:193-202.
69. Thurnher M, Radmayr C, Ramoner R, et al. Human renal-cell carcinoma tissue contains dendritic cells. *International journal of cancer*, 1996;68(1):1-7.
70. Zong J, Keskinov AA, Shurin GV, et al. Tumor-derived factors modulating dendritic cell function. *Cancer Immunology, Immunotherapy*, 2016;65(7):821-33.
71. Vose B, Vanky F, Argov S, et al. Natural cytotoxicity in man: activity of lymph node and tumor-infiltrating lymphocytes. *European journal of immunology*, 1977;7(11):753-7.
72. Vose BM, Moore M, editors. *Human tumor-infiltrating lymphocytes: a marker of host response*. Seminars in hematology; 1985.
73. Chiou S-H, Sheu B-C, Chang W-C, et al. Current concepts of tumor-infiltrating lymphocytes in human malignancies. *Journal of reproductive immunology*, 2005;67(1-2):35-50.
74. Fridman WH, Pagès F, Sautès-Fridman C, et al. The immune contexture in human tumours: impact on clinical outcome. *Nature Reviews Cancer*, 2012;12(4):298-306.
75. Hsieh C-S, Lee H-M, Lio C-WJ. Selection of regulatory T cells in the thymus. *Nature Reviews Immunology*, 2012;12(3):157-67.
76. Campbell DJ, Koch MA. T reg cells: patrolling a dangerous neighborhood. *Nature medicine*, 2011;17(8):929-30.
77. Bates GJ, Fox SB, Han C, et al. Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse. *Journal of Clinical Oncology*, 2006;24(34):5373-80.
78. De Visser KE, Korets LV, Coussens LM. De novo carcinogenesis promoted by chronic inflammation is B lymphocyte dependent. *Cancer cell*, 2005;7(5):411-23.
79. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. *Nature*, 2011;480(7378):480-9.

80. Schioppa T, Moore R, Thompson RG, et al. B regulatory cells and the tumor-promoting actions of TNF- α during squamous carcinogenesis. *Proceedings of the National Academy of Sciences*, 2011;108(26):10662-7.
81. Olkhanud PB, Damdinsuren B, Bodogai M, et al. Tumor-evoked regulatory B cells promote breast cancer metastasis by converting resting CD4+ T cells to T-regulatory cells. *Cancer research*, 2011;71(10):3505-15.
82. Vivier E, Raulet DH, Moretta A, et al. Innate or adaptive immunity? The example of natural killer cells. *Science*, 2011;331(6013):44-9.
83. Langers I, Renoux VM, Thiry M, et al. Natural killer cells: role in local tumor growth and metastasis. *Biologics: targets & therapy*, 2012;6:73.
84. Sceneay J, Chow MT, Chen A, et al. Primary tumor hypoxia recruits CD11b+/Ly6Cmed/Ly6G+ immune suppressor cells and compromises NK cell cytotoxicity in the premetastatic niche. *Cancer research*, 2012;72(16):3906-11.
85. Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science*, 2005;307(5706):58-62.
86. Dvorak H. Angiogenesis: update 2005. *Journal of Thrombosis and Haemostasis*, 2005;3(8):1835-42.
87. Ribatti D, Vacca A, Dammacco F. New non-angiogenesis dependent pathways for tumour growth. *European Journal of Cancer*, 2003;39(13):1835-41.
88. Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. *Nature*, 2011;473(7347):298-307.
89. O'Keeffe MB, Devlin AH, Burns AJ, et al. Investigation of pericytes, hypoxia, and vascularity in bladder tumors: association with clinical outcomes. *Oncology Research Featuring Preclinical and Clinical Cancer Therapeutics*, 2008;17(3):93-101.
90. Nieman KM, Kenny HA, Penicka CV, et al. Adipocytes promote ovarian cancer metastasis and provide energy for rapid tumor growth. *Nature medicine*, 2011;17(11):1498.
91. Frantz C, Stewart KM, Weaver VM. The extracellular matrix at a glance. *Journal of cell science*, 2010;123(24):4195-200.
92. Weigelt B, Bissell MJ, editors. *Unraveling the microenvironmental influences on the normal mammary gland and breast cancer*. Seminars in cancer biology; 2008: Elsevier.
93. Levental KR, Yu H, Kass L, et al. Matrix crosslinking forces tumor progression by enhancing integrin signaling. *Cell*, 2009;139(5):891-906.
94. Edovitsky E, Elkin M, Zcharia E, et al. Heparanase gene silencing, tumor invasiveness, angiogenesis, and metastasis. *Journal of the national cancer institute*, 2004;96(16):1219-30.
95. Pezzuto A, Carico E. Role of HIF-1 in cancer progression: Novel insights. A review. *Current molecular medicine*, 2018;18(6):343-51.
96. Semenza GL. The hypoxic tumor microenvironment: A driving force for breast cancer progression. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 2016;1863(3):382-91.
97. Valastyan S, Weinberg RA. Tumor metastasis: molecular insights and evolving paradigms. *Cell*, 2011;147(2):275-92.
98. Koh BI, Kang Y. The pro-metastatic role of bone marrow-derived cells: a focus on MSCs and regulatory T cells. *EMBO reports*, 2012;13(5):412-22.
99. Dayan F, Mazure NM, Brahimi-Horn MC, et al. A dialogue between the hypoxia-inducible factor and the tumor microenvironment. *Cancer Microenvironment*, 2008;1(1):53-68.
100. Gilkes DM, Semenza GL, Wirtz D. Hypoxia and the extracellular matrix: drivers of tumour metastasis. *Nature Reviews Cancer*, 2014;14(6):430-9.
101. Coussens LM, Zitvogel L, Palucka AK. Neutralizing tumor-promoting chronic inflammation: a magic bullet? *Science*, 2013;339(6117):286-91.
102. DeNardo DG, Brennan DJ, Rexhepaj E, et al. Leukocyte complexity predicts breast cancer survival and functionally regulates response to chemotherapy. *Cancer discovery*, 2011;1(1):54-67.