

BÖLÜM 6

MAŞ FASULYESİNDE MUTASYON ISLAHININ ÖNEMİ VE UYGULAMALARI

Cengiz TÜRKEY¹
İlknur AKGÜN²
Ruziye KARAMAN³

GİRİŞ

Tarım, iklim değişikliği, nüfus artışı, arazilerin küçülmesi ve doğal kaynakların tükenmesi gibi etmenlerin birleşimiyle, küresel gıda ve beslenme güvenliğini tehdit eden ciddi zorluklar altındadır. Bu bağlamda bitki ıslahı, verimliliği sürdürülebilir bir şekilde artırmak, kalite özelliklerini iyileştirmek, biyotik ve abiyotik streslere karşı dayanıklılığı artırmak için temel bir strateji olmaya devam etmektedir (1, 2). Geleneksel ıslah yaklaşımları, temel teşkil etse de, özellikle kendine tozlaşan bitkilerde ve gen havuzları kısıtlı olan baklagillerde, mevcut genetik çeşitliliğe ve uzun ıslah döngülerine olan bağımlılıkları nedeniyle sınırlı düzeyde kalmaktadır (3). Ayrıca, geleneksel melezleme ve seleksiyon yöntemlerinin etkinliği, genetik çeşitliliği dar olan veya doğal rekombinasyonun sınırlı olduğu bitkilerde azalma eğilimindedir. Bu durum, yeni ve hedefe yönelik genetik varyasyonlar üretebilen alternatif stratejilerin geliştirilmesine olanak sağlamaktadır (4). Mutasyon ıslahı, genel genomik bütünlüğü koruyarak üstün ve istenen özelliklerin kazandırılmasına olanak tanıyan; aynı zamanda gizli

¹ Arş. Gör., Isparta Uygulamalı Bilimler Üniversitesi, Ziraat Fakültesi, Tarla Bitkileri Bölümü
cengizturkay@isparta.edu.tr, ORCID iD: 0000-0003-3857-0140

² Prof. Dr., Isparta Uygulamalı Bilimler Üniversitesi, Ziraat Fakültesi, Tarla Bitkileri Bölümü
ilknurakgun@isparta.edu.tr, ORCID iD: 0000-0002-7476-7226

³ Doç. Dr., Isparta Uygulamalı Bilimler Üniversitesi, Ziraat Fakültesi, Tarla Bitkileri Bölümü
ruziyekaraman@isparta.edu.tr, ORCID iD: 0000-0001-5088-8253

genel genetik altyapısını bozmadan erkencilik, hastalık direnci ve abiyotik stres toleransı gibi belirli özelliklerin hedefli olarak iyileştirilmesini sağlamaktadır. Son yıllarda, gama ışınları ve EMS gibi mutajenler kullanılarak maş fasulyesinde çok sayıda başarılı mutant hattı geliştirilmiş ve bu yöntem, hem verim hem de kalite kısıtlamalarını ele alma potansiyelini göstermiştir. Ancak bu potansiyel, çeşitli biyolojik, teknik ve kurumsal zorluklarla dengelenmektedir. Bunlar arasında, yararlı mutasyonların tespiti ve mutajen dozlarının optimize edilmesi, seleksiyon aşamalarındaki ayrıntılı incelemeler ve mutantların ticari çeşit olarak kullanılmasına kadar geçen sürede karşılaşılan zorluklar yer almaktadır.

Maş fasulyesinde mutasyon ıslahının geleceği, genotipik, fenotipik ve dijital tarımla stratejik entegrasyonuna bağlıdır. Markör destekli tarama yöntemleri, mutasyona uğrayan alellerin hassas bir şekilde tanımlanmasını ve doğrulanmasını sağlayacaktır. Mutasyon ıslahını yabancı akrabaların zengin çeşitliliğiyle birleştirmek ıslahçıların kullanabileceği genetik tabanı daha da genişletecektir. Mutasyon ıslahı yalnızca geleneksel ve transgenik ıslah yöntemlerine pratik bir alternatif değil, aynı zamanda yemeklik baklagil ıslahında önemli bir stratejidir. Etkili bir şekilde kullanıldığında, iklim dostu tarımın ve küresel gıda güvenliğinin değişen taleplerini karşılayan dayanıklı, yüksek verimli ve besin değeri artırılmış maş fasulyesi çeşitlerinin geliştirilmesine önemli ölçüde katkıda bulunabilir.

KAYNAKLAR

1. IAEA (2008). Induced Mutations in Plants. Proceedings of a Joint FAO/IAEA Symposium, International Atomic Energy Agency, Vienna.
2. Penna S, Jain SM. *Mutation Breeding for Sustainable Food Production and Climate Resilience*. Singapore: Springer Nature Singapore, 2023.
3. Solanki RK, Gill RK, Verma P, Singh S. *Mutation breeding in pulses: an overview*. Breeding of pulse crops. Ludhiana: Kalyani Publishers, 2011. p. 85-103.
4. Kharkwal MC. *Mutation breeding for crop improvement*. In: Jain HK, Kharkwal MC (Eds) Plant Breeding - Mendelian to Molecular Approaches. New Delhi, India, Narosa Publishing House, 2004. P. 601-645
5. Auti SG. Induced morphological and quantitative mutations in mungbean. *Bioremediation, Biodiversity and Bioavailability*, 2012; 6(1): 27-39.
6. Javed I, Ahsan M, Ahmad H M, Ali Q. Role of mutation breeding to improve mungbean (*Vigna radiata* L. Wilczek) yield: An overview. *Nature and Science*. 2016;14(1): 63-77. doi:10.7537/marsnsj140116.09
7. Kumar R, Sharma NK, Meena S, Balakrishnan AP. Mutation breeding: A way forward for genetic improvement in mungbean. *International Journal of Plant and Environment*. 2021; 7(4): 255-262.
8. Ghotbi V, Mahrokh A, Tehrani AM, Asadi H. Evaluation of forage yield and quality of cowpea, guar, and mung bean under drought stress conditions. *Chemistry Proceedings*. 2022; 10(1): 62.

9. Akbay F, Günaydın T, Kızılyar EN, vd. İkinci ürün koşullarında mısır-maş fasulye üretiminde farklı ekim sistemlerinin silaj kalitesi ve fermentasyonuna etkisi. *ISPEC Tarım Bilimleri Dergisi*. 2024; 8 (4): 992-1000.
10. Imran İ, Khan, AA, Inam I, et al. Yield and yield attributes of Mungbean (*Vigna radiata* L.) cultivars as affected by phosphorous levels under different tillage systems. *Cogent Food & Agriculture*. 2016; 2(1): 1151982.
11. Sangsiri C, Worawit S, Peerasak S. Gamma radiation induced mutations in mungbean. *Science Asia*. 2005; 31: 251-255
12. Khan S, Goyal S. Mutation genetic studies in mungbean IV. Selection of early maturing mutants. *Thai Journal of Agricultural Science*. 2009; 42(2): 109-113
13. Muller HJ. Artificial transmutation of the gene. *Science*. 1927; 66(1699): 84-87.
14. Maluszynski M, Nichterlein K, Van Zanten L, et al. Officially released mutant varieties-the FAO/IAEA Database. 2000.
15. Jankowicz-Cieslak J, Mba C, Till BJ. Mutagenesis for crop breeding and functional genomics. *Biotechnologies for Plant Mutation Breeding*. 2017; 30: 3-18.
16. Ahloowalia BS, Maluszynski M, Nichterlein K. Global impact of mutation-derived varieties. *Euphytica*. 2004; 135(2): 187-204.
17. Stadler LJ. Chromosome number and the mutation rate in *Avena* and *Triticum*. *Proceedings of the National Academy of Sciences*. 1929; 15(12): 876-881.
18. Shu QY, Forster BP, Nakagawa H, Nakagawa H.. *Plant mutation breeding and biotechnology*. Cabi. 2012.
19. Henikoff S, Till BJ, Comai L. *Traditional mutagenesis meets functional genomics*. *Plant Physiology*, 2004; 135(2): 630-636.
20. Hasan N, Choudhary S, Joshi DC et al. A comparative study of mutagenic efficacy of EMS and MMS in inducing quantitative and cytological variations in *Capsicum annuum* L. var. NS1101. *Discover Plants*, 2025; 2(1): 184.
21. Sikora P, Chawade A, Larsson M, et al. Mutagenesis as a tool in plant genetics, functional genomics, and breeding. *International Journal of Plant Genomics*. 2011; (1): 314829.
22. Konzak CF, Nilan RA, Wagner J., et al. Efficient chemical mutagenesis. 1965.
23. Dhoooghe E, Van Laere K, Eeckhaut T, Leus L, Van Huylenbroeck J. Mitotic chromosome doubling of plant tissues in vitro. *Plant Cell, Tissue and Organ Culture*. 2011; 104(3), 359-373. <https://doi.org/10.1007/s11240-010-9786-5>
24. Roy P, Mandal A, Das D. A review on colchicine-induced polyploidy in legume crops. *Legume Research*. 2021; 44(7): 778-785. <https://doi.org/10.18805/LR-4895>
25. Wongpiyasatid A, Chotechuen S, Hormchan P, et al. Induced mutations in mungbean breeding: regional yield trial of mungbean mutant lines. *Agriculture and Natural Resources*. 2000; 34(4): 443-449.
26. Tah PR. Induced synchrony in pod maturity in mungbean [*Vigna radiata* (L.) Wilczek]. *ARPN Journal of Agricultural and Biological Science*. 2009; 4(1): 41-44.
27. Sahu BC, Patra G. Evaluation of quantitative characteristics in some mutant lines of green gram. *Indian Journal of Agricultural Sciences*. 1997; 67: 533- 535
28. Lal N, Mishra R. Induced genetic variability and divergence in M2 generation of mungbean. *Indian Journal of Pulses Research*. 2006; 19 (1): 47-49
29. Yaqoob M, Rashid A. Induced mutation studies in some mungbean (*Vigna radiata* L. Wilczek). *Journal of Biological Sciences*. 2001; 1: 805-808
30. Sarwar G, Ahmed M. Development of new high yielding mungbean variety "AEM 96" through induced mutations. *SAARC Journal of Agriculture*. 2003; 1: 173-180
31. Tah PR. Induced macromutation in mungbean [*Vigna radiata* (L.) Wilczek]. *International Journal of Botany*. 2006; 2(3): 219-228.

32. Khan S, Wani M. Induced mutations affecting quantitative characters in mungbean. *Agricultural Science Digest*. 2006; 4: 241-244
33. Auti SG, Barshile JD, Dalave SC et al. Frequency and spectrum of chlorophyll mutants in mungbean. *Journal of Food Legumes*. 2007; 20(2): 156-157.
34. Singh A. Induced genetic variability in M3 generation of mungbean. *Journal of Food Legumes*. 2009;22(3):162-165
35. Singh A, Kumar D. Genetic parameters and co-efficient analysis in M4 generation of mungbean (*Vigna radiata* L. Wilczek). *Journal of Food Legumes*. 2009;22 (3): 166-170
36. Kumar A, Parmhansh P, Prasad R. Induced chlorophyll and morphological mutations in mungbean (*Vigna radiata* L. Wilczek). *Legume Research*. 2009; 32(1): 41-45
37. Auti SG, Apparao BJ. *Induced mutagenesis in mungbean (Vigna radiata (L.) Wilczek)*. In: Shu Q-Y (Ed) *Induced Plant Mutations in the Genomics Era*, Food and Agriculture Organization of the United Nations, Rome, 2009, p. 97- 100.
38. Ngampongsai, S., Watanasit, A., Srinives, P et al. Current status of mungbean and the use of mutation breeding in Thailand. In: Shu Q-Y (Ed) *Induced Plant Mutations in the Genomics Era*, Food and Agriculture Organization of the United Nations, Rome, 2009, p. 355-357.
39. Reddy KS. *A new mutant for yellow mosaic virus resistant in mungbean (Vigna radiata L. Wilczek) variety SML 668 by recurrent gamma irradiation*. In: Shu Q-Y (Ed) *Induced Plant Mutations in the Genomics Era*, Food and Agriculture Organization of the United Nations, Rome, 2009, p. 361-362
40. Balai OP, Krishna KR. Efficiency and effectiveness of chemical mutagens in mungbean. *Journal of Food Legumes*. 2009; 22(2): 105-108.
41. Dhole VJ, Reddy KS. Gamma rays induced moisture stress tolerant long root mutant in mungbean (*Vigna radiata* L. Wilczek). *Electronic Journal of Plant Breeding*. 2010;1(5): 1299-1305
42. Kozgar MI, Goyal S, Khan S. EMS induced mutational variability in *Vigna radiata* and *Vigna mungo*. *Research Journal of Botany*. 2011; 6 (1): 31-37
43. Lavanya R, Yadav L, Suresh B et al. Sodium azide mutagenic effect on biological parameters and induced genetic variability in mungbean. *Journal of Food Legumes*. 2011; 24 (1): 46-49
44. Bado S, Forster BP, Nielen S et al. Plant mutation breeding: current progress and future assessment. *Plant Breeding Reviews*. 2015; 39: 23-88.
45. Yali W, Mitiku T. Mutation breeding and its importance in modern plant breeding. *Journal of Plant Sciences*. 2022; 10(2): 64-70.
46. Gandhi S, Umavath, S, Mullainathan L. Studies on induced chlorophyll mutants in green gram (*Vigna radiata* (L.) Wilczek). *Int J of Adv Res*. 2014; 2: 00-04.
47. Raina A, Laskar RA, Tantray YR, et al. Characterization of induced high yielding cowpea mutant lines using physiological, biochemical and molecular markers. *Scientific Reports*. 2020; 10(1): 3687.
48. Wani MR. Characterization of chlorophyll deficient mutants in mungbean (*Vigna radiata* (L.) Wilczek). *Bangladesh Journal of Botany*. 2020; 49(4): 1013-1019.
49. Dhole VJ, Souframanien J, Reddy KS, et al. Comparison of effectiveness and efficiency of electron beam over gamma rays to induce novel mutations in mungbean (*Vigna radiata* L. Wilczek). *Applied Radiation and Isotopes*. 2023; 194: 110719.
50. Kumar S. Gamma radiation induced mutations in mungbean (*Vigna radiata* (L.) Wilczek). *Scholarly Journal of Agricultural Science*. 2014;4:240-243
51. Girija M, Dhanavel D, Gnanamurthy S. Gamma rays and EMS induced flower color and seed mutants in cowpea (*Vigna unguiculata* L. Walp). *Advances in Applied Science Research*. 2013; 4:134-139.

52. Rahevar PM, Chauhan RM, Patel PT, et al. Isolation and evaluation of novel male sterile and self-incompatible mutant lines of mungbean (*Vigna radiata* L.). *Plant Breeding*. (2024); 143(5): 666-674.
53. Usharani KS, Kumar CA. Mutagenic efficiency and effectiveness of gamma rays and EMS and their combination in inducing chlorophyll mutations in M2 generation of Urdbean (*Vigna mungo* (L.) Hepper). *Electronic Journal of Plant Breeding*. 2015; 6(1): 210-217.
54. Roslim DI, Fiatin ISRO. Lethal dose 50 (LD 50) of mungbean (*Vigna radiata* L. Wilczek) cultivar kampar. *SABRAO Journal of Breeding & Genetics*. 2015; 47(4).
55. Smith, GF, Kerstein H. Auxins and calines in seedlings from X-rayed seeds. *American Journal of Botany*. 1942; 29: 785-792
56. Evans HJ, Sparrow AH. Nuclear factors affecting radiosensitivity II. Dependence on nuclear and chromosome structure and organization. *Brook haven Symposia in Biology*. 1961; 14: 101-127
57. Pele SK, Howard A. Effect of various doses of X-rays on the number of cells synthesizing DNA. *Radiation Research*. 1955; 3: 135-142
58. Stein OL, Sparrow AH. *Effect of chronic gamma radiation on the growth of Kalanchoe cv. Brilliant Star*. *Radiation Botany*. 1963; 3: 207-222
59. Evans HJ. Effects of radiations on meristematic cells. *Radiation Botany* 1965; 5: 171-182.
60. Webber E, Gottschalk W. The relation between cell size and internode length in induced mutants of *Pisum sativum*. *Beitrage zur Biologie der Pflanzen*. 1973; 49: 101-126
61. Blonstein AD, Gale MD. *Cell size and cell number in dwarf barley and semidwarf cereal mutants and their use in cross breeding II (Teidse 407)*. FAO/IAEA, Vienna, 1984, p. 19-29
62. Bhatia CR, Nichterlein K, Maluszynsky M. Oilseed cultivars developed from induced mutations and mutations altering fatty acid composition. *Mutations Breeding Review*, 1999; 11: 1-36
63. Yadav RDS. Effect of mutagens on mitotic index, seedling vigour and chlorophyll mutations in mungbean. *Journal of Nuclear and Agricultural Biology*. (1987); 16: 13-17
64. Khan IA. Mutation studies in mung bean (*Vigna radiata* (L.) Wilczek) iv. Increased variability in quantitative characters. *Journal of Bangladesh Academy of Sciences*. 1989; 13(1): 41.
65. Subramanian D. Effect of gamma radiation in Vigna. *Indian Journal of Genetics and Plant Breeding*. 1980; 40: 187-194
66. Sharma R, Singh VP. The influence of altered moisture levels in seeds on the induction of mutations with EMS and gamma rays in mungbean. *Journal of the Indian Botanical Society*. 1992; 71: 125-128
67. Bhal JR, Gupta PK. Chlorophyll mutations in mungbean (*Vigna radiata* (L) Wilczek). *Theoretical and Applied Genetics*. 1982; 63: 23-26
68. Kharakwal MC. *Accomplishment of mutation breeding in crop improvement in India*. In: Sachdev MS, Sachdev P, Deb DL (Eds) *Isotopes and Radiations in Agriculture and Environment Research*, Indian Society of Nuclear Techniques in Agriculture and Biology, New Delhi, India, 1996, p. 196-218
69. Ahmed ZU, Shaikh MAQ, & Begum S. *Development of the high yielding and disease resistant mungbean variety, BINA Moong-2 using nuclear and cross breeding techniques*. In: *Proceedings of International Atomic Energy Agency and the Food and Agriculture Organization. Symposium on Induced Mutations and Molecular Techniques for Crop Improvement*, Vienna, June 19-23, 1995, p. 638-640.
70. IAEA. *Physical Mapping Technologies for the Identification and Characterization of Mutated Genes Contributing to Crop Quality* (IAEA-TECDOC-1664). Vienna: IAEA, 2011.

71. Kharkwal MC. Induced mutation in chickpea (*Cicer arietinum* L.) V. Evaluation of micro-mutation. *Indian Journal of Genetics*. 2001;61: 115-124
72. Awan MA. Use of induced mutations for crop improvement in Pakistan. In: Proceedings of a Symposium on Plant Mutation Breeding for Crop Improvement, International Atomic Energy Agency and Food and Agriculture Organization, Vienna, Austria, 1991, 18-22,
73. Ali M, Aslam MM, Jaffar MAB. Selection of mungbean (*Vigna radiata* L.) mutants with respect to seasonal variation of summer and spring using discriminant function analysis. *Heliyon*. 2024; 10(10).
74. Wani MR, Khan S. Estimates of genetic variability in mutated populations and the scope of selection for yield attributes in *Vigna radiata* (L.) Wilczek. *Egyptian Journal of Biology*. 2006; 8: 1-6
75. Afzal MA, Bakr MA, Luna NK, et al. Registration of 'Barimung-5' mungbean. *Crop Science*. 2003; 43: 2304-2305.
76. Chen Y, Nelson RL. Identification and characterization of a white-flowered wild soybean plant. *Crop Science*. 2004; 44: 339-342
77. Hamid A, Afzal M, Haque M, et al. Registration of 'BUMug-1' mungbean. *Crop Science*. 2004; 44: 1489-1489.
78. Corbesier L, Gadisseur I, Silvestre G, et al. Design in *Arabidopsis thaliana* of a synchronous system of floral induction by one long day. *The Plant Journal*. 2003; 9: 947-952
79. Bushby H, Lawn R. Accumulation and partitioning of nitrogen and dry matter by contrasting genotypes of mungbean (*Vigna radiata* (L.) Wilczek). *Australian Journal of Agricultural Research*. 1992; 43, 1609-1628
80. Bisht I, Mahajan R, Patel D. The use of characterization data to establish the Indian mungbean core collection and assessment of genetic diversity. *Genetic Resources and Crop Evolution*. (1998; 45: 127-133
81. Ahloowalia BS, Maluszynski M. Induced mutations—A new paradigm in plant breeding. *Euphytica*. 2001; 118(2): 167-173.
82. Kumar R, Kumar R, Goel H, et al. Whole exome sequencing identifies novel variants of PIK3CA and validation of hotspot mutation by droplet digital PCR in breast cancer among Indian population. *Cancer Cell International*, 2023; 23(1): 236.
83. Aklilu, E. (2021). Review on forward and reverse genetics in plant breeding. *All Life*, 14(1), 127-135.
84. Pathak D, Leroy AK, Thompson TA, et al. A two-component probability distribution function describes the mid-IR emission from the disks of star-forming galaxies. *The Astronomical Journal*. (2023); 167(1): 39.
85. Ali A, Altaf MT, Nadeem MA, et al. Recent advancement in OMICS approaches to enhance abiotic stress tolerance in legumes. *Frontiers in Plant Science*. 2022; 13: 952759.