

Güncel Biyokimya Çalışmaları VII

Editör
Doğan YÜCEL



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1. Tıbbi Biyokimya.

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Akademisyen Yayınevi A.Ş.

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Bölüm 1

NÖRODEJENERATİF HASTALIKLARDA SİRTUİNLER VE MİTOKONDRIYAL AKTİVİTE

Yasemin ATICI¹
Zeynep ÖKTEN²
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1. SİRTUİNLERİN YAPISI, LOKALİZASYONU VE ENZİMATİK AKTİVİTELERİ

Sessiz bilgi düzenleyici 2 (Sir2) proteinleri olarak da bilinen sirtuinler ilk olarak mayada (*Saccharomyces cerevisiae*) tanımlanmıştır. Nikotinamid adenin dinükleotid (NAD) bağımlı protein deasetilazlar ailesinin bir alt tipidir ve sınıf III histon deasetilazlar (HDAC'ler) olarak kategorize edilirler. Sirtuinler (SIRT), histonlardan, transkripsiyon faktörlerinden, enzimlerden, histon olmayan proteinlerden asetil grubunu çıkarır. İnsanlarda ve diğer memelilerde yedi farklı sirtuin vardır. SIRT 1'den SIRT 7'ye kadar tanımlanan sirtuinlerden SIRT1 en iyi tanımlanmış olanıdır (1).

SIRT'ler, C- ve N-terminal alanlarında uzunluk ve dizi bakımından farklılık gösterir. Bu nedenle hücrenin farklı yerlerinde lokalize olurlar. SIRT1 ve SIRT2 nükleusta ve sitoplazmada lokalize olurken, SIRT3-SIRT5 mitokondride lokalize olur (Şekil 1) (2,3).

Sirtuinler, NAD+ bağımlı lizin deasetilasyonu yaparlar. Aynı zamanda yapılarından süksinil, malonil ve uzun zincirli açıl gruplarını da çıkardıkları gösterilmiştir (5,6). Bu enzimler, hücre sağkalımı, proliferasyon, yaşlanma, apoptoz, DNA onarımı ve kalori kısıtlaması gibi çeşitli biyolojik süreçlere dahil olmuşlardır (7,8).

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stresle mücadelede rol oynayabilir, enerji metabolizmasını düzenleyebilir ve DNA onarım mekanizmalarını etkinleştirebilir. Bu nedenle, SIRT1'in potansiyel olarak yaşlanma süreçleri ve çeşitli hastalıklar üzerinde olumlu etkileri olabileceği düşünülmektedir. Sirtuin aktivitesinin artırılması, yaşlanma süreçlerini geciktirme ve yaşa bağlı hastalıkların gelişimini önleme potansiyeline sahip olabilir. Ancak, bu konudaki araştırmalar henüz tam olarak netleşmemiş olup, sirtuinlerin fonksiyonları ve potansiyel uygulamaları üzerindeki çalışmalar devam etmektedir. Sirtuinler arasındaki kesin moleküler mekanizmaları aydınlatmak için gelecekteki çalışmalara ihtiyaç vardır. Ayrıca, yeni sirtuin modülatörlerinin keşfi ve hücreler, hayvanlar ve klinik deneylerin karşılaştırmalı çalışmalarının geliştirilmesi, etkili anti-nörodejenerasyon ilaçlarının keşfi ve geliştirilmesine katkıda bulunabilir.

KAYNAKÇA

1. Finkel T, Deng C-X, Mostoslavsky R. Recent progress in the biology and physiology of sirtuins. *Nature*. 2009; 460: 587–91.
2. Huang J-Y, Hirschev MD, Shimazu T, et al. Mitochondrial sirtuins. *Biochimica et Biophysica Acta (BBA)- Proteins and Proteomics*. 2010; 1804: 1645–51.
3. Vaquero A, Scher MB, Lee DH, et al. SirT2 is a histone deacetylase with preference for histone H4 Lys 16 during mitosis. *Genes Dev*. 2006; 20: 1256–61.
4. Yıldırım S, Demirel R, İcen M, et al. Sirtuin1-3 Deasetilazlar: Biyolojik Fonksiyonları ve Kanserde Terapötik Potansiyelleri. *Iğdır Üniversitesi FBED*, 2022; 12(2): 1055-1069.
5. Du J, Jiang H, Lin H. Investigating the ADP-ribosyltransferase Activity of Sirtuins with NAD Analogues and 32 P-NAD. *Biochemistry*. 2009; 48: 2878–90.
6. Roessler C, Nowak T, Pannek M et al. Chemical Probing of the Human Sirtuin 5 Active Site Reveals Its Substrate Acyl Specificity and Peptide-Based Inhibitors. *Angewandte Chemie International Edition*. 2014; 53: 10728–32.
7. Bordone L, Cohen D, Robinson A, et al. SIRT1 transgenic mice show phenotypes resembling calorie restriction. *Aging Cell*. 2007; 6: 759–67.
8. Rogina B, Helfand SL. Sir2 mediates longevity in the fly through a pathway related to calorie restriction. *PNAS*. 2004; 101: 15998–6003.
9. Frye RA. Characterization of Five Human cDNAs with Homology to the Yeast SIR2 Gene: Sir2-like Proteins (Sirtuins) Metabolize NAD and May Have Protein ADP-Ribosyltransferase Activity. *Biochem Biophys Res Commun*. 1999; 260: 273–9.
10. Frye RA. Phylogenetic Classification of Prokaryotic and Eukaryotic Sir2-like Proteins. *Biochem Biophys Res Commun*. 2000; 273: 793–8.
11. Haigis MC, Guarente LP. Mammalian sirtuins—emerging roles in physiology, aging, and calorie restriction. *Genes Dev*. 2006; 20: 2913–21.
12. Jin Q, Yan T, Ge X, et al. Cytoplasm-localized SIRT1 enhances apoptosis. *J Cell Physiol*. 2007; 213: 88–97.
13. Dryden SC, Nahhas FA, Nowak JE, et al. Role for Human SIRT2 NAD-Dependent Deacetylase Activity in Control of Mitotic Exit in the Cell Cycle. *Mol Cell Biol*. 2003; 23: 3173–85.

14. Serrano L, Martínez-Redondo P, Marazuela-Duque A, et al. The tumor suppressor SirT2 regulates cell cycle progression and genome stability by modulating the mitotic deposition of H4K20 methylation. *Genes Dev.* 2013; 27: 639–53.
15. Chang H-C, Guarente L. SIRT1 and other sirtuins in metabolism. *Trends Endocrinol. Metab.* 2014; 25: 138–45.
16. Mahlknecht U, Voelter-Mahlknecht S. Fluorescence in situ hybridization and chromosomal organization of the sirtuin 4 gene (*Sirt4*) in the mouse. *Biochem Biophys Res Commun.* 2009; 382: 685–90.
17. Anderson KA, Huynh FK, Fisher-Wellman K, et al. SIRT4 Is a Lysine Deacylase that Controls Leucine Metabolism and Insulin Secretion. *Cell Metab.* 2017; 25: 838-855. e15.
18. Mathias RA, Greco TM, Oberstein A, et al. Sirtuin 4 Is a Lipoamidase Regulating Pyruvate Dehydrogenase Complex Activity. *Cell.* 2014; 159: 1615–25.
19. Rardin MJ, He W, Nishida Y, et al. SIRT5 regulates the mitochondrial lysine succinylome and metabolic networks. *Cell Metab.* 2013;18(6):920-933.
20. Kumar S, Lombard DB. Functions of the sirtuin deacylase SIRT5 in normal physiology and pathobiology. *Crit Rev Biochem Mol Biol.* 2018; 53: 311–34.
21. Korotkov A, Seluanov A, Gorbunova V. Sirtuin 6: linking longevity with genome and epigenome stability. *Trends Cell Biol.* 2021; 31: 994–1006.
22. Zhang M, Tang Z. Therapeutic potential of natural molecules against Alzheimer's disease via SIRT1 modulation. *Biomed. Pharmacother.* 2023; 161: 114474.
23. Jiang H, Khan S, Wang Y, et al. SIRT6 regulates TNF- α secretion through hydrolysis of long-chain fatty acyl lysine. *Nature.* 2013; 496: 110–3.
24. Meng H, Yan W-Y, Lei Y-H, et al. SIRT3 Regulation of Mitochondrial Quality Control in Neurodegenerative Diseases. *Front Aging Neurosci.* 2019; 11.
25. Scarpulla RC. Transcriptional activators and coactivators in the nuclear control of mitochondrial function in mammalian cells. *Gene.* 2002; 286: 81–9.
26. Chuang YC, Chen SD, Jou SB, et al. Sirtuin 1 Regulates Mitochondrial Biogenesis and Provides an Endogenous Neuroprotective Mechanism Against Seizure-Induced Neuronal Cell Death in the Hippocampus Following Status Epilepticus. *Int J Mol Sci.* 2019;20(14):3588.
27. Bernier M, Paul RK, Martin-Montalvo A, et al. Negative regulation of STAT3 protein-mediated cellular respiration by SIRT1 protein. *J Biol Chem.* 2011;286(22):19270-19279.
28. Schartner E, Sabbir MG, Saleh A, et al. High glucose concentration suppresses a SIRT2 regulated pathway that enhances neurite outgrowth in cultured adult sensory neurons. *Exp Neurol.* 2018; 309: 134–47.
29. Fourcade S, Morató L, Parameswaran J, et al. Loss of SIRT2 leads to axonal degeneration and locomotor disability associated with redox and energy imbalance. *Aging Cell.* 2017;16(6):1404-1413.
30. Xin T, Lu C. SirT3 activates AMPK-related mitochondrial biogenesis and ameliorates sepsis-induced myocardial injury. *Aging (Albany NY).* 2020;12(16):16224-16237.
31. Cantó C, Gerhart-Hines Z, Feige JN et al. AMPK regulates energy expenditure by modulating NAD⁺ metabolism and SIRT1 activity. *Nature.* 2009; 458: 1056–60.
32. Hasan-Olive MM, Lauritzen KH, Ali M, et al. A Ketogenic Diet Improves Mitochondrial Biogenesis and Bioenergetics via the PGC1 α -SIRT3-UCP2 Axis. *Neurochem Res.* 2019; 44: 22–37.

33. Tseng AHH, Shieh S-S, Wang DL. SIRT3 deacetylates FOXO3 to protect mitochondria against oxidative damage. *Free Radic Biol Med.* 2013; 63: 222–34.
34. Sun Q, Kang R, Chen K, et al. Sirtuin 3 is required for the protective effect of Resveratrol on Manganese-induced disruption of mitochondrial biogenesis in primary cultured neurons. *J Neurochem.* 2021; 156: 121–35.
35. Ho L, Titus AS, Banerjee KK, et al. SIRT4 regulates ATP homeostasis and mediates a retrograde signaling via AMPK. *Aging.* 2013; 5: 835–49.
36. Buler M, Aatsinki S, Izzi V, et al. SIRT5 is under the control of PGC-1 α and AMPK and is involved in regulation of mitochondrial energy metabolism. *The FASEB Journal.* 2014; 28: 3225–37.
37. Ding W-X, Yin X-M. Mitophagy: mechanisms, pathophysiological roles, and analysis. *bchm.* 2012; 393: 547–64.
38. Wang H, Dou S, Zhu J, et al. Ghrelin protects against rotenone-induced cytotoxicity: Involvement of mitophagy and the AMPK/SIRT1/PGC1 α pathway. *Neuropeptides.* 2021; 87: 102134.
39. Huang S, Hong Z, Zhang L, et al. CERKL alleviates ischemia reperfusion-induced nervous system injury through modulating the SIRT1/PINK1/Parkin pathway and mitophagy induction. *Biol Chem.* 2022; 403: 691–701.
40. Zhao N, Xia J, Xu B. Physical exercise may exert its therapeutic influence on Alzheimer's disease through the reversal of mitochondrial dysfunction via SIRT1-FOXO1/3-PINK1-Parkin-mediated mitophagy. *J Sport Health Sci.* 2021; 10: 1–3.
41. Chang C-C, Tsou S-H, Chen W-J, et al. miR-302 Attenuates Mutant Huntingtin-Induced Cytotoxicity through Restoration of Autophagy and Insulin Sensitivity. *Int J Mol Sci.* 2021; 22: 8424.
42. Silva DF, Esteves AR, Oliveira CR, Cardoso SM. Mitochondrial Metabolism Power SIRT2-Dependent Deficient Traffic Causing Alzheimer's-Disease Related Pathology. *Mol Neurobiol.* 2017; 54: 4021–40.
43. Sampaio-Marques B, Felgueiras C, Silva A, et al. Autophagy. 2012; 8: 1494–509.
44. Zhou ZD, Tan EK. Oxidized nicotinamide adenine dinucleotide-dependent mitochondrial deacetylase sirtuin-3 as a potential therapeutic target of Parkinson's disease. *Ageing Res Rev.* 2020; 62: 101107.
45. Lang A, Anand R, Altinluk-Hambüchen S, et al. SIRT4 interacts with OPA1 and regulates mitochondrial quality control and mitophagy. *Aging.* 2017; 9: 2163–89.
46. Polletta L, Vernucci E, Carnevale I, et al. SIRT5 regulation of ammonia-induced autophagy and mitophagy. *Autophagy.* 2015;11(2):253-270.
47. Hong Y-X, Wu W-Y, Song F, et al. Cardiac senescence is alleviated by the natural flavone acacetin via enhancing mitophagy. *Aging.* 2021; 13: 16381–403.
48. Chang AL, Doering TL. Maintenance of Mitochondrial Morphology in *Cryptococcus neoformans* Is Critical for Stress Resistance and Virulence. *mBio.* 2018;9(6):e01375-18.
49. Palmer CS, Elgass KD, Parton RG, et al. Adaptor proteins MiD49 and MiD51 can act independently of Mff and Fis1 in Drp1 recruitment and are specific for mitochondrial fission. *J Biol Chem.* 2013;288(38):27584-27593.
50. Oanh NTK, Park Y-Y, Cho H. Mitochondria elongation is mediated through SIRT1-mediated MFN1 stabilization. *Cell Signal.* 2017; 38: 67–75.

51. Song SB, Park JS, Jang SY, Hwang ES. Nicotinamide Treatment Facilitates Mitochondrial Fission through Drp1 Activation Mediated by SIRT1-Induced Changes in Cellular Levels of cAMP and Ca²⁺. *Cells*. 2021; 10: 612.
52. Cha Y, Kim T, Jeon J, Jang Y, Kim PB, Lopes C, Leblanc P, Cohen BM, Kim K-S. SIRT2 regulates mitochondrial dynamics and reprogramming via MEK1-ERK-DRP1 and AKT1-DRP1 axes. *Cell Rep*. 2021; 37: 110155.
53. Samant SA, Zhang HJ, Hong Z, et al. SIRT3 Deacetylates and Activates OPA1 To Regulate Mitochondrial Dynamics during Stress. *Mol Cell Biol*. 2014; 34: 807–19.
54. Tyagi A, Nguyen CU, Chong T, et al. Reusch JEB, Pugazhenth S. SIRT3 deficiency-induced mitochondrial dysfunction and inflammasome formation in the brain. *Sci Rep*. 2018; 8: 17547.
55. Guedouari H, Daigle T, Scorrano L et al.. Sirtuin 5 protects mitochondria from fragmentation and degradation during starvation. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*. 2017; 1864: 169–76.
56. Kao T-Y, Chiu Y-C, Fang W-C, et al. Mitochondrial Lon regulates apoptosis through the association with Hsp60–mtHsp70 complex. *Cell Death Dis*. 2015; 6: e1642–e1642.
57. Mouchiroud L, Houtkooper RH, Moullan N, et al. The NAD⁺/Sirtuin Pathway Modulates Longevity through Activation of Mitochondrial UPR and FOXO Signaling. *Cell*. 2013; 154: 430–41.
58. Wang B, Ge S, Xiong W, et al. Effects of resveratrol pretreatment on endoplasmic reticulum stress and cognitive function after surgery in aged mice. *BMC Anesthesiol*. 2018; 18: 141.
59. Hou M, Bao W, Gao Y, et al. Honokiol improves cognitive impairment in APP/PS1 mice through activating mitophagy and mitochondrial unfolded protein response. *Chem Biol Interact*. 2022; 351: 109741.
60. Shi H, Deng H-X, Gius D, et al. Sirt3 protects dopaminergic neurons from mitochondrial oxidative stress. *Hum Mol Genet*. 2017; 26: 1915–26.
61. Lu Z, Chen Y, Aponte AM, et al. Prolonged fasting identifies heat shock protein 10 as a Sirtuin 3 substrate: elucidating a new mechanism linking mitochondrial protein acetylation to fatty acid oxidation enzyme folding and function. *J Biol Chem*. 2015;290(4):2466-2476.
62. Gibellini L, Pinti M, Beretti F, et al. Sirtuin 3 interacts with Lon protease and regulates its acetylation status. *Mitochondrion*. 2014; 18: 76–81.
63. Xu H, Liu Y-Y, Li L-S, et al. Sirtuins at the Crossroads between Mitochondrial Quality Control and Neurodegenerative Diseases: Structure, Regulation, Modifications, and Modulators. *Aging Dis*. 2023; 14: 794.
64. Silva MVF, Loures CMG, Alves LCV, et al. Alzheimer's disease: risk factors and potentially protective measures. *J Biomed Sci*. 2019;26(1):33.
65. Lin C-L, Cheng Y-S, Li H-H, et al. Amyloid- β suppresses AMP-activated protein kinase (AMPK) signaling and contributes to α -synuclein-induced cytotoxicity. *Exp Neurol*. 2016; 275: 84–98.
66. Pradhan R, Singh AK, Kumar P, et al. Blood Circulatory Level of Seven Sirtuins in Alzheimer's Disease: Potent Biomarker Based on Translational Research. *Mol Neurobiol*. 2022;59(3):1440-1451.
67. Lutz MI, Milenkovic I, Regelsberger G, et al. Distinct Patterns of Sirtuin Expression During Progression of Alzheimer's Disease. *Neuromolecular Med*. 2014; 16: 405–14.

68. Cheng A, Wang J, Ghena N, et al. SIRT3 Haploinsufficiency Aggravates Loss of GABAergic Interneurons and Neuronal Network Hyperexcitability in an Alzheimer's Disease Model. *J. Neurosci. Res.* 2020; 40: 694–709.
69. Verdin E, Hirschey MD, Finley LWS, Haigis MC. Sirtuin regulation of mitochondria: energy production, apoptosis, and signaling. *Trends Biochem Sci.* 2010; 35: 669–75.
70. Song S, Li B, Jia Z, et al. Sirtuin 3 mRNA Expression is Downregulated in the Brain Tissues of Alzheimer's Disease Patients: A Bioinformatic and Data Mining Approach. *Med Sci Monit.* 2020;26:e923547.
71. Tyagi A, Mirita C, Taher N, et al. Metabolic syndrome exacerbates amyloid pathology in a comorbid Alzheimer's mouse model. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866(10):165849.
72. Chen J, Zhou Y, Mueller-Steiner S, et al. SIRT1 protects against microglia-dependent amyloid-beta toxicity through inhibiting NF-kappaB signaling. *J Biol Chem.* 2005;280(48):40364-40374.
73. Xiao H, Xie Y, Xi K, et al. Targeting Mitochondrial Sirtuins in Age-Related Neurodegenerative Diseases and Fibrosis. *Aging Dis.* 2023; 14: 1583.
74. Wu S, Wei Y, Li J, et al. SIRT5 Represses Neurotrophic Pathways and A β Production in Alzheimer's Disease by Targeting Autophagy. *ACS Chem Neurosci.* 2021; 12: 4428–37.
75. Kaluski S, Portillo M, Besnard A, et al. Neuroprotective Functions for the Histone Deacetylase SIRT6. *Cell Rep.* 2017;18(13):3052-3062.
76. Maxwell MM, Tomkinson EM, Nobles J, et al. The Sirtuin 2 microtubule deacetylase is an abundant neuronal protein that accumulates in the aging CNS. *Hum Mol Genet.* 2011; 20: 3986–96.
77. Pascoal TA, Benedet AL, Ashton NJ, et al. Publisher Correction: Microglial activation and tau propagate jointly across Braak stages. *Nat Med.* 2021;27(11):2048-2049.
78. Gaetani L, Bellomo G, Parnetti L, et al. Neuroinflammation and Alzheimer's Disease: A Machine Learning Approach to CSF Proteomics. *Cells.* 2021; 10: 1930.
79. Koo J-H, Kang E-B, Oh Y-S, et al. Treadmill exercise decreases amyloid- β burden possibly via activation of SIRT-1 signaling in a mouse model of Alzheimer's disease. *Exp Neurol.* 2017; 288: 142–52.
80. Julien C, Tremblay C, Émond V, et al. Sirtuin 1 Reduction Parallels the Accumulation of Tau in Alzheimer Disease. *J Neuropathol Exp Neurol.* 2009; 68: 48–58.
81. Kumar R, Nigam L, Singh AP, et al. Design, synthesis of allosteric peptide activator for human SIRT1 and its biological evaluation in cellular model of Alzheimer's disease. *Eur J Med Chem.* 2017; 127:909-916.
82. Gomes BAQ, Silva JPB, Romeiro CFR, et al. Neuroprotective Mechanisms of Resveratrol in Alzheimer's Disease: Role of SIRT1. *Oxid Med Cell Longev.* 2018; 2018: 1–15.
83. Yu S, Zhou X, Xiang H, et al. Resveratrol Reduced Liver Damage After Liver Resection in a Rat Model by Upregulating Sirtuin 1 (SIRT1) and Inhibiting the Acetylation of High Mobility Group Box 1 (HMGB1). *Med Sci Monit.* 2019; 25:3212-3220.
84. Kumar V, Pandey A, Jahan S, et al. Differential responses of Trans-Resveratrol on proliferation of neural progenitor cells and aged rat hippocampal neurogenesis. *Sci Rep.* 2016; 6:28142.
85. Ma CY, Yao MJ, Zhai QW, et al. SIRT1 suppresses self-renewal of adult hippocampal neural stem cells. *Development.* 2014 Dec;141(24):4697-709.

86. Saharan S, Jhaveri DJ, Bartlett PF. SIRT1 regulates the neurogenic potential of neural precursors in the adult subventricular zone and hippocampus. *J Neurosci Res.* 2013; 91: 642–59.
87. Abozaid OAR, Sallam MW, Ahmed ESA. Mesenchymal Stem Cells Modulate SIRT1/ MiR-134/ GSK3 β Signaling Pathway in a Rat Model of Alzheimer's Disease. *J Prev Alzheimers Dis (Internet).* *J Prev Alzheimers Dis;* 2022; 9: 458–68.
88. Ascherio A, Schwarzschild MA. The epidemiology of Parkinson's disease: risk factors and prevention. *Lancet Neurol.* 2016; 15: 1257–72.
89. Rocha EM, De Miranda B, Sanders LH. Alpha-synuclein: Pathology, mitochondrial dysfunction and neuroinflammation in Parkinson's disease. *Neurobiol Dis.* 2018; 109: 249–57.
90. Maszlag-Török R, Boros FA, Vécsei L, et al. Gene variants and expression changes of SIRT1 and SIRT6 in peripheral blood are associated with Parkinson's disease. *Sci Rep.* 2021; 11: 10677.
91. Nicholatos JW, Francisco AB, Bender CA, et al. Nicotine promotes neuron survival and partially protects from Parkinson's disease by suppressing SIRT6. *Acta Neuropathol Commun.* 2018; 6: 120.
92. Li X, Feng Y, Wang X-X, et al. The Critical Role of SIRT1 in Parkinson's Disease: Mechanism and Therapeutic Considerations. *Aging Dis.* 2020; 11: 1608.
93. Park JH, Burgess JD, Faruqi AH, et al. Alpha-synuclein-induced mitochondrial dysfunction is mediated via a sirtuin 3-dependent pathway. *Mol Neurodegener.* 2020; 15: 5.
94. Chopra V, Quinti L, Kim J, et al. The Sirtuin 2 Inhibitor AK-7 Is Neuroprotective in Huntington's Disease Mouse Models. *Cell Rep.* 2012; 2: 1492–7.
95. Singh P, Hanson PS, Morris CM. Sirtuin-2 Protects Neural Cells from Oxidative Stress and Is Elevated in Neurodegeneration. *Parkinsons Dis.* 2017; 2017: 1–17.
96. Hou Y, Dan X, Babbar M, et al. Ageing as a risk factor for neurodegenerative disease. *Nat Rev Neurol.* 2019; 15: 565–81.
97. Shi H, Deng H-X, Gius D, et al. Sirt3 protects dopaminergic neurons from mitochondrial oxidative stress. *Hum Mol Genet.* 2017; 26: 1915–26.
98. Liu L, Peritore C, Ginsberg J, et al. Protective role of SIRT5 against motor deficit and dopaminergic degeneration in MPTP-induced mice model of Parkinson's disease. *Behav. Brain Res.* 2015; 281: 215–21.
99. Browne SE, Bowling AC, Macgarvey U, et al. Oxidative damage and metabolic dysfunction in Huntington's disease: Selective vulnerability of the basal ganglia. *Ann Neurol.* 1997; 41: 646–53.
100. Panov AV, Gutekunst C-A, Leavitt BR, et al. Early mitochondrial calcium defects in Huntington's disease are a direct effect of polyglutamines. *Nat Neurosci.* 2002; 5: 731–6.
101. Kim J, Moody JP, Edgerly CK, et al. Mitochondrial loss, dysfunction and altered dynamics in Huntington's disease. *Hum Mol Genet.* 2010; 19: 3919–35.
102. Baldo B, Gabery S, Soyly-Kucharz R, et al. SIRT1 is increased in affected brain regions and hypothalamic metabolic pathways are altered in Huntington disease. *Neuropathol Appl Neurobiol.* 2019; 45: 361–79.
103. Salamon A, Maszlag-Török R, Veres G, et al. Cerebellar Predominant Increase in mRNA Expression Levels of Sirt1 and Sirt3 Isoforms in a Transgenic Mouse Model of Huntington's Disease. *Neurochem Res.* 2020; 45: 2072–81.

104. Marin B, Fontana A, Arcuti S, et al. Age-specific ALS incidence: a dose-response meta-analysis. *Eur J Epidemiol.* 2018; 33: 621–34.
105. Bosco DA, Morfini G, Karabacak NM, et al. Wild-type and mutant SOD1 share an aberrant conformation and a common pathogenic pathway in ALS. *Nat Neurosci.* 2010; 13: 1396–403.
106. Song W, Song Y, Kincaid B, et al. Mutant SOD1G93A triggers mitochondrial fragmentation in spinal cord motor neurons: Neuroprotection by SIRT3 and PGC-1 α . *Neurobiol Dis.* 2013; 51: 72–81.

Bölüm 2

LİPİT METABOLİZMASININ VE APOLİPOPROTEİNLERİN ALZHEIMER HASTALIĞINA ETKİSİ

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1. GİRİŞ

Alzheimer Hastalığı (AH), normal yaşlanma süreci dışında düşünme ve hafıza becerilerini zamanla azaltan, birçok işlevin tek başına gerçekleştirilmesini geri dönüşümsüz bir şekilde engelleyen; davranışsal ve fonksiyonel bozukluklarla devam eden özellikle hipokampus üzerinde etkili olan bir beyin hastalığıdır. AH bilişsel işlevlerin kaybıyla kendini gösteren demansın en yaygın tipi olan nörodejeneratif bir hastalıktır. AH'nin belirtileri arasında beyinde hafıza ve işlevsel bozukluklar meydana gelmektedir. Hastalığın kendine özgü görülen önemli patolojik bulguları vardır. Bunlar; amiloid beta (A β), Tau proteinlerinin hiperfosforile olması ve hücre içerisinde çözünemeyen nörofibril yumak (NFY)'lerin birikimi ile birlikte sinaptik disfonksiyondan kaynaklanan bozukluklardır (1).

Lipitler beynin kuru ağırlığının büyük kısmını oluşturmaktadır. Beyin, lipitleri glukozdan gerekli enerjiyi sağlayamadığı durumlarda kullanır. Genetik faktörler ve yaşam tarzı lipit metabolizmasını önemli derecede etkiler ve lipit metabolizmasında görülen tüm değişiklikler AH ile ilişkilendirilmektedir (1). Lipit metabolizmasında ve lipitlerin hücre içerisinde taşınmasında önemli rol oynayan apolipoprotein E4 (APOE4) hastalığın bilinen en yaygın genetik faktörüdür. Ek olarak lipit metabolizması AH'de etkili olan kan-beyin bariyerinin işlevi, amiloid öncül proteininin (APP) işlenmesi, miyelinasyon, membran yeniden modellenmesi,

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Kolesterolün serebral ve hipokampal kortekste artması, hafıza ve öğrenmeyi geri dönüşümü olmayacak şekilde etkilemektedir. Orta yaşlarda yüksek kolesterollü diyet uygulanması AH için risk faktörü oluştururken, ileri yaşlardaki yüksek kolesterolün AH ile bağlantısı bulunamamıştır. Kolesterolün AH için koruyucu bir etkiye sahip olup olmadığı henüz tam anlamıyla belirlenememiştir. Bunun yanında, AH ile kolesterol arasındaki ilişkinin yaşa ve doza bağımlı olarak farklılık gösterdiği belirtilmektedir.

Normal yaşlanmada lipit homeostazı beyin temel işlevlerinin karşılanmasını sağlar. AH'de lipit metabolizmasında homeostazın bozulması söz konusudur ve bu durum AH'nin ilerlemesini karakterize eden anormal beyin fonksiyonlarıyla sonuçlanır. AH'nin daha kapsamlı mekanizmalarını, spesifik biyobelirteçlerini ve yeni tedavilerini ortaya çıkarmak için beyin lipit homeostazının araştırılacağı daha kapsamlı çalışmalara ihtiyaç vardır.

KAYNAKÇA

1. Husain M, Schott JM. (Ed). Oxford textbook of cognitive neurology and dementia. *Oxford University Press*. 2016.
2. Chew H, Solomon VA, Fonteh AN. Involvement of Lipids in Alzheimer's Disease Pathology and Potential Therapies. *Front Physiol*. 2020;9(11):598. doi: 10.3389/fphys.2020.00598
3. Ann Dipika Binoshia Fernando, WM, Rainey-Smith SR, Martins IJ, et al. In Vitro Study to Assess the Potential of Short Chain Fatty Acids (SCFA) as Therapeutic Agents for Alzheimer's Disease. *Alzheimer's & Dementia*, 2014; 10, 626.
4. Dinkova-Kostova AT, Kostov RV. Glucosinolates and isothiocyanates in health and disease. *Trends Mol Med*. 2012;18(6):337-47. doi: 10.1016/j.molmed.2012.04.003
5. Lei E, Vacy K, Boon WC. Fatty acids and their therapeutic potential in neurological disorders. *Neurochem Int*. 2016;95:75-84. doi: 10.1016/j.neuint.2016.02.014
6. Bianca Velasco A, Tan ZS. Fatty Acids and the Aging Brain. In: Watson RR, De Meester F, (ed). Omega-3 Fatty Acids in Brain and Neurological Health., *Elsevier*; 2014 p. 201-19.
7. Hooijmans CR, Kiliaan AJ. Fatty acids, lipid metabolism and Alzheimer pathology. *Eur J Pharmacol*. 2008;6,585(1):176-96. doi: 10.1016/j.ejphar.2007.11.081
8. Sultana R, Perluigi M, Butterfield DA. Protein oxidation and lipid peroxidation in brain of subjects with Alzheimer's disease: insights into mechanism of neurodegeneration from redox proteomics. *Antioxid Redox Signal*. 2006;8(11-12):2021-37. doi: 10.1089/ars.2006.8.2021
9. Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med*. 1991;11(1):81-128. doi: 10.1016/0891-5849(91)90192-6
10. Markesbery WR. Oxidative stress hypothesis in Alzheimer's disease. *Free Radic Biol Med*. 1997;23(1):134-47. doi: 10.1016/s0891-5849(96)00629-6
11. Morris MC, Tangney CC. Dietary fat composition and dementia risk. *Neurobiol Aging*. 2014;35,2:S59-64. doi: 10.1016/j.neurobiolaging.2014.03.038

12. Naqvi AZ, Harty B, Mukamal KJ, et al. Monounsaturated, trans, and saturated Fatty acids and cognitive decline in women. *J Am Geriatr Soc.* 2011;59(5):837-43. doi: 10.1111/j.1532-5415.2011.03402.x
13. Takechi R, Galloway S, Pallegage-Gamarallage MM, et al. Dietary fats, cerebrovasculature integrity and Alzheimer's disease risk. *Prog Lipid Res.* 2010;49(2):159-70. doi: 10.1016/j.plipres.2009.10.004
14. Schaefer EJ, Bongard V, Beiser AS, et al. Plasma phosphatidylcholine docosahexaenoic acid content and risk of dementia and Alzheimer disease: the Framingham Heart Study. *Arch Neurol.* 2006;63(11):1545-50. doi: 10.1001/archneur.63.11.1545
15. Florent-Bécharde S, Desbène C, Garcia P, et al. The essential role of lipids in Alzheimer's disease. *Biochimie.* 2009;91(6):804-9. doi: 10.1016/j.biochi.2009.03.004
16. Hooijmans CR, Van der Zee CE, Dederen PJ, et al. DHA and cholesterol containing diets influence Alzheimer-like pathology, cognition and cerebral vasculature in AP-Pswe/PS1dE9 mice. *Neurobiol Dis.* 2009;33(3):482-98. doi: 10.1016/j.nbd.2008.12.002
17. Lim GP, Calon F, Morihara T, et al. A diet enriched with the omega-3 fatty acid docosahexaenoic acid reduces amyloid burden in an aged Alzheimer mouse model. *J Neurosci.* 2005;23;25(12):3032-40. doi: 10.1523/JNEUROSCI.4225-04.2005
18. Oster T, Pillot T. Docosahexaenoic acid and synaptic protection in Alzheimer's disease mice. *Biochim Biophys Acta.* 2010;1801(8):791-8. doi: 10.1016/j.bbailip.2010.02.011
19. Wu A, Ying Z, Gomez-Pinilla F. The interplay between oxidative stress and brain-derived neurotrophic factor modulates the outcome of a saturated fat diet on synaptic plasticity and cognition. *Eur J Neurosci.* 2004;19(7):1699-707. doi: 10.1111/j.1460-9568.2004.03246.x
20. Morris MC, Evans DA, Tangney CC, et al. Dietary copper and high saturated and trans fat intakes associated with cognitive decline. *Arch Neurol.* 2006;63(8):1085-8. doi: 10.1001/archneur.63.8.1085
21. Pistell PJ, Morrison CD, Gupta S, et al. Cognitive impairment following high fat diet consumption is associated with brain inflammation. *J Neuroimmunol.* 2010;26;219(1-2):25-32. doi: 10.1016/j.jneuroim.2009.11.010
22. Park HR, Park M, Choi J, et al. A high-fat diet impairs neurogenesis: involvement of lipid peroxidation and brain-derived neurotrophic factor. *Neurosci Lett.* 2010;4;482(3):235-9. doi: 10.1016/j.neulet.2010.07.046
23. Zhang X, Dong F, Ren J, et al. High dietary fat induces NADPH oxidase-associated oxidative stress and inflammation in rat cerebral cortex. *Exp Neurol.* 2005;191(2):318-25. doi: 10.1016/j.expneurol.2004.10.011
24. Solfrizzi V, D'Introno A, Colacicco AM, et al. Dietary fatty acids intake: possible role in cognitive decline and dementia. *Exp Gerontol.* 2005;40(4):257-70. doi: 10.1016/j.exger.2005.01.001
25. Simopoulos AP. Evolutionary aspects of diet: the omega-6/omega-3 ratio and the brain. *Mol Neurobiol.* 2011;44(2):203-15. doi: 10.1007/s12035-010-8162-0
26. Fernando WM, Martins IJ, Goozee KG, et al. The role of dietary coconut for the prevention and treatment of Alzheimer's disease: potential mechanisms of action. *Br J Nutr.* 2015;14;114(1):1-14. doi: 10.1017/S0007114515001452
27. Hashimoto M, Hossain S, Shimada T, et al. Docosahexaenoic acid-induced protective effect against impaired learning in amyloid beta-infused rats is associated with increased synaptosomal membrane fluidity. *Clin Exp Pharmacol Physiol.* 2006;33(10):934-9. doi: 10.1111/j.1440-1681.2006.04467.x

28. Yurko-Mauro K, Alexander DD, Van Elswyk ME. Docosahexaenoic acid and adult memory: a systematic review and meta-analysis. *PLoS One*. 2015;18;10(3):e0120391. doi: 10.1371/journal.pone.0120391
29. Agrawal R, Gomez-Pinilla F. Metabolic syndrome' in the brain: deficiency in omega-3 fatty acid exacerbates dysfunctions in insulin receptor signalling and cognition. *J Physiol*. 2012;15;590(10):2485-99. doi: 10.1113/jphysiol.2012.230078
30. Seneff S, Wainwright G, Mascitelli L. Nutrition and Alzheimer's disease: The detrimental role of a high carbohydrate diet. *Eur J Int Med* 2011;7(1):8-20
31. Cecchi C, Nichino D, Zampagni M, et al. A protective role for lipid raft cholesterol against amyloid-induced membrane damage in human neuroblastoma cells. *Biochim Biophys Acta*. 2009;1788(10):2204-16. doi: 10.1016/j.bbame.2009.07.019
32. Crichton GE, Elias MF, Davey A, et al. Higher HDL cholesterol is associated with better cognitive function: the Maine-Syracuse study. *J Int Neuropsychol Soc*. 2014;20(10):961-70. doi: 10.1017/S1355617714000885
33. Sparks DL, Scheff SW, Hunsaker JC 3rd, et al. Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. *Exp Neurol*. 1994;126(1):88-94. doi: 10.1006/exnr.1994.1044
34. Wang D, Zheng W. Dietary cholesterol concentration affects synaptic plasticity and dendrite spine morphology of rabbit hippocampal neurons. *Brain Res*. 2015;5;1622:350-60. doi: 10.1016/j.brainres.2015.06.049
35. Solomon A, Kivipelto M, Wolozin B, et al. Midlife serum cholesterol and increased risk of Alzheimer's and vascular dementia three decades later. *Dement Geriatr Cogn Disord*. 2009;28(1):75-80. doi: 10.1159/000231980
36. Alonso A, Jacobs DR Jr, Menotti A, et al. Cardiovascular risk factors and dementia mortality: 40 years of follow-up in the Seven Countries Study. *J Neurol Sci*. 2009;15;280(1-2):79-83. doi: 10.1016/j.jns.2009.02.004
37. Hughes TF, Ganguli M. Modifiable Midlife Risk Factors for Late-Life Cognitive Impairment and Dementia. *Curr Psychiatry Rev*. 2009;1;5(2):73-92. doi: 10.2174/157340009788167347
38. Lim WL, Lam SM, Shui G, et al. Effects of a high-fat, high-cholesterol diet on brain lipid profiles in apolipoprotein E ϵ 3 and ϵ 4 knock-in mice. *Neurobiol Aging*. 2013;34(9):2217-24. doi: 10.1016/j.neurobiolaging.2013.03.012
39. Pensalfini A, Zampagni M, Liguri G, et al. Membrane cholesterol enrichment prevents A β -induced oxidative stress in Alzheimer's fibroblasts. *Neurobiol Aging*. 2011;32(2):210-22. doi: 10.1016/j.neurobiolaging.2009.02.010
40. Martins IJ, Hone E, Foster JK, et al. Apolipoprotein E, cholesterol metabolism, diabetes, and the convergence of risk factors for Alzheimer's disease and cardiovascular disease. *Mol Psychiatry*. 2006;11(8):721-36. doi: 10.1038/sj.mp.4001854
41. Agarwal M, Khan S. Plasma Lipids as Biomarkers for Alzheimer's Disease: A Systematic Review. *Cureus*. 2020;10;12(12):e12008. doi: 10.7759/cureus.12008
42. Jeong W, Lee H, Cho S, et al. ApoE4-Induced Cholesterol Dysregulation and Its Brain Cell Type-Specific Implications in the Pathogenesis of Alzheimer's Disease. *Mol Cells*. 2019 30;42(11):739-746. doi: 10.14348/molcells.2019.0200
43. Martín MG, Pfrieger F, Dotti CG. Cholesterol in brain disease: sometimes determinant and frequently implicated. *EMBO Rep*. 2014;15(10):1036-52. doi: 10.15252/embr.201439225

44. Wong MW, Braidy N, Poljak A, et al. Dysregulation of lipids in Alzheimer's disease and their role as potential biomarkers. *Alzheimers Dement.* 2017;3(7):810-827. doi: 10.1016/j.jalz.2017.01.008
45. Braun V, Hantke K. Lipoproteins: Structure, Function, Biosynthesis. *Subcell Biochem.* 2019;92:39-77. doi: 10.1007/978-3-030-18768-2_3
46. Ito J, Nagayasu Y, Miura Y, et al. Astrocyte s endogenous apoE generates HDL-like lipoproteins using previously synthesized cholesterol through interaction with ABCA1. *Brain Res.* 2014;27;1570:1-12. doi: 10.1016/j.brainres.2014.04.037
47. Filou S, Lhomme M, Karavia EA, et al. Distinct Roles of Apolipoproteins A1 and E in the Modulation of High-Density Lipoprotein Composition and Function. *Biochemistry.* 2016;12;55(27):3752-62. doi: 10.1021/acs.biochem.6b00389
48. Wong MWK, Braidy N, Crawford J, et al. APOE Genotype Differentially Modulates Plasma Lipids in Healthy Older Individuals, with Relevance to Brain Health. *J Alzheimers Dis.* 2019;72(3):703-716. doi: 10.3233/JAD-190524
49. Zhao N, Liu CC, Qiao W, et al. Apolipoprotein E, Receptors, and Modulation of Alzheimer's Disease. *Biol Psychiatry.* 2018;15;83(4):347-357. doi: 10.1016/j.biopsych.2017.03.003
50. Merino-Zamorano C, Fernández-de Retana S, Montañola A, et al. Modulation of Amyloid- β 1-40 Transport by ApoA1 and ApoJ Across an in vitro Model of the Blood-Brain Barrier. *J Alzheimers Dis.* 2016;25;53(2):677-91. doi: 10.3233/JAD-150976
51. Nelson AR, Sagare AP, Zlokovic BV. Role of clusterin in the brain vascular clearance of amyloid- β . *Proc Natl Acad Sci U S A.* 2017;15;114(33):8681-8682. doi: 10.1073/pnas.1711357114
52. Zandl-Lang M, Fanaee-Danesh E, Sun Y, et al. Regulatory effects of simvastatin and apoJ on APP processing and amyloid- β clearance in blood-brain barrier endothelial cells. *Biochim Biophys Acta Mol Cell Biol Lipids.* 2018;1863(1):40-60. doi: 10.1016/j.bbalip.2017.09.008
53. Foster EM, Dangla-Valls A, Lovestone S, et al. Clusterin in Alzheimer's Disease: Mechanisms, Genetics, and Lessons From Other Pathologies. *Front Neurosci.* 2019;28;13:164. doi: 10.3389/fnins.2019.00164
54. Panza F, D'Introno A, Colacicco AM, et al. Lipid metabolism in cognitive decline and dementia. *Brain Res Rev.* 2006;51(2):275-92. doi: 10.1016/j.brainresrev.2005.11.007
55. Formiga F, Ferrer A, Chivite D, et al. Serum high-density lipoprotein cholesterol levels correlate well with functional but not with cognitive status in 85-year-old subjects. *J Nutr Health Aging.* 2012;16(5):449-53. doi: 10.1007/s12603-012-0018-z
56. Agarwal R, Tripathi CB. Association of apolipoprotein E genetic variation in Alzheimer's disease in Indian population: a meta-analysis. *Am J Alzheimers Dis Other Demen.* 2014;29(7):575-82. doi: 10.1177/1533317514531443
57. Raygani AV, Rahimi Z, Kharazi H, et al. Association between apolipoprotein E polymorphism and serum lipid and apolipoprotein levels with Alzheimer's disease. *Neurosci Lett.* 2006;6;408(1):68-72. doi: 10.1016/j.neulet.2006.08.048
58. Wingo TS, Cutler DJ, Wingo AP, et al. Association of Early-Onset Alzheimer Disease With Elevated Low-Density Lipoprotein Cholesterol Levels and Rare Genetic Coding Variants of APOB. *JAMA Neurol.* 2019;1;76(7):809-817. doi: 10.1001/jama-neurol.2019.0648

59. Caramelli P, Nitrini R, Maranhão R, et al. Increased apolipoprotein B serum concentration in Alzheimer's disease. *Acta Neurol Scand.* 1999 Jul;100(1):61-3. doi: 10.1111/j.1600-0404.1999.tb00724.x
60. Nunan J, Small DH. Regulation of APP cleavage by alpha-, beta- and gamma-secretases. *FEBS Lett.* 2000;13;483(1):6-10. doi: 10.1016/s0014-5793(00)02076-7
61. Liu K, Liu Y, Xu Y et al. Regulatory role of Golgi brefeldin A resistance factor-1 in amyloid precursor protein trafficking, cleavage and A β formation. *J Cell Biochem.* 2019;120(9):15604-15615. doi: 10.1002/jcb.28827
62. Volmar CH, Salah-Uddin H, Janczura KJ, et al. M344 promotes nonamyloidogenic amyloid precursor protein processing while normalizing Alzheimer's disease genes and improving memory. *Proc Natl Acad Sci U S A.* 2017;24;114(43):E9135-E9144. doi: 10.1073/pnas.1707544114
63. Andrew RJ, Kellett KA, Thinakaran G, et al. A Greek Tragedy: The Growing Complexity of Alzheimer Amyloid Precursor Protein Proteolysis. *J Biol Chem.* 2016;9;291(37):19235-44. doi: 10.1074/jbc.R116.746032
64. Grimm MO, Hauptenthal VJ, Mett J, et al. Oxidized Docosahexaenoic Acid Species and Lipid Peroxidation Products Increase Amyloidogenic Amyloid Precursor Protein Processing. *Neurodegener Dis.* 2016;16(1-2):44-54. doi: 10.1159/000440839
65. Audagnotto M, Kengo Lorkowski A, Dal Peraro M. Recruitment of the amyloid precursor protein by γ -secretase at the synaptic plasma membrane. *Biochem Biophys Res Commun.* 2018;29;498(2):334-341. doi: 10.1016/j.bbrc.2017.10.164
66. Bhattacharyya R, Barren C, Kovacs DM. Palmitoylation of amyloid precursor protein regulates amyloidogenic processing in lipid rafts. *J Neurosci.* 2013;3;33(27):11169-83. doi: 10.1523/JNEUROSCI.4704-12.2013
67. Mukadam AS, Breusegem SY, Seaman MNJ. Analysis of novel endosome-to-Golgi retrieval genes reveals a role for PLD3 in regulating endosomal protein sorting and amyloid precursor protein processing. *Cell Mol Life Sci.* 2018;75(14):2613-2625. doi: 10.1007/s00018-018-2752-9
68. Bartzokis G. Alzheimer's disease as homeostatic responses to age-related myelin breakdown. *Neurobiol Aging.* 2011;32(8):1341-71. doi: 10.1016/j.neurobiolaging.2009.08.007
69. Simons M, Nave KA. Oligodendrocytes: Myelination and Axonal Support. *Cold Spring Harb Perspect Biol.* 2015;22;8(1):a020479. doi: 10.1101/cshperspect.a020479
70. Desai MK, Mastrangelo MA, Ryan DA, et al. Early oligodendrocyte/myelin pathology in Alzheimer's disease mice constitutes a novel therapeutic target. *Am J Pathol.* 2010;177(3):1422-35. doi: 10.2353/ajpath.2010.100087
71. Kohama SG, Rosene DL, Sherman LS. Age-related changes in human and non-human primate white matter: from myelination disturbances to cognitive decline. *Age (Dordr).* 2012;34(5):1093-110. doi: 10.1007/s11357-011-9357-7
72. Wang WY, Tan MS, Yu JT, et al. Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease. *Ann Transl Med.* 2015;3(10):136. doi: 10.3978/j.issn.2305-5839.2015.03.49
73. Zhang C, Wang K, Yang L, et al. Lipid metabolism in inflammation-related diseases. *Analyst.* 2018;24;143(19):4526-4536. doi: 10.1039/c8an01046c
74. Ntambi JM. Highlighting inflammation and lipid metabolism. *Biochem Biophys Res Commun.* 2019;17;520(4):688-689. doi: 10.1016/j.bbrc.2019.10.014

75. Umamaheswaran S, Dasari SK, Yang P, et al. Stress, inflammation, and eicosanoids: an emerging perspective. *Cancer Metastasis Rev.* 2018;37(2-3):203-211. doi: 10.1007/s10555-018-9741-1
76. Chiurchiù V, Leuti A, Maccarrone M. Bioactive Lipids and Chronic Inflammation: *Managing the Fire Within.* *Front Immunol.* 2018;29:9:38. doi: 10.3389/fimmu.2018.00038. PMID: 29434586
77. Whittington RA, Planel E, Terrando N. Impaired Resolution of Inflammation in Alzheimer's Disease: A Review. *Front Immunol.* 2017;6:8:1464. doi: 10.3389/fimmu.2017.01464. PMID: 29163531
78. Cheignon C, Tomas M, Bonnefont-Rousselot D, et al. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* 2018;14:450-464. doi: 10.1016/j.redox.2017.10.014. Epub 2017 Oct 18. PMID: 29080524
79. Shinohara M, Tachibana M, Kanekiyo T, et al. Role of LRP1 in the pathogenesis of Alzheimer's disease: evidence from clinical and preclinical studies. *J Lipid Res.* 2017;58(7):1267-1281. doi: 10.1194/jlr.R075796

Bölüm 3

ALZHEIMER HASTALIĞININ PATOFİZYOLOJİSİ VE OKSİDATİF STRES

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Berat AKTOLUN³

1. ALZHEIMER HASTALIĞI VE OKSİDATİF STRES

Alzheimer hastalığı (AH), yaşlı nüfusta demansın yaygın prevalansından sorumlu olan ilerleyici nörodejeneratif bir bozukluktur (1). Dünya Sağlık Örgütü'nün verileri, dünya çapında demanstan etkilenen yaklaşık 55 milyon birey olduğunu göstermektedir. Her yıl neredeyse 10 milyon yeni AH vakası, demans teşhisi almaktadır. AH'nin küresel yükünün 2050 yılına kadar iki katına çıkması tahmin edilmektedir (2). Bu nedenle AH'nin klinik belirtilerini hafifletmek için potansiyel terapötik müdahalelere ihtiyaç vardır.

Amiloid- β protein öncülünün (A β PP) işleme ve imha mekanizmasındaki kusurlar, çözünen amiloid- β 40 (A β 40), çözünmeyen A β 42 ve beyindeki C-terminal parçaların birikimi, AH'nin patolojisi ile ilişkilendirilmiştir. Ayrıca amiloid plakların aracılık ettiği biyokimyasal, metabolik, moleküler ve hücrel olaylar nörodejenerasyona ve hipokampüsteki nörojenik süreçlerde zararlı etkilere neden olabilmektedir (3). Demansın potansiyel nedeni olarak tanımlanan nöral rejeneratif plastisitenin, nöral kök hücrelerin (NKH) hücre döngüsündeki kusurlar ve hipokampüste yeni gelişen nöronlarda apoptoz nedeniyle bozulduğu belirlenmiştir [4].

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faktördür. Oksidatif stres, antioksidan savunmanın yetersiz olduğu durumlarda reaktif türlerin birikmesine neden olur. Bu durum, lipid peroksidasyonu, protein ve nükleik asitlerin oksidatif modifikasyonu gibi biyokimyasal değişikliklere yol açabilir. Oksidatif stres, A β birikimi, mikroglia aktivasyonu, redoks aktif metal iyonlarının düzensizliği ve mitokondriyal fonksiyon bozukluğu gibi AH ile ilişkilendirilen belirtilerle bağlantılıdır. Mikroglia ve astrosit gibi hücreler, AH'nin histopatolojik belirtileri olan nöroenflamasyona katkıda bulunabilir. Aktive edilen mikroglialar, proenflamatuvar sitokinleri üreterek ve A β birikimine yanıt olarak nöronal hasara aracılık ederek AH patofizyolojisinde rol oynar. Mitokondriyal disfonksiyon da AH ile ilişkilendirilen bir başka önemli faktördür, çünkü mitokondriler reaktif oksijen türlerinin ana kaynağıdır. Metal iyonlarının özellikle bakır ve demir düzensizliğinin, AH ile ilişkilendirilen oksidatif stresi artırabileceği ve A β 'nin nörotoksitesini etkileyebileceği belirtilmiştir. AH'nin kompleks patogenezinde oksidatif stresin önemli bir rol oynadığı ve bu süreçlere müdahale eden terapötik stratejilere ihtiyaç olduğu vurgulanmıştır.

KAYNAKÇA

1. Botchway B, Iyer IC. Alzheimer's disease—the past, the present and the future. *Science*, 2017; 6, 1-19.
2. GBD 2019 Dementia Forecasting Collaborators. Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the Global Burden of Disease Study 2019. *Lancet Public Health*. 2022;7(2): e105-e125. doi: 10.1016/S2468-2667(21)00249-8.
3. Babcock KR, Page JS, Fallon JR, Webb AE. Adult hippocampal neurogenesis in aging and Alzheimer's disease. *Stem Cell Rep*. 2021; 16, 681-693.
4. Lee H, Casadesus G, Zhu X, Castellani RJ, McShea A, Perry G, Petersen RB, Bajic V, Smith MA. Cell cycle re-entry mediated neurodegeneration and its treatment role in the pathogenesis of Alzheimer's disease. *Neurochem Int*. 2009;54, 84-88.
5. Monteiro AR, Barbosa DJ, Remião F, Silva R. Alzheimer's disease: Insights and new prospects in disease pathophysiology, biomarkers and disease-modifying drugs. *Biochem Pharmacol*. 2023 May; 211:115522. doi: 10.1016/j.bcp.2023.115522.
6. Cassidy L, Fernandez F, Johnson JB, Naiker M, Owoola AG, Broszczak DA. Oxidative stress in Alzheimer's disease: A review on emergent natural polyphenolic therapeutics. *Complement Ther Med*. 2020 Mar; 49:102294. doi: 10.1016/j.ctim.2019.102294.
7. Ito F, Sono Y, Ito T. Measurement and Clinical Significance of Lipid Peroxidation as a Biomarker of Oxidative Stress: Oxidative Stress in Diabetes, Atherosclerosis, and Chronic Inflammation. *Antioxidants*. 2019; 8(3):72.
8. Forman HJ, Zhang H. Targeting oxidative stress in disease: promise and limitations of antioxidant therapy. *Nat Rev Drug Discov*. 2021 Sep;20(9):689-709. doi: 10.1038/s41573-021-00233-1.
9. Savelieff MG, Nam G, Kang J, Lee HJ, Lee M, Lim MH. Development of Multifunctional Molecules as Potential Therapeutic Candidates for Alzheimer's Disease, Parkin-

- son's Disease, and Amyotrophic Lateral Sclerosis in the Last Decade. *Chem Rev.* 2019 Jan 23;119(2):1221-1322. doi: 10.1021/acs.chemrev.8b00138.
10. Circu ML, Aw TY. Reactive oxygen species, cellular redox systems, and apoptosis. *Free Radic Biol Med.* 2010 Mar 15;48(6):749-62. doi: 10.1016/j.freeradbiomed.2009.12.022.
 11. Zhao Y, Zhao B. Oxidative stress and the pathogenesis of Alzheimer's disease. *Oxid Med Cell Longev.* 2013; 2013:316523. doi: 10.1155/2013/316523.
 12. Ganguly G, Chakrabarti S, Chatterjee U, Saso L. Proteinopathy, oxidative stress and mitochondrial dysfunction: cross talk in Alzheimer's disease and Parkinson's disease. *Drug Des Devel Ther.* 2017 Mar 16; 11:797-810. doi: 10.2147/DDDT.S130514.
 13. Huang X, Moir RD, Tanzi RE, Bush AI, Rogers JT. Redox-active metals, oxidative stress, and Alzheimer's disease pathology. *Ann N Y Acad Sci.* 2004 Mar; 1012:153-63. doi: 10.1196/annals.1306.012.
 14. Butterfield DA, Di Domenico F, Swomley AM, Head E, Perluigi M. Redox proteomics analysis to decipher the neurobiology of Alzheimer-like neurodegeneration overlaps in Down's syndrome and Alzheimer's disease brain. *Biochem J.* 2014 Oct 15;463(2):177-89. doi: 10.1042/BJ20140772.
 15. Butterfield DA, Swomley AM, Sultana R. Amyloid β -peptide (1-42)-induced oxidative stress in Alzheimer disease: importance in disease pathogenesis and progression. *Antioxid Redox Signal.* 2013 Sep 10;19(8):823-35. doi: 10.1089/ars.2012.5027.
 16. Rinaldi C, Donato L, Alibrandi S, Scimone C, D'Angelo R, Sidoti A. Oxidative Stress and the Neurovascular Unit. *Life (Basel).* 2021 Jul 29;11(8):767. doi: 10.3390/life11080767.
 17. Colonna M, Butovsky O. Microglia Function in the Central Nervous System During Health and Neurodegeneration. *Annu Rev Immunol.* 2017 Apr 26; 35:441-468. doi: 10.1146/annurev-immunol-051116-052358.
 18. Hansen DV, Hanson JE, Sheng M. Microglia in Alzheimer's disease. *J Cell Biol.* 2018 Feb 5;217(2):459-472. doi: 10.1083/jcb.201709069.
 19. Kwon HS, Koh SH. Neuroinflammation in neurodegenerative disorders: the roles of microglia and astrocytes. *Transl Neurodegener.* 2020 Nov 26;9(1):42. doi: 10.1186/s40035-020-00221-2.
 20. Cai Z, Zhao B, Ratka A. Oxidative stress and β -amyloid protein in Alzheimer's disease. *Neuromolecular Med.* 2011 Dec;13(4):223-50. doi: 10.1007/s12017-011-8155-9.
 21. Juźwik CA, S Drake S, Zhang Y, Paradis-Isler N, Sylvester A, Amar-Zifkin A, Douglas C, Morquette B, Moore CS, Fournier AE. microRNA dysregulation in neurodegenerative diseases: A systematic review. *Prog Neurobiol.* 2019 Nov;182:101664. doi: 10.1016/j.pneurobio.2019.101664.
 22. Acioglu C, Li L, Elkabes S. Contribution of astrocytes to neuropathology of neurodegenerative diseases. *Brain Res.* 2021 May 1; 1758:147291. doi: 10.1016/j.brainres.2021.147291.
 23. Escartin C, Galea E, Lakatos A, et al., Reactive astrocyte nomenclature, definitions, and future directions. *Nat Neurosci.* 2021 Mar;24(3):312-325. doi: 10.1038/s41593-020-00783-4.
 24. Sarkar S, Biswas SC. Astrocyte subtype-specific approach to Alzheimer's disease treatment. *Neurochem Int.* 2021 May; 145:104956. doi: 10.1016/j.neuint.2021.104956.

25. Arranz AM, De Strooper B. The role of astroglia in Alzheimer's disease: pathophysiology and clinical implications. *Lancet Neurol.* 2019 Apr;18(4):406-414. doi: 10.1016/S1474-4422(18)30490-3.
26. Hong P, Zhang X, Gao S, Wang P. Role of monocarboxylate transporter 4 in Alzheimer disease. *Neurotoxicology.* 2020 Jan; 76:191-199. doi: 10.1016/j.neuro.2019.11.006.
27. Liu B, Teschemacher AG, Kasparov S. Neuroprotective potential of astroglia. *J Neurosci Res.* 2017 Nov;95(11):2126-2139. doi: 10.1002/jnr.24140.
28. Veyrat-Durebex C, Corcia P, Piver E, et al., Disruption of TCA Cycle and Glutamate Metabolism Identified by Metabolomics in an In Vitro Model of Amyotrophic Lateral Sclerosis. *Mol Neurobiol.* 2016 Dec;53(10):6910-6924. doi: 10.1007/s12035-015-9567-6.
29. Perez-Nievas BG, Serrano-Pozo A. Deciphering the Astrocyte Reaction in Alzheimer's Disease. *Front Aging Neurosci.* 2018 Apr 25; 10:114. doi: 10.3389/fnagi.2018.00114.
30. Walker KA, Ficek BN, Westbrook R. Understanding the Role of Systemic Inflammation in Alzheimer's Disease. *ACS Chem Neurosci.* 2019 Aug 21;10(8):3340-3342. doi: 10.1021/acscemneuro.9b00333.
31. Ibrahim MM, Gabr MT. Multitarget therapeutic strategies for Alzheimer's disease. *Neural Regen Res.* 2019 Mar;14(3):437-440. doi: 10.4103/1673-5374.245463.
32. Jomova K, Baros S & Valko M. Redox active metal-induced oxidative stress in biological systems. *Transition Met Chem.* 2012; 37, 127-134. <https://doi.org/10.1007/s11243-012-9583-6>.
33. Gammella E, Buratti P, Cairo G, Recalcati S. The transferrin receptor: the cellular iron gate. *Metallomics.* 2017 Oct 18;9(10):1367-1375. doi: 10.1039/c7mt00143f.
34. Nakamura T, Naguro I, Ichijo H. Iron homeostasis and iron-regulated ROS in cell death, senescence and human diseases. *Biochim Biophys Acta Gen Subj.* 2019 Sep;1863(9):1398-1409. doi: 10.1016/j.bbagen.2019.06.010.
35. Wessling-Resnick M. Crossing the Iron Gate: Why and How Transferrin Receptors Mediate Viral Entry. *Annu Rev Nutr.* 2018 Aug 21; 38:431-458. doi: 10.1146/annurev-nutr-082117-051749.
36. Wang X, Su B, Lee HG, Li X, Perry G, Smith MA, Zhu X. Impaired balance of mitochondrial fission and fusion in Alzheimer's disease. *J Neurosci.* 2009 Jul 15;29(28):9090-103. doi: 10.1523/JNEUROSCI.1357-09.2009.
37. Devi L, Prabhu BM, Galati DF, Avadhani NG, Anandatheerthavarada HK. Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. *J Neurosci.* 2006 Aug 30;26(35):9057-68. doi: 10.1523/JNEUROSCI.1469-06.2006.
38. Cenini G, Voos W. Mitochondria as Potential Targets in Alzheimer Disease Therapy: An Update. *Front Pharmacol.* 2019 Aug 23; 10:902. doi: 10.3389/fphar.2019.00902.
39. Sharma C, Kim SR. Linking Oxidative Stress and Proteinopathy in Alzheimer's Disease. *Antioxidants (Basel).* 2021 Jul 30;10(8):1231. doi: 10.3390/antiox10081231.
40. Müller UC, Deller T, Korte M. Not just amyloid: physiological functions of the amyloid precursor protein family. *Nat Rev Neurosci.* 2017 May;18(5):281-298. doi: 10.1038/nrn.2017.29.
41. O'Brien RJ, Wong PC. Amyloid precursor protein processing and Alzheimer's disease. *Annu Rev Neurosci.* 2011; 34:185-204. doi: 10.1146/annurev-neuro-061010-113613.

42. Thinakaran G, Koo EH. Amyloid precursor protein trafficking, processing, and function. *J Biol Chem*. 2008 Oct 31;283(44):29615-9. doi: 10.1074/jbc.R800019200.
43. Carrillo-Mora P, Luna R, Colín-Barenque L. Amyloid beta: multiple mechanisms of toxicity and only some protective effects? *Oxid Med Cell Longev*. 2014;2014:795375. doi: 10.1155/2014/795375.
44. Nicholas M Kanaan, Diana S Himmelstein, Sarah M Ward, Benjamin Combs, Lester I Binder. *Tau Protein: Biology and Pathobiology*, Editor(s): Mark S. LeDoux, *Movement Disorders (Second Edition)*, Academic Press, 2015, 857-874.
45. Iqbal K, Alonso Adel C, Chen S, Chohan MO, El-Akkad E, Gong CX, Khatoon S, Li B, Liu F, Rahman A, Tanimukai H, Grundke-Iqbal I. Tau pathology in Alzheimer disease and other tauopathies. *Biochim Biophys Acta*. 2005 Jan 3;1739(2-3):198-210. doi: 10.1016/j.bbadis.2004.09.008.
46. Yang K, Chen Z, Gao J, Shi W, Li L, Jiang S, Hu H, Liu Z, Xu D, Wu L. The Key Roles of GSK-3 β in Regulating Mitochondrial Activity. *Cell Physiol Biochem*. 2017;44(4):1445-1459. doi: 10.1159/000485580.
47. Souder DC, Anderson RM. An expanding GSK3 network: implications for aging research. *Geroscience*. 2019 Aug;41(4):369-382. doi: 10.1007/s11357-019-00085-z.
48. Sinha K, Das J, Pal PB, Sil PC. Oxidative stress: the mitochondria-dependent and mitochondria-independent pathways of apoptosis. *Arch Toxicol*, 2013; 87 (7), 1157-1180.
49. Singh A, Kukreti R, Saso L, Kukreti S. Oxidative stress: a key modulator in neurodegenerative diseases. *Molecules*,2019;24 (8).
50. Islam MT. Oxidative stress and mitochondrial dysfunction-linked neurodegenerative disorders. *Neurol Res*, 2017;39 (1), 73-82.
51. Sies H, Berndt C, Jones DP. Oxidative stress. *Annu Rev Biochem* ,2017; 86, 715-748.
52. Lee J, Giordano S, Zhang J. Autophagy, mitochondria and oxidative stress: crosstalk and redox signalling. *Biochem J*, 2012;441 (2), 523-540.
53. Maynard S, Fang EF, Scheibye-Knudsen M, Croteau DL, Bohr VA. DNA damage. DNA Repair, Aging, Neurodegeneration. *Cold Spring Harb Perspect Med*, 2015;5 (10).
54. Guillaumet-Adkins A, Yañez Y, Peris-Diaz MD, Calabria I, Palanca-Ballester C, Sandoval J. Epigenetics and oxidative stress in aging. *Oxid Med Cell Longev*, 2017; 9175806.
55. Grimm A, Eckert A. Brain aging and neurodegeneration: from a mitochondrial point of view. *J Neurochem*, 2017;143 (4), 418-431.
56. Lu T, Pan Y, Kao SY, Li C, Kohane I, Chan J, Yankner BA. Gene regulation and DNA damage in the ageing human brain. *Nature*, 2004;429 (6994), 883-891.
57. Mecocci P, Boccardi V, Cecchetti R, Bastiani P, Scamosci M, Ruggiero C, Baroni M. A long journey into aging, brain aging, and Alzheimer's disease following the oxidative stress tracks. *J Alzheimers Dis*, 2018;62 (3), 1319-1335.
58. Pérez VI, Van Remmen H, Bokov A, Epstein CJ, Vijg J, Richardson A. The overexpression of major antioxidant enzymes does not extend the lifespan of mice. *Aging Cell*, 2009;8 (1), 73-75.
59. Tower J. Transgenic methods for increasing *Drosophila* life span. *Mech Ageing Dev*,2000; 118 (1-2), 1-14.
60. Dai DF, Santana LF, Vermulst M, Tomazela DM, Emond MJ, MacCoss MJ, Gollahon K, Martin GM, Loeb LA, Ladiges WC, Rabinovitch PS. Overexpression of catalase

- targeted to mitochondria attenuates murine cardiac aging. *Circulation*, 2009;119 (21), 2789–2797.
61. Paglialunga S, Ludzki A, Root-McCaig J, Holloway GP. In adipose tissue, increased mitochondrial emission of reactive oxygen species is important for short-term high-fat diet-induced insulin resistance in mice. *Diabetologia*, 2015;58 (5), 1071–1080.
 62. Schriener SE, Linford NJ, Martin GM, Treuting P, Ogburn CE, Emond M, Coskun PE, Ladiges W, Wolf N, Van Remmen H, Wallace DC, Rabinovitch PS. Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science*, 2005;308 (5730), 1909–1911.
 63. Campisi J, Kapahi P, Lithgow GJ, Melov S, Newman JC, Verdin E. From discoveries in ageing research to therapeutics for healthy ageing. *Nature*, 2019;571 (7764), 183–192.
 64. Kim SJ, Cheresch P, Jablonski RP, Morales-Nebreda L, Cheng Y, Hogan E, Yeldandi A, Chi M, Piseaux R, Ridge K, Hart Michael C, Chandel N, Scott Budinger GR, Kamp DW. Mitochondrial catalase overexpressed transgenic mice are protected against lung fibrosis in part via preventing alveolar epithelial cell mitochondrial DNA damage. *Free Radic Biol Med*, 2016;101, 482–490.
 65. Swerdlow RH, Burns JM, Khan SM. The Alzheimer's disease mitochondrial cascade hypothesis: progress and perspectives. *Biochim Biophys Acta*, 2014;1842 (8), 1219–1231.
 66. Cheignon C, Tomas M, Bonnefont-Rousselot D, Faller P, Hureau C, Collin F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol*, 2018;14, 450–464.
 67. Hensley K, Carney JM, Mattson MP, Aksenova M, Harris M, Wu JF, Floyd RA, Butterfield DA. A model for beta-amyloid aggregation and neurotoxicity based on free radical generation by the peptide: relevance to Alzheimer disease. *Proc Natl Acad Sci U S A*. 1994 Apr 12;91(8):3270-4. doi: 10.1073/pnas.91.8.3270.
 68. Pike CJ, Cummings BJ, Cotman CW. beta-Amyloid induces neuritic dystrophy in vitro: similarities with Alzheimer pathology. *Neuroreport*. 1992 Sep;3(9):769-72. doi: 10.1097/00001756-199209000-00012.
 69. Mark RJ, Lovell MA, Markesbery WR, Uchida K, Mattson MP. A role for 4-hydroxynonenal, an aldehydic product of lipid peroxidation, in disruption of ion homeostasis and neuronal death induced by amyloid beta-peptide. *J Neurochem*. 1997 Jan;68(1):255-64. doi: 10.1046/j.1471-4159.1997.68010255.x.
 70. Butterfield DA. Brain lipid peroxidation and Alzheimer disease: Synergy between the Butterfield and Mattson laboratories. *Ageing Res Rev*. 2020 Dec; 64:101049. doi: 10.1016/j.arr.2020.101049.
 71. Di Domenico F, Tramutola A, Butterfield DA. Role of 4-hydroxy-2-nonenal (HNE) in the pathogenesis of Alzheimer disease and other selected age-related neurodegenerative disorders. *Free Radic Biol Med*. 2017 Oct; 111:253-261. doi: 10.1016/j.freeradbiomed.2016.10.490.
 72. Pike CJ, Burdick D, Walencewicz AJ, Glabe CG, Cotman CW. Neurodegeneration induced by beta-amyloid peptides in vitro: the role of peptide assembly state. *J Neurosci*. 1993 Apr;13(4):1676-87. doi: 10.1523/JNEUROSCI.13-04-01676.1993.
 73. Keller JN, Mark RJ, Bruce AJ, Blanc E, Rothstein JD, Uchida K, Waeg G, Mattson MP. 4-Hydroxynonenal, an aldehydic product of membrane lipid peroxidation, impairs glutamate transport and mitochondrial function in synaptosomes. *Neuroscience*. 1997 Oct;80(3):685-96. doi: 10.1016/s0306-4522(97)00065-1.

74. Tönnies E, Trushina E. Oxidative Stress, Synaptic Dysfunction, and Alzheimer's Disease. *J Alzheimers Dis.* 2017;57(4):1105-1121. doi: 10.3233/JAD-161088.
75. Abolhassani N, Leon J, Sheng Z, Oka S, Hamasaki H, Iwaki T, Nakabeppu Y. Molecular pathophysiology of impaired glucose metabolism, mitochondrial dysfunction, and oxidative DNA damage in Alzheimer's disease brain. *Mech Ageing Dev.* 2017 Jan;161(Pt A):95-104. doi: 10.1016/j.mad.2016.05.005.
76. Nakamura T, Cieplak P, Cho DH, Godzik A, Lipton SA. S-nitrosylation of Drp1 links excessive mitochondrial fission to neuronal injury in neurodegeneration. *Mitochondrion.* 2010 Aug;10(5):573-8. doi: 10.1016/j.mito.2010.04.007.
77. Khandelwal PJ, Herman AM, Hoe HS, Rebeck GW, Moussa CE. Parkin mediates beclin-dependent autophagic clearance of defective mitochondria and ubiquitinated Abeta in AD models. *Hum Mol Genet.* 2011 Jun 1;20(11):2091-102. doi: 10.1093/hmg/ddr091.
78. Martín-Maestro P, Gargini R, Perry G, Avila J, García-Escudero V. PARK2 enhancement is able to compensate mitophagy alterations found in sporadic Alzheimer's disease. *Hum Mol Genet.* 2016 Feb 15;25(4):792-806. doi: 10.1093/hmg/ddv616.
79. Tan S, Sagara Y, Liu Y, Maher P, Schubert D. The regulation of reactive oxygen species production during programmed cell death. *J Cell Biol.* 1998 Jun 15;141(6):1423-32. doi: 10.1083/jcb.141.6.1423.
80. Grivennikova VG, Vinogradov AD. Generation of superoxide by the mitochondrial Complex I. *Biochim Biophys Acta.* 2006 May-Jun;1757(5-6):553-61. doi: 10.1016/j.bbabi.2006.03.013.
81. Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, Johnson AB, Kress Y, Vinters HV, Tabaton M, Shimohama S, Cash AD, Siedlak SL, Harris PL, Jones PK, Petersen RB, Perry G, Smith MA. Mitochondrial abnormalities in Alzheimer's disease. *J Neurosci.* 2001 May 1;21(9):3017-23. doi: 10.1523/JNEUROSCI.21-09-03017.2001.
82. Zhu X, Perry G, Moreira PI, Aliev G, Cash AD, Hirai K, Smith MA. Mitochondrial abnormalities and oxidative imbalance in Alzheimer disease. *J Alzheimers Dis.* 2006 Jul;9(2):147-53. doi: 10.3233/jad-2006-9207.
83. Mutisya EM, Bowling AC, Beal MF. Cortical cytochrome oxidase activity is reduced in Alzheimer's disease. *J Neurochem.* 1994 Dec;63(6):2179-84. doi: 10.1046/j.1471-4159.1994.63062179.x.
84. Manczak M, Anekonda TS, Henson E, Park BS, Quinn J, Reddy PH. Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: implications for free radical generation and oxidative damage in disease progression. *Hum Mol Genet.* 2006 May 1;15(9):1437-49. doi: 10.1093/hmg/ddl066.
85. Caspersen C, Wang N, Yao J, Sosunov A, Chen X, Lustbader JW, Xu HW, Stern D, McKhann G, Yan SD. Mitochondrial Abeta: a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease. *FASEB J.* 2005 Dec;19(14):2040-1. doi: 10.1096/fj.05-3735fj.
86. Rodrigues CM, Solá S, Brito MA, Brondino CD, Brites D, Moura JJ. Amyloid beta-peptide disrupts mitochondrial membrane lipid and protein structure: protective role of tauroursodeoxycholate. *Biochem Biophys Res Commun.* 2001 Feb 23;281(2):468-74. doi: 10.1006/bbrc.2001.4370.
87. Casley CS, Canevari L, Land JM, Clark JB, Sharpe MA. Beta-amyloid inhibits integrated mitochondrial respiration and key enzyme activities. *J Neurochem.* 2002 Jan;80(1):91-100. doi: 10.1046/j.0022-3042.2001.00681.x.

88. Anantharaman M, Tangpong J, Keller JN, Murphy MP, Markesbery WR, Kiningham KK, St Clair DK. Beta-amyloid mediated nitration of manganese superoxide dismutase: implication for oxidative stress in a APPNLH/NLH X PS-1P264L/P264L double knock-in mouse model of Alzheimer's disease. *Am J Pathol.* 2006 May;168(5):1608-18. doi: 10.2353/ajpath.2006.051223.
89. Rousset S, Alves-Guerra MC, Mozo J, Miroux B, Cassard-Doulcier AM, Bouillaud F, Ricquier D. The biology of mitochondrial uncoupling proteins. *Diabetes.* 2004 Feb;53 Suppl 1:S130-5. doi: 10.2337/diabetes.53.2007.s130.
90. Echtay KS. Mitochondrial uncoupling proteins--what is their physiological role? *Free Radic Biol Med.* 2007 Nov 15;43(10):1351-71. doi: 10.1016/j.freeradbiomed.2007.08.011.
91. de la Monte SM, Wands JR. Molecular indices of oxidative stress and mitochondrial dysfunction occur early and often progress with severity of Alzheimer's disease. *J Alzheimers Dis.* 2006 Jul;9(2):167-81. doi: 10.3233/jad-2006-9209.
92. Wu Z, Zhang J, Zhao B. Superoxide anion regulates the mitochondrial free Ca²⁺ through uncoupling proteins. *Antioxid Redox Signal.* 2009 Aug;11(8):1805-18. doi: 10.1089/ars.2009.2427
93. Schlieff ML, Gitlin JD. Copper homeostasis in the CNS: a novel link between the NMDA receptor and copper homeostasis in the hippocampus. *Mol Neurobiol.* 2006 Apr;33(2):81-90. doi: 10.1385/MN:33:2:81.
94. Smart TG, Hosie AM, Miller PS. Zn²⁺ ions: modulators of excitatory and inhibitory synaptic activity. *Neuroscientist.* 2004 Oct;10(5):432-42. doi: 10.1177/1073858404263463.
95. Muñoz P, Humeres A. Iron deficiency on neuronal function. *Biometals.* 2012 Aug;25(4):825-35. doi: 10.1007/s10534-012-9550-x.
96. Kenche VB, Barnham KJ. Alzheimer's disease & metals: therapeutic opportunities. *Br J Pharmacol.* 2011;163(2):211-219. doi:10.1111/j.1476-5381.2011.01221.x.
97. Deibel MA, Ehmann WD, Markesbery WR. Copper, iron, and zinc imbalances in severely degenerated brain regions in Alzheimer's disease: possible relation to oxidative stress. *J Neurol Sci.* 1996 Nov;143(1-2):137-42. doi: 10.1016/s0022-510x(96)00203-1.
98. Lovell MA, Robertson JD, Teesdale WJ, Campbell JL, Markesbery WR. Copper, iron and zinc in Alzheimer's disease senile plaques. *J Neurol Sci.* 1998 Jun 11;158(1):47-52. doi: 10.1016/s0022-510x(98)00092-6.
99. Lee JY, Mook-Jung I, Koh JY. Histochemically reactive zinc in plaques of the Swedish mutant beta-amyloid precursor protein transgenic mice. *J Neurosci.* 1999 Jun 1;19(11):RC10. doi: 10.1523/JNEUROSCI.19-11-j0002.1999.
100. Zhang J, Liu Q, Chen Q, Liu NQ, Li FL, Lu ZB, Qin C, Zhu H, Huang YY, He W, Zhao BL. Nicotine attenuates beta-amyloid-induced neurotoxicity by regulating metal homeostasis. *FASEB J.* 2006 Jun;20(8):1212-4. doi: 10.1096/fj.05-5214fe.
101. Curtain CC, Ali F, Volitakis I, et al. Alzheimer's disease amyloid-beta binds copper and zinc to generate an allosterically ordered membrane-penetrating structure containing superoxide dismutase-like subunits. *J Biol Chem.* 2001 Jun 8;276(23):20466-73. doi: 10.1074/jbc.M100175200.
102. Hesse L, Behr D, Masters CL, Multhaup G. The beta A4 amyloid precursor protein binding to copper. *FEBS Lett.* 1994 Jul 25;349(1):109-16. doi: 10.1016/0014-5793(94)00658-x.

103. Atwood CS, Moir RD, Huang X, et al. Dramatic aggregation of Alzheimer abeta by Cu (II) is induced by conditions representing physiological acidosis. *J Biol Chem.* 1998 May 22;273(21):12817-26. doi: 10.1074/jbc.273.21.12817.
104. Bush AI, Pettingell WH, Multhaup G, d Paradis M, Vonsattel JP, Gusella JF, Beyreuther K, Masters CL, Tanzi RE. Rapid induction of Alzheimer A beta amyloid formation by zinc. *Science.* 1994 Sep 2;265(5177):1464-7. doi: 10.1126/science.8073293.
105. Strausak D, Mercer JF, Dieter HH, Stremmel W, Multhaup G. Copper in disorders with neurological symptoms: Alzheimer's, Menkes, and Wilson diseases. *Brain Res Bull.* 2001 May 15;55(2):175-85. doi: 10.1016/s0361-9230(01)00454-3.
106. Opazo C, Huang X, Cherny RA, Moir RD, Roher AE, White AR, Cappai R, Masters CL, Tanzi RE, Inestrosa NC, Bush AI. Metalloenzyme-like activity of Alzheimer's disease beta-amyloid. Cu-dependent catalytic conversion of dopamine, cholesterol, and biological reducing agents to neurotoxic H(2)O(2). *J Biol Chem.* 2002 Oct 25;277(43):40302-8. doi: 10.1074/jbc.M206428200.
107. Opazo C, Huang X, Cherny RA, Moir RD, Roher AE, White AR, Cappai R, Masters CL, Tanzi RE, Inestrosa NC, Bush AI. Metalloenzyme-like activity of Alzheimer's disease beta-amyloid. Cu-dependent catalytic conversion of dopamine, cholesterol, and biological reducing agents to neurotoxic H(2)O(2). *J Biol Chem.* 2002 Oct 25;277(43):40302-8. doi: 10.1074/jbc.M206428200.
108. Huang X, Cuajungco MP, Atwood CS, et al. Cu (II) potentiation of alzheimer abeta neurotoxicity. Correlation with cell-free hydrogen peroxide production and metal reduction. *J Biol Chem.* 1999 Dec 24;274(52):37111-6. doi: 10.1074/jbc.274.52.37111.
109. Huang X, Atwood CS, Hartshorn MA, et al. The A beta peptide of Alzheimer's disease directly produces hydrogen peroxide through metal ion reduction. *Biochemistry.* 1999 Jun 15;38(24):7609-16. doi: 10.1021/bi990438f.
110. Lynch T, Cherny RA, Bush AI. Oxidative processes in Alzheimer's disease: the role of abeta-metal interactions. *Exp Gerontol.* 2000 Jul;35(4):445-51. doi: 10.1016/s0531-5565(00)00112-1.
111. Rottkamp CA, Raina AK, Zhu X, Gaier E, Bush AI, Atwood CS, Chevion M, Perry G, Smith MA. Redox-active iron mediates amyloid-beta toxicity. *Free Radic Biol Med.* 2001 Feb 15;30(4):447-50. doi: 10.1016/s0891-5849(00)00494-9.
112. Wan L, Nie G, Zhang J, Luo Y, Zhang P, Zhang Z, Zhao B. β -Amyloid peptide increases levels of iron content and oxidative stress in human cell and *Caenorhabditis elegans* models of Alzheimer disease. *Free Radic Biol Med.* 2011 Jan 1;50(1):122-9. doi: 10.1016/j.freeradbiomed.2010.10.707.
113. Zheng W, Xin N, Chi ZH, Zhao BL, Zhang J, Li JY, Wang ZY. Divalent metal transporter 1 is involved in amyloid precursor protein processing and Abeta generation. *FASEB J.* 2009 Dec;23(12):4207-17. doi: 10.1096/fj.09-135749.
114. Zhang LH, Wang X, Zheng ZH, Ren H, Stoltenberg M, Danscher G, Huang L, Rong M, Wang ZY. Altered expression and distribution of zinc transporters in APP/PS1 transgenic mouse brain. *Neurobiol Aging.* 2010 Jan;31(1):74-87. doi: 10.1016/j.neurobiolaging.2008.02.018.
115. Zhang LH, Wang X, Stoltenberg M, Danscher G, Huang L, Wang ZY. Abundant expression of zinc transporters in the amyloid plaques of Alzheimer's disease brain. *Brain Res Bull.* 2008 Sep 5;77(1):55-60. doi: 10.1016/j.brainresbull.2008.03.014.

116. Li H, Li F, Sun H, Qian ZM. Membrane-inserted conformation of transmembrane domain 4 of divalent-metal transporter. *Biochem J.* 2003;372(Pt 3):757-766. doi:10.1042/BJ20030075.
117. Bellingham SA, Ciccotosto GD, Needham BE, et al. Gene knockout of amyloid precursor protein and amyloid precursor-like protein-2 increases cellular copper levels in primary mouse cortical neurons and embryonic fibroblasts. *J Neurochem.* 2004 Oct;91(2):423-8. doi: 10.1111/j.1471-4159.2004.02731.x.
118. Acevedo KM, Hung YH, Dalziel AH, et al. Copper promotes the trafficking of the amyloid precursor protein. *J Biol Chem.* 2011;286(10):8252-8262. doi:10.1074/jbc.M110.128512.
119. White AR, Reyes R, Mercer JF, et al. Copper levels are increased in the cerebral cortex and liver of APP and APLP2 knockout mice. *Brain Res.* 1999 Sep 25;842(2):439-44. doi: 10.1016/s0006-8993(99)01861-2.
120. Bolognin S, Messori L, Zatta P. Metal ion physiopathology in neurodegenerative disorders. *Neuromolecular Med.* 2009;11(4):223-38. doi: 10.1007/s12017-009-8102-1.
121. Cherny RA, Atwood CS, Xilinas ME, et al. Treatment with a copper-zinc chelator markedly and rapidly inhibits beta-amyloid accumulation in Alzheimer's disease transgenic mice. *Neuron.* 2001 Jun;30(3):665-76. doi: 10.1016/s0896-6273(01)00317-8.
122. Ritchie CW, Bush AI, Mackinnon A, Macfarlane S, Mastwyk M, MacGregor L, Kiers L, Cherny R, Li QX, Tammer A, Carrington D, Mavros C, Volitakis I, Xilinas M, Ames D, Davis S, Beyreuther K, Tanzi RE, Masters CL. Metal-protein attenuation with iodochlorhydroxyquin (clioquinol) targeting Abeta amyloid deposition and toxicity in Alzheimer disease: a pilot phase 2 clinical trial. *Arch Neurol.* 2003 Dec;60(12):1685-91. doi: 10.1001/archneur.60.12.1685.
123. Zhao Y, Zhao B. Oxidative stress and the pathogenesis of Alzheimer's disease. *Oxid Med Cell Longev.* 2013; 2013:316523. doi:10.1155/2013/316523.
124. Kreuz S, Fischle W. Oxidative stress signaling to chromatin in health and disease. *Epigenomics.* 2016;8(6):843-862. doi:10.2217/epi-2016-0002.
125. Chia N, Wang L, Lu X, Senut MC, Brenner C, Ruden DM. Hypothesis: environmental regulation of 5-hydroxymethylcytosine by oxidative stress. *Epigenetics.* 2011; 6(7):853-6. doi: 10.4161/epi.6.7.16461.
126. Thanan R, Oikawa S, Hiraku Y, Ohnishi S, Ma N, Pinlaor S, Yongvanit P, Kawanishi S, Murata M. Oxidative stress and its significant roles in neurodegenerative diseases and cancer. *Int J Mol Sci.* 2014 Dec 24;16(1):193-217. doi: 10.3390/ijms16010193.
127. Lewandowska J, Bartoszek A. DNA methylation in cancer development, diagnosis and therapy--multiple opportunities for genotoxic agents to act as methylome disruptors or remediators. *Mutagenesis.* 2011 Jul;26(4):475-87. doi: 10.1093/mutage/ger019.
128. Niu Y, DesMarais TL, Tong Z, Yao Y, Costa M. Oxidative stress alters global histone modification and DNA methylation. *Free Radic Biol Med.* 2015; 82:22-28. doi: 10.1016/j.freeradbiomed.2015.01.028.
129. Gu X, Sun J, Li S, Wu X, Li L. Oxidative stress induces DNA demethylation and histone acetylation in SH-SY5Y cells: potential epigenetic mechanisms in gene transcription in A β production. *Neurobiol Aging.* 2013; 34(4):1069-79. doi: 10.1016/j.neurobiolaging.2012.10.013.

Bölüm 4

İNME BİYOBELİRTEÇLERİ

Yeşim GÜVENÇ DEMİRAĞCI¹

GİRİŞ

İnme, merkezi sinir sisteminin vasküler hasarına atfedilen ve klinik olarak tanımlanmış akut, fokal nörolojik defisit sendromudur. Beynin fokal hasarına bağlı olarak hızla gelişmiş olan klinik bulgular 24 saatten daha fazla sürmektedir. Ülkemizde ve Batı'da morbiditenin ve mortalitenin üçüncü nedenidir. Hayatta kalan hastalar beş yıl içinde yeniden inme geçirmektedir. Yaşlıların hastalığı olarak sınırlandırılmamalıdır. Bütün yaşlar inme açısından risk altındadır (1).

İNME SINIFLANDIRMASI

İki çeşit inme tipi bulunmaktadır:

İskemik İnme

Hemorajik İnme

İnmelerin %80'ini iskemik nedenler oluşturmaktadır. Bunların %50'si intrakraniyal tromboz (ateroskleroz), %30'u da ekstrakraniyal embolilerle meydana gelmektedir. Hemorajik inmeler (%20) intraserebral kanama (ISK) ve subaraknoid kanama (SAK) nedeniyle meydana gelmektedir. İntraserebral kanamalar çoğunlukla rüptür ve hematoma oluşturan zayıflamış damarlardan meydana gelmektedir. Subaraknoid kanamalarda kanama beyin dışına ya da BOS içine olmaktadır (2).

Geçici iskemik atak (GİA) mini-stroke olarak adlandırılmaktadır. İskemik inmeye benzemekle birlikte fokal nörolojik defisitler 24 saatten kısa sürmektedir (1).

İNME FİZYOPATOLOJİSİ

İskemik olaylarda serebral hipoperfüzyon, hücrel biyoenerji yetmezliği, oksidatif hasar, kan-beyin bariyeri disfonksiyonu, mikrovasküler hasar, hemostatik

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KAYNAKÇA

1. Sacco RL, Kasner SE, Broderick JP, et al. An updated definition of stroke for the 21st century: a statement for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke*. 2013;44(7): 2064-2089. doi: 10.1161/STR.0b013e318296aeca
2. Johnston SC, Mendis S, Mathers CD. Global variation in stroke burden and mortality: Estimates from monitoring, surveillance, and modelling. *Lancet Neurol*. 2009;8(4):345-354. doi: 10.1016/S1474-4422(09)70023-7.
3. Brouns R, De Deyn PP. The complexity of neurobiological processes in acute ischemic stroke. *Clin Neurol Neurosurg*. 2009; 111(6):483-495. doi: 10.1016/j.clin-neuro.2009.04.001.
4. Xi G, Keep RF, Hoff JT. Mechanisms of brain injury after intracerebral haemorrhage. *The Lancet. Neurology*. 2006;5(1):53-63. doi: 10.1016/S1474-4422(05)70283-0.
5. Meschia JF, Bushnell C, Boden-Albala B, et al. Guidelines for the Primary Prevention of Stroke. A Statement for Healthcare Professionals From the American Heart Association/American Stroke Association. *Stroke*. 2014;45(12):3754-3832. doi: 10.1161/STR.0000000000000046.
6. Wityk RJ, Beauchamp NJ Jr. Diagnostic evaluation of stroke. *Neurol Clin*. 2000;18(2):357-378. doi: 10.1016/s0733-8619(05)70197-3.
7. Strimbu K, Tavel JA. What are biomarkers? *Curr Opin HIV AIDS*. 2010;5(6):463-466. doi: 10.1097/COH.0b013e32833ed177.
8. Whiteley W, Tseng MC, Sandercock P. Blood biomarkers in the diagnosis of ischemic stroke: a systematic review. *Stroke*. 2008; 39(10):2902-2909. doi: 10.1161/STROKEA-HA.107.511261.
9. Baudier J, Glasser N, Gerard D. Ions binding to S100 proteins. I. Calcium- and zinc-binding properties of bovine brain S100 alpha alpha, S100a (alpha beta), and S100b (beta beta) protein: Zn²⁺ regulates Ca²⁺ binding on S100b protein. *J Biol Chem*. 1986; 261:8192-8203.
10. Elting JW, de Jager AE, Teelken AW, et al. Comparison of serum S-100 protein levels following stroke and traumatic brain injury. *J Neurol Sci*. 2000;181(1-2):104-110. doi: 10.1016/s0022-510x(00)00442-1.
11. Koh SX, Lee JK. S100B as a marker for brain damage and blood-brain barrier disruption following exercise. *Sports Med*. 2014;44(3):369-385. doi: 10.1007/s40279-013-0119-9.
12. Eng LF. Glial fibrillary acidic protein (GFAP): The major protein of glial intermediate filaments in differentiated astrocytes. *J Neuroimmunol*. 1985;8(4-6):203-214. doi: 10.1016/s0165-5728(85)80063-1.
13. Schiff L, Hadker N, Weiser S, et al. A literature review of the feasibility of glial fibrillary acidic protein as a biomarker for stroke and traumatic brain injury. *Mol Diagn Ther*. 2012;16(2):79-92. doi: 10.2165/11631580-000000000-00000.
14. Isgro MA, Bottoni P, Scatena R. Neuron-Specific Enolase as a Biomarker: Biochemical and Clinical Aspects. *Adv Exp Med Biol*. 2015; 867:125-143. doi: 10.1007/978-94-017-7215-0_9.
15. Mochetti MM, Silva EGP, Correa AAF, et al. Neuron-specific enolase at admission as a predictor for stroke volume, severity and outcome in ischemic stroke patients: a prognostic biomarker review. *Sci Rep*. 2024;14(1):2688. doi: 10.1038/s41598-024-53080-6.

16. Ramos-Fernandez M, Bellolio MF, Stead LG. Matrix metalloproteinase-9 as a marker for acute ischemic stroke: a systematic review *J Stroke Cerebrovasc Dis.* 2011; 20(1):47-54. doi: 10.1016/j.jstrokecerebrovasdis.2009.10.008.
17. Castellanos M, Leira R, Serena J, et al. Plasma metalloproteinase-9 concentration predicts hemorrhagic transformation in acute ischemic stroke. *Stroke.* 2003; 34(1):40-46.
18. Lonn E. Lipoprotein-associated phospholipase A2: a new therapeutic target. *Can J Cardiol.* 2010; 26 Suppl A:27A-31A. doi: 10.1016/s0828-282x(10)71058-8.
19. Elkind MS, Tai W, Coates K, et al. High-sensitivity C-reactive protein, lipoprotein-associated phospholipase A2, and outcome after ischemic stroke. *Arch Intern Med.* 2006; 166(19):2073-2080. doi: 10.1001/archinte.166.19.2073.
20. Shufen Chen I, Na Li, Milani Deb-Chatterji, Qiang Dong, Jan T Kielstein, Karin Weisenborn, Hans Worthmann. Asymmetric dimethylarginine as marker and mediator in ischemic stroke. *Int J Mol Sci.* 2012; 13(12):15983-16004. doi: 10.3390/ijms131215983.
21. Furukawa H, Singh SK, Mancusso R, et al. Subunit arrangement and function in NMDA receptors. *Nature.* 2005;438(7065):185-192. doi: 10.1038/nature04089.
22. Dambinova SA, Bettermann K, Glynn T, et al. Diagnostic potential of the NMDA receptor peptide assay for acute ischemic stroke. *PLoS One.* 2012;7(7): e42362. doi: 10.1371/journal.pone.0042362.
23. Allard L, Burkhard PR, Lescuyer P, Burgess JA, Walter N, Hochstrasser DF, Sanchez JC. PARK7 and nucleoside diphosphate kinase A as plasma markers for the early diagnosis of stroke. *Clin Chem.* 2005; 51(11):2043-2051. doi: 10.1373/clinchem.2005.053942.
24. Glushakova OY, Glushakov AV, Miller ER, Valadka AB, Hayes RL. Biomarkers for acute diagnosis and management of stroke in neurointensive care units. *Brain Circ.* 2016; 2(1):28-47. doi: 10.4103/2394-8108.178546.
25. Jickling GC, Sharp FR. Biomarker panels in ischemic stroke. *Stroke.* 2015;46(3):915-920. doi: 10.1161/STROKEAHA.114.005604.
26. Flint AC, Banki NM, Ren X, Rao VA, Go AS. Detection of paroxysmal atrial fibrillation by 30-day event monitoring in cryptogenic ischemic stroke: the Stroke and Monitoring for PAF in Real Time (SMART) Registry. *Stroke.* 2012; 43:2788-2790. doi: 10.1161/STROKEAHA.112.665844.
27. Llombart V, Antolin-Fontes A, Bustamante A, et al. B-type natriuretic peptides help in cardioembolic stroke diagnosis: pooled data meta-analysis. *Stroke.* 2015;46(5):1187-1195. doi: 10.1161/STROKEAHA.114.008311.
28. Bos MJ, Schipper CM, Koudstaal PJ, et al. High serum C-reactive protein level is not an independent predictor for stroke: the Rotterdam Study. *Circulation.* 2006;114(15):1591-1598. doi:10.1161/CIRCULATIONAHA.106.619833.
29. Isenegger J, Meier N, Lämmle B, et al. D-dimers predict stroke subtype when assessed early. *Cerebrovasc Dis.* 2010; 29(1):82-86. doi: 10.1159/000256652.
30. Matosevic B, Knoflach M, Werner P, et al. Fibrinogen degradation coagulopathy and bleeding complications after stroke thrombolysis. *Neurology.* 2013;80(13):1216-1224. doi: 10.1212/WNL.0b013e3182897015
31. Barba L, Vollmuth C, Abu-Rumeileh S, et al. Serum β -synuclein, neurofilament light chain and glial fibrillary acidic protein as prognostic biomarkers in moderate-to-severe acute ischemic stroke. *Sci Rep.* 2023;13(1):20941. doi: 10.1038/s41598-023-47765-7.
32. Fakhari MS, Poorsaadat L, Almasi-Hashiani A, et al. Inflammatory markers and functional outcome score in different subgroups of ischaemic stroke: a prospective cohort

- study. *BMJ Neurology Open*. 2024;6(1):e000556. doi: 10.1136/bmjno-2023-000556. eCollection 2024.DOI: 10.1136/bmjno-2023-000556.
33. Chu M, Niu H, Yang N, et al. High serum lactate dehydrogenase to albumin ratio is associated with increased risk of poor prognosis after ischemic stroke. *Clin Neurol Neurosurg*. 2024; 237:108120. doi: 10.1016/j.clineuro.2024.108120.
 34. Zaharia AL, Tutunaru D, Oprea VD, et al. Thrombomodulin serum levels- a predictable biomarker for the acute onset of ischemic stroke. *Curr Issues Mol Biol*. 2024;46(1):677-688. doi: 10.3390/cimb46010044.
 35. Wang T, Zhao W, Liu Y, et al. Differentially expressed miR-511-3p in stroke patients predicts the present of post-stroke cognitive impairment. *Dement Geriatr Cogn Disord*. 2023 Dec 6. doi: 10.1159/000535631.
 36. Mainali S, Nepal G, Webb A, et al. MicroRNA Expression profile in acute ischemic stroke. *Res Sq*. [Preprint]. 2024 Jan 3: rs.3.rs-3754883. doi: 10.21203/rs.3.rs-3754883/v1.

Bölüm 5

KANSERDE TERAPÖTİK HEDEF OLARAK AXL RESEPTÖRÜ VE AXL İNHİBİTÖRLERİ

Hatibe KARA¹

GİRİŞ

Kanser hücrelerinde ve tümör mikro çevresinde farklı sinyal yollarının düzensiz işlediği görülür. Transmembran proteinler olarak karakterize edilen Reseptör Tirozin Kinaz (RTK)'lerin anormallikleri de birçok malignitede yer almaktadır. Bir RTK üyesi olan AXL'nin aşırı ekspresyonu kanserde kötü prognoz ve tümör büyümesiyle ilişkilidir. AXL, K vitaminine bağımlı protein ailesinde yer alan Büyüme Durdurma Spesifik Protein 6 (GAS6)'nın bağlanmasıyla aktive olur. GAS6/AXL sinyal yolu PI3K/AKT, MAPK/ERK ve STAT3 dahil olmak üzere aşağı akış yollarını aktive ederek kanserin ilerlemesi, metastazı, kemoterapötik direnci ve immün bağışıklıktan kaçınmayı teşvik eder. Kanserinin farklı süreçlerinden sorumlu tutuluyor olması AXL'yi kemoterapötik hedef haline getirmiş ve AXL'a özgü pek çok terapötik ajan geliştirilmesinde etkili olmuştur. Bu ajanların tek başına veya kombine rejimler halinde kullanımı hem prelinik hem de klinik ortamlarda ümit verici sonuçlar ortaya koymuştur. Bu bölümde, AXL reseptörü hakkında kısaca bilgi verilecek ve AXL'a özgü geliştirilen tedavi stratejilerine değinilerek, kanser tedavisinde değerlendirmeye alınan AXL inhibitörleri tanıtılacaktır.

AXL'in Yapısı ve İşleyişi

AXL, RTK'lerin TAM (TYRO3, AXL ve MERTK) alt ailesinin bir üyesidir. ANXELKTO adı, "kontrol edilemez" anlamına gelen ve Yunanca "anexelekto" kavramından gelmektedir (1). AXL ilk olarak 1988'de kronik miyeloid lösemi hücrelerinden (KML) izole edildi (2). AXL proteininin yapısı; i) iki immüoglobulin benzeri alan ve iki fibronektin III alanı içeren membran dışı bölge; ii) bir transmembran alan; iii) bir hücre içi kinaz alanı olmak üzere üç kısımdan oluşur (3). AXL molekülü, K vitaminine bağımlı protein ailesinden olan GAS6 tarafından aktive edilir. Bu aktivasyonda bir AXL molekülü bir

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bulunmuştur. Bu çalışmadan elde edilen gözlemler, KHDAK hastalarında glesatinib ile ilgili bir faz II çalışmasının başlatılmasını sağlamıştır (78).

SONUÇ

AXL kanserde, hücrenin hayatta kalması, EMT, metastaz ve terapötik direnç mekanizmalarında önemli bir oyuncudur. Son on yılda, TKI'ler, mAb'ler, ADC'ler ve çözünebilir reseptörler dahil olmak üzere bir dizi spesifik AXL hedefli tedavi, klinik gelişime dahil olmuştur. Özellikle, AXL inhibitörleri ön çalışmalarda umut verici sonuçlar ortaya koymuştur. Gün geçtikçe artan araştırma sonuçları, AXL inhibisyonunu potansiyel kanser tedavi stratejilerinden biri yapmaktadır. AXL inhibitörlerinin keşfi ve araştırılması, özellikle ilaç direncindeki etkinliğinin ortaya çıkmasıyla birlikte kanser tedavisi seçeneklerinin iyileştirilmesi açısından kritik önem taşımaktadır. Bu bölümde AXL'in kanserdeki rolüne kısaca değinilmiş ve AXL'i hedef alan tedavi stratejilerinden AXL inhibitörleri tanıtılmıştır.

KAYNAKÇA

1. Antony J, Huang RYJ. AXL-driven EMT state as a targetable conduit in cancer, *Cancer Res.* 2017; 77 (14) 3725–3732.
2. Liu E, Hjelle B, Bishop JM. Transforming genes in chronic myelogenous leukemia. *Proc Natl Acad Sci U S A.* 1988;85(6):1952-6. <https://doi.org/10.1073/pnas.85.6.1952>
3. Di Stasi R, De Rosa L, D'Andrea LD. Therapeutic aspects of the Axl/Gas6 molecular system. *Drug Discov Today.* 2020 Dec;25(12):2130-2148. doi: 10.1016/j.drudis.2020.09.022. Epub 2020 Sep 28. PMID: 33002607.
4. Mudduluru, G. Axl and its mediated Signaling Axis in cancer. Johannes Haybaeck (Ed.), *Mechanisms of molecular carcinogenesis.* Switzerland: Springer International Publishing, 2017; 39-60.
5. Levin PA, Brekken RA, Byers LA, Heymach JV, Gerber DE. Axl receptor axis: a new therapeutic target in lung cancer. *J Thorac Oncol.* 2016;11(8):1357–62. <https://doi.org/10.1016/j.jtho.2016.04.015>.
6. Lemke G, Rothlin CV. Immunobiology of the TAM receptors, *Nat. Rev. Immunol.* 8 (5) (2008) 327–336.
7. Scaltriti M, Elkabets M, Baselga J. Molecular Pathways: AXL, a membrane receptor mediator of resistance to therapy. *Clin Cancer Res.* 2016;22(6):1313–7. <https://doi.org/10.1158/1078-0432.CCR-15-1458>.
8. Gay CM, Balaji K, Byers LA. Giving AXL the axe: targeting AXL in human malignancy. *Br J Cancer.* 2017;116(4):415–23. <https://doi.org/10.1038/bjc.2016.428>.
9. Zhu C, Wei Y, Wei X. AXL receptor tyrosine kinase as a promising anti-cancer approach: functions, molecular mechanisms and clinical applications. *Mol Cancer.* 2019;18(1):153. <https://doi.org/10.1186/s12943-019-1090-3>.
10. Atıcı Y. ve Kara H. (2022) Bölüm 9 “Gas6/Axl Sinyal Yolu ve Kansere İlişkisi”, *Güncel Biyokimya Çalışmaları III*, Editör Doğan YÜCEL, Akademisyen Yayınevi, Ankara

11. Son HY, Jeong HK. Immune Evasion Mechanism and AXL. *Front Oncol*. 2021 Oct 28;11:756225. doi: 10.3389/fonc.2021.756225. PMID: 34778071; PMCID: PMC8581356.
12. Zhang G, Kong X, Wang M, Zhao H, Han S, Hu R, et al. AXL is a marker for epithelial-mesenchymal transition in esophageal squamous cell carcinoma. *Oncol Lett*. 2018;15(2):1900–6. <https://doi.org/10.3892/ol.2017.7443>.
13. Bhalla S, Gerber DE. AXL Inhibitors: Status of Clinical Development. *Curr Oncol Rep*. 2023 May;25(5):521–529. doi: 10.1007/s11912-023-01392-7. Epub 2023 Mar 15. PMID: 36920638.
14. Byers LA, Diao L, Wang J, Saintigny P, Girard L, Peyton M, et al. An epithelial-mesenchymal transition gene signature predicts resistance to EGFR and PI3K inhibitors and identifies Axl as a therapeutic target for overcoming EGFR inhibitor resistance. *Clin Cancer Res*. 2013;19(1):279–90. <https://doi.org/10.1158/1078-0432.CCR-12-1558>.
15. Wu F, Li J, Jang C, Wang J, Xiong J. The role of Axl in drug resistance and epithelial-to-mesenchymal transition of non-small cell lung carcinoma. *Int J Clin Exp Pathol*. 2014 Sep 15;7(10):6653–61. PMID: 25400744; PMCID: PMC4230140.
16. Malvankar C, Kumar D. AXL kinase inhibitors- A prospective model for medicinal chemistry strategies in anticancer drug discovery. *Biochim Biophys Acta Rev Cancer*. 2022 Sep;1877(5):188786. doi: 10.1016/j.bbcan.2022.188786. Epub 2022 Sep 1. PMID: 36058379.
17. Zaman A, Bivona TG. Targeting AXL in NSCLC. *Lung Cancer (Auckl)*. 2021 Aug 10;12:67–79. doi: 10.2147/LCTT.S305484. PMID: 34408519; PMCID: PMC8364399.
18. Lin JZ, Wang ZJ, De W, Zheng M, Xu WZ, Wu HF, et al. Targeting AXL overcomes resistance to docetaxel therapy in advanced prostate cancer. *Oncotarget*. 2017;8(25):41064–77. <https://doi.org/10.18632/oncotarget.17026>.
19. Taniguchi H, Yamada T, Wang R, Tanimura K, Adachi Y, Nishiyama A, et al. AXL confers intrinsic resistance to osimertinib and advances the emergence of tolerant cells. *Nat Commun*. 2019;10(1):259. <https://doi.org/10.1038/s41467-018-08074-0>.
20. Hugo W, Zaretsky JM, Sun L, Song C, Moreno BH, Hu-Lieskovan S, et al. Genomic and transcriptomic features of response to anti-PD-1 therapy in metastatic melanoma. *Cell*. 2016;165(1):35–44. <https://doi.org/10.1016/j.cell.2016.02.065>.
21. Auyez A, Sayan AE, Kriajevska M, Tulchinsky E. AXL Receptor in Cancer Metastasis and Drug Resistance: When Normal Functions Go Askew. *Cancers (Basel)*. 2021 Sep 28;13(19):4864. doi: 10.3390/cancers13194864. PMID: 34638349; PMCID: PMC8507788.
22. Tanaka M, Siemann DW. Gas6/Axl signaling pathway in the tumor immune microenvironment. *Cancers (Basel)*. 2020;12(7):1850. <https://doi.org/10.3390/cancers12071850>.
23. Scott AM, Wolchok JD, Old LJ. Antibody therapy of cancer. *Nat Rev Cancer*. 2012;12(4):278–87. <https://doi.org/10.1038/nrc3236>.
24. Leconet W, Larbouret C, Chardes T, Thomas G, Neiveyans M, Busson M, et al. Pre-clinical validation of AXL receptor as a target for antibody-based pancreatic cancer immunotherapy. *Oncogene*. 2014;33(47):5405–14. <https://doi.org/10.1038/onc.2013.487>.

25. Chang H, An R, Li X, Lang X, Feng J, Lv M. Anti-Axl monoclonal antibodies attenuate the migration of MDA-MB-231 breast cancer cells. *Oncol Lett.* 2021 Nov;22(5):749. doi: 10.3892/ol.2021.13010. Epub 2021 Aug 27.
26. Davra V, Kumar S, Geng K, Calianese D, Mehta D, Gadiyar V, Kasikara C, Lahey KC, Chang YJ, Wichroski M, Gao C, De Lorenzo MS, Kotenko SV, Bergsbaken T, Mishra PK, Gause WC, Quigley M, Spires TE, Birge RB. Axl and Mertk Receptors Cooperate to Promote Breast Cancer Progression by Combined Oncogenic Signaling and Evasion of Host Antitumor Immunity. *Cancer Res.* 2021 Feb 1;81(3):698-712. doi: 10.1158/0008-5472.CAN-20-2066. Epub 2020 Nov 25.
27. Van Renterghem B, Wozniak A, Castro PG, Franken P, Pencheva N, Sciot R, Schöffski P. Enapotamab Vedotin, an AXL-Specific Antibody-Drug Conjugate, Demonstrates Antitumor Efficacy in Patient-Derived Xenograft Models of Soft Tissue Sarcoma. *Int J Mol Sci.* 2022 Jul 6;23(14):7493. doi: 10.3390/ijms23147493.
28. Sharp LL, Chang C, Frey G, et al. Anti-tumor efficacy of BA3011, a novel conditionally active biologic (CAB) anti-AXLADC. *Cancer Res.* 2018;78(13_Supplement):827.
29. Zammarchi F, Havenith KE, Chivers S, Hogg P, Bertelli F, Tyrer P, Janghra N, Reinert HW, Hartley JA, van Berkel PH. Preclinical Development of ADCT-601, a Novel Pyrrollobenzodiazepine Dimer-based Antibody-drug Conjugate Targeting AXL-expressing Cancers. *Mol Cancer Ther.* 2022 Apr 1;21(4):582-593. doi: 10.1158/1535-7163.MCT-21-0715.
30. Mullen MM, Lomonosova E, Toboni MD, Opl A, Cybulla E, Blachut B, Zhao P, Noia H, Wilke D, Rankin EB, Kuroki LM, Hagemann AR, Hagemann IS, McCourt CK, Thaker PH, Mutch DG, Powell MA, Mosammamarast N, Vindigni A, Fuh KC. GAS6/AXL Inhibition Enhances Ovarian Cancer Sensitivity to Chemotherapy and PARP Inhibition through Increased DNA Damage and Enhanced Replication Stress. *Mol Cancer Res.* 2022 Feb;20(2):265-279. doi: 10.1158/1541-7786.MCR-21-0302. Epub 2021 Oct 20.
31. Kariolis, M.S.; Miao, Y.R.; Jones, D.S., 2nd; Kapur, S.; Mathews, I.I.; Giaccia, A.J.; Cochran, J.R. An engineered Axl 'decoy receptor' effectively silences the Gas6-Axl signaling axis. *Nat. Chem. Biol.* 2014, 10, 977–983.
32. Tanaka M, Siemann DW. Therapeutic Targeting of the Gas6/Axl Signaling Pathway in Cancer. *Int J Mol Sci.* 2021 Sep 15;22(18):9953. doi: 10.3390/ijms22189953. PMID: 34576116; PMCID: PMC8469858.
33. Rankin, E.B.; Fuh, K.C.; Castellini, L.; Viswanathan, K.; Finger, E.C.; Diep, A.N.; Lagory, E.L.; Kariolis, M.S.; Chan, A.; Lindgren, D.; et al. Direct regulation of GAS6/AXL signaling by HIF promotes renal metastasis through SRC and MET. *Proc. Natl. Acad. Sci. USA* 2014, 111, 13373–13378.
34. Xiao, Y.; Zhao, H.; Tian, L.; Nolley, R.; Diep, A.N.; Ernst, A.; Fuh, K.C.; Miao, Y.R.; von Eyben, R.; Leppert, J.T.; et al. S100A10 Is a Critical Mediator of GAS6/AXL-Induced Angiogenesis in Renal Cell Carcinoma. *Cancer Res.* 2019, 79, 5758–5768.
35. Zhao Z, Li Y, Liu W, Li X. Engineered IL-7 Receptor Enhances the Therapeutic Effect of AXL-CAR-T Cells on Triple-Negative Breast Cancer. *Biomed Res Int.* 2020 Jan 2;2020:4795171. doi: 10.1155/2020/4795171.
36. C. Wilson, X. Ye, T. Pham, E. Lin, S. Chan, E. McNamara, R.M. Neve, L. Belmont, H. Koeppen, R.L. Yauch, AXL inhibition sensitizes mesenchymal cancer cells to antimetabolic drugs, *Cancer Res.* 74 (20) (2014) 5878–5890.

37. Sheridan C. First Axl inhibitor enters clinical trials. *Nat Biotechnol.* 2013 Sep;31(9):775-6. doi: 10.1038/nbt0913-775a.
38. Quinn JM, Greenwade MM, Palisoul ML, Opara G, Massad K, Guo L, Zhao P, Beck-Noia H, Hagemann IS, Hagemann AR, McCourt CK, Thaker PH, Powell MA, Mutch DG, Fuh KC. Therapeutic Inhibition of the Receptor Tyrosine Kinase AXL Improves Sensitivity to Platinum and Taxane in Ovarian Cancer. *Mol Cancer Ther.* 2019 Feb;18(2):389-398. doi: 10.1158/1535-7163.MCT-18-0537. Epub 2018 Nov 26.
39. Bhalla S, Farjana FJ, Williams JN, et al. Phase 1 dose escalation and expansion study of bemcentinib (BGB324), a first-in-class, selective AXL inhibitor, with docetaxel in patients with previously treated advanced NSCLC. *J Clin Oncol.* 2022;40(16_suppl):9081.
40. Shen Y, Chen X, He J, Liao D, Zu X. Axl inhibitors as novel cancer therapeutic agents. *Life Sci.* 2018 Apr 1;198:99-111. doi: 10.1016/j.lfs.2018.02.033. Epub 2018 Feb 27.
41. A. Mollard, S.L. Warner, L.T. Call, M.L. Wade, J.J. Bearss, A. Verma, S. Sharma, H. Vankayalapati, D.J. Bearss, Design, synthesis, and biological evaluation of a series of novel AXL kinase inhibitors, *ACS Med. Chem. Lett.* 2 (12) (2011) 907–912.
42. Aveic S, Corallo D, Porcù E, Pantile M, Boso D, Zanon C, Viola G, Sidarovich V, Mariotto E, Quattrone A, Basso G, Tonini GP. TP-0903 inhibits neuroblastoma cell growth and enhances the sensitivity to conventional chemotherapy. *Eur J Pharmacol.* 2018 Jan 5;818:435-448. doi: 10.1016/j.ejphar.2017.11.016. Epub 2017 Nov 14.
43. Wang D, Bi L, Ran J, Zhang L, Xiao N, Li X. Gas6/Axl signaling pathway promotes proliferation, migration and invasion and inhibits apoptosis in A549 cells. *Exp Ther Med.* 2021 Nov;22(5):1321. doi: 10.3892/etm.2021.10756. Epub 2021 Sep 20.
44. Terragno M, Vetrova A, Semenov O, Sayan AE, Kriajevskaja M, Tulchinsky E. Mesenchymal-epithelial transition and AXL inhibitor TP-0903 sensitise triple-negative breast cancer cells to the antimalarial compound, artesunate. *Sci Rep.* 2024 Jan 3;14(1):425. doi: 10.1038/s41598-023-50710-3.
45. Kim SH, Choi S, Lee WS. Bevacizumab and anelexleto inhibitor, TP-0903 inhibits TGF- β 1-induced epithelial-mesenchymal transition of colon cancer cells. *Anticancer Drugs.* 2022 Jan 1;33(1):e453-e461. doi: 10.1097/CAD.0000000000001239.
46. Sinha S, Boysen J, Nelson M, Secreto C, Warner SL, Bearss DJ, et al. Targeted Axl inhibition primes chronic lymphocytic leukemia B cells to apoptosis and shows synergistic/additive effects in combination with BTK inhibitors. *Clin Cancer Res.* 2015;21(9):2115
47. Zhang Y, Arner EN, Rizvi A, Toombs JE, Huang H, Warner SL, Foulks JM, Brekken RA. AXL Inhibitor TP-0903 Reduces Metastasis and Therapy Resistance in Pancreatic Cancer. *Mol Cancer Ther.* 2022 Jan;21(1):38-47. doi: 10.1158/1535-7163.MCT-21-0293. Epub 2021 Oct 21.
48. Adjei AA, Melear J, Thompson J, et al. 536MO a phase I, first-in-human, safety, pharmacokinetic, and pharmacokinetic study of oral duberminib (TP-0903) in patients with advanced solid tumours. *Ann Oncol.* 2020;31(suppl_4):S469
49. Mims AS, Huang Y, Eisenmann E, et al. A phase 1b/2 study of TP-0903 and decitabine targeting mutant TP53 and/or complex karyotype in patients with untreated acute myeloid leukemia \geq age 60 years: phase 1b interim results. *J Clin Oncol.* 2022;40(16_suppl):9081.

50. Jeon JY, Buelow DR, Garrison DA, Niu M, Eisenmann ED, Huang KM, Zavorka Thomas ME, Weber RH, Whatcott CJ, Warner SL, Orwick SJ, Carmichael B, Stahl E, Brinton LT, Lapalombella R, Blachly JS, Hertlein E, Byrd JC, Bhatnagar B, Baker SD. TP-0903 is active in models of drug-resistant acute myeloid leukemia. *JCI Insight*. 2020 Dec 3;5(23):e140169. doi: 10.1172/jci.insight.140169.
51. Kostecki KL, Iida M, Wiley AL, Kimani S, Mehall B, Tetreault K, Alexandridis R, Yu M, Hong S, Salgia R, Bruce JY, Birge RB, Harari PM, Wheeler DL. Dual Axl/MerTK inhibitor INCB081776 creates a proinflammatory tumor immune microenvironment and enhances anti-PDL1 efficacy in head and neck cancer. *Head Neck*. 2023 May;45(5):1255-1271. doi: 10.1002/hed.27340. Epub 2023 Mar 20.
52. Rios-Doria J, Favata M, Lasky K, Feldman P, Lo Y, Yang G, Stevens C, Wen X, Sehra S, Katiyar K, Liu K, Wynn R, Harris JJ, Ye M, Spitz S, Wang X, He C, Li YL, Yao W, Covington M, Scherle P, Koblisch H. A Potent and Selective Dual Inhibitor of AXL and MERTK Possesses Both Immunomodulatory and Tumor-Targeted Activity. *Front Oncol*. 2020 Dec 7;10:598477. doi: 10.3389/fonc.2020.598477.
53. Okura N, Nishioka N, Yamada T, Taniguchi H, Tanimura K, Katayama Y, Yoshimura A, Watanabe S, Kikuchi T, Shiotsu S, Kitazaki T, Nishiyama A, Iwasaku M, Kaneko Y, Uchino J, Uehara H, Horinaka M, Sakai T, Tanaka K, Kozaki R, Yano S, Takayama K. ONO-7475, a Novel AXL Inhibitor, Suppresses the Adaptive Resistance to Initial EGFR-TKI Treatment in EGFR-Mutated Non-Small Cell Lung Cancer. *Clin Cancer Res*. 2020 May 1;26(9):2244-2256. doi: 10.1158/1078-0432.CCR-19-2321. Epub 2020 Jan 17.
54. Post SM, Ma H, Malaney P, Zhang X, Aitken MJL, Mak PY, Ruvolo VR, Yasuhiro T, Kozaki R, Chan LE, Ostermann LB, Konopleva M, Carter BZ, DiNardo C, Andreeff MD, Khoury JD, Ruvolo PP. AXL/MERTK inhibitor ONO-7475 potently synergizes with venetoclax and overcomes venetoclax resistance to kill FLT3-ITD acute myeloid leukemia. *Haematologica*. 2022 Jun 1;107(6):1311-1322. doi: 10.3324/haematol.2021.278369. PMID: 34732043; PMCID: PMC9152975.
55. Mahadevan D, Theiss N, Morales C, Stejskal AE, Cooke LS, Zhu M, Kurtzman D, Swart R, Ong E, Qi W. Novel receptor tyrosine kinase targeted combination therapies for imatinib-resistant gastrointestinal stromal tumors (GIST). *Oncotarget*. 2015 Feb 10;6(4):1954-66. doi: 10.18632/oncotarget.3021..
56. Fedorenko IV, Fang B, Koomen JM, Gibney GT, Smalley KS. Amuvatinib has cytotoxic effects against NRAS-mutant melanoma but not BRAF-mutant melanoma. *Melanoma Res*. 2014 Oct;24(5):448-53. doi: 10.1097/CMR.000000000000103. PMID: 24950457; PMCID: PMC4384823.
57. Dantas-Barbosa, C.; Lesluyes, T.; Loarer, F.L.; Chibon, F.; Treilleux, I.; Coindre, J.M.; Meeus, P.; Brahmi, M.; Bally, O.; Ray-Coquard, I.; et al. Expression and role of TYRO3 and AXL as potential therapeutical targets in leiomyosarcoma. *Br. J. Cancer* 2017, 117, 1787–1797.
58. Lin JJ, Choudhury NJ, Yoda S, Zhu VW, Johnson TW, Sakhtemani R, Dagogo-Jack I, Digumarthy SR, Lee C, Do A, Peterson J, Prutisto-Chang K, Malik W, Hubbeling HG, Langenbucher A, Schoenfeld AJ, Falcon CJ, Temel JS, Sequist LV, Yeap BY, Lennerz JK, Shaw AT, Lawrence MS, Ou SI, Hata AN, Drilon A, Gainor JF. Spectrum of Mechanisms of Resistance to Crizotinib and Lorlatinib in ROS1 Fusion-Positive Lung Cancer. *Clin Cancer Res*. 2021 May 15;27(10):2899-2909. doi: 10.1158/1078-0432.CCR-21-0032. Epub 2021 Mar 8.

59. Burbridge, M.F.; Bossard, C.J.; Saunier, C.; Fejes, I.; Bruno, A.; Leonce, S.; Ferry, G.; Da Violante, G.; Bouzom, F.; Cattan, V.; et al. S49076 is a novel kinase inhibitor of MET, AXL, and FGFR with strong preclinical activity alone and in association with bevacizumab. *Mol. Cancer Ther.* 2013, 12, 1749–1762. [CrossRef]
60. Viteri S, Chang G-C, Chiari R, Cho BC, Ciardiello F, Curigliano G, Hida T, Lee DH, Lim WTD, Lin C-C, Martinez A, Murakami H, Natsume I, Nishio M, Paz-Ares L, Soo RA, Cattan V, Gandossi E, Heck H, Park K. 1452P - Combination of the S49076 with gefitinib in NSCLC patients progressing on EGFR-TKI and harboring MET/AXL dysregulation, *Annals of Oncology*, Volume 29, Supplement 8, 2018, Page viii525, ISSN 0923-7534, <https://doi.org/10.1093/annonc/mdy292.074>.
61. Clemenson, C.; Chargari, C.; Liu, W.; Mondini, M.; Ferte, C.; Burbridge, M.F.; Cattan, V.; Jacquet-Bescond, A.; Deutsch, E. The MET/AXL/FGFR Inhibitor S49076 Impairs Aurora B Activity and Improves the Antitumor Efficacy of Radiotherapy. *Mol. Cancer Ther.* 2017, 16, 2107–2119.
62. Keunchil Park, Gee-Chen Chang, Giuseppe Curigliano, Wan-Teck Lim, Ross A. Soo, Miguel A. Molina-Vila, Valérie Cattan, Hélène Darville, Eric Gandossi, Veronika Smutna, Isabelle Sudey, Santiago Viteri, Phase I results of S49076 plus gefitinib in patients with EGFR TKI-resistant non-small cell lung cancer harbouring MET/AXL dysregulation, *Lung Cancer*, Volume 155, 2021, Pages 127-135, ISSN 0169-5002, <https://doi.org/10.1016/j.lungcan.2021.03.012>.
63. van der Mijn, J.C., Broxterman, H.J., Knol, J.C., Piersma, S.R., De Haas, R.R., Dekker, H., Pham, T.V., Van Beusechem, V.W., Halmos, B., Mier, J.W., Jiménez, C.R. and Verheul, H.M.W. (2016), Sunitinib activates Axl signaling in renal cell cancer. *Int. J. Cancer*, 138: 3002-3010. <https://doi.org/10.1002/ijc.30022>
64. Rini BI, Plimack ER, Stus V, Gafanov R, Hawkins R, Nosov D, Pouliot F, Alekseev B, Soulières D, Melichar B, Vynnychenko I, Kryzhanivska A, Bondarenko I, Azevedo SJ, Borchiellini D, Szczylik C, Markus M, McDermott RS, Bedke J, Tartas S, Chang YH, Tamada S, Shou Q, Perini RF, Chen M, Atkins MB, Powles T; KEYNOTE-426 Investigators. Pembrolizumab plus Axitinib versus Sunitinib for Advanced Renal-Cell Carcinoma. *N Engl J Med.* 2019 Mar 21;380(12):1116-1127. doi: 10.1056/NEJMoa1816714. Epub 2019 Feb 16.
65. Zhou L, Liu XD, Sun M, Zhang X, German P, Bai S, Ding Z, Tannir N, Wood CG, Martin SF, Karam JA, Tamboli P, Sircar K, Rao P, Rankin EB, Laird DA, Hoang AG, Walker CL, Giaccia AJ, Jonasch E. Targeting MET and AXL overcomes resistance to sunitinib therapy in renal cell carcinoma. *Oncogene.* 2016 May;35(21):2687-97. doi: 10.1038/onc.2015.343. Epub 2015 Sep 14.
66. Msaouel P, Goswami S, Thall PF, Wang X, Yuan Y, Jonasch E, Gao J, Campbell MT, Shah AY, Corn PG, Tam AL, Ahrar K, Rao P, Sircar K, Cohen L, Basu S, Duan F, Jindal S, Zhang Y, Chen H, Yadav SS, Shazer R, Der-Torossian H, Allison JP, Sharma P, Tannir NM. A phase 1-2 trial of sitravatinib and nivolumab in clear cell renal cell carcinoma following progression on antiangiogenic therapy. *Sci Transl Med.* 2022 Apr 20;14(641):eabm6420. doi: 10.1126/scitranslmed.abm6420. Epub 2022 Apr 20.
67. Martinelli E, Martini G, Cardone C, Troiani T, Liguori G, Vitagliano D, Napolitano S, Morgillo F, Rinaldi B, Melillo RM, Liotti F, Nappi A, Bianco R, Berrino L, Ciuffreda LP, Ciardiello D, Iaffaioli V, Botti G, Ferraiolo F, Ciardiello F. AXL is an oncotarget in human colorectal cancer. *Oncotarget.* 2015 Sep 15;6(27):23281-96. doi: 10.18632/oncotarget.3962.

68. Chia, S.K.; Ellard, S.L.; Mates, M.; Welch, S.; Mihalciou, C.; Miller, W.H., Jr.; Gelmon, K.; Lohrisch, C.; Kumar, V.; Taylor, S.; et al. A phase-I study of lapatinib in combination with foretinib, a c-MET, AXL and vascular endothelial growth factor receptor inhibitor, in human epidermal growth factor receptor 2 (HER-2)-positive metastatic breast cancer. *Breast Cancer Res.* 2017, 19, 54.
69. Kataoka Y, Mukohara T, Tomioka H, Funakoshi Y, Kiyota N, Fujiwara Y, Yashiro M, Hirakawa K, Hirai M, Minami H. Foretinib (GSK1363089), a multi-kinase inhibitor of MET and VEGFRs, inhibits growth of gastric cancer cell lines by blocking inter-receptor tyrosine kinase networks. *Invest New Drugs.* 2012 Aug;30(4):1352-60. doi: 10.1007/s10637-011-9699-0. Epub 2011 Jun 8. PMID: 21655918.
70. Hassan MS, Williams F, Awasthi N, Schwarz MA, Schwarz RE, Li J, von Holzen U. Combination effect of lapatinib with foretinib in HER2 and MET co-activated experimental esophageal adenocarcinoma. *Sci Rep.* 2019 Nov 26;9(1):17608. doi: 10.1038/s41598-019-54129-7.
71. Liu, L.; Greger, J.; Shi, H.; Liu, Y.; Greshock, J.; Annan, R.; Halsey, W.; Sathe, G.M.; Martin, A.M.; Gilmer, T.M. Novel mechanism of lapatinib resistance in HER2-positive breast tumor cells: Activation of AXL. *Cancer Res.* 2009, 69, 6871–6878.
72. Yau TCC, Lencioni R, Sukeepaisarnjaroen W, Chao Y, Yen CJ, Lausoontornsiri W, Chen PJ, Sanpajit T, Camp A, Cox DS, Gagnon RC, Liu Y, Raffensperger KE, Kulkarni DA, Kallender H, Ottesen LH,
73. Yang, P.W.; Liu, Y.C.; Chang, Y.H.; Lin, C.C.; Huang, P.M.; Hua, K.T.; Lee, J.M.; Hsieh, M.S. Cabozantinib (XL184) and R428 (BGB324) Inhibit the Growth of Esophageal Squamous Cell Carcinoma (ESCC). *Front. Oncol.* 2019, 9, 1138.
74. Choueiri, T.K.; Powles, T.; Burotto, M.; Escudier, B.; Bours, M.T.; Zurawski, B.; Oyervides Juarez, V.M.; Hsieh, J.J.; Basso, U.; Shah, A.Y.; et al. Nivolumab plus Cabozantinib versus Sunitinib for Advanced Renal-Cell Carcinoma. *N. Engl. J. Med.* 2021, 384, 829–841.
75. Diane M. Bodenmiller, Julie A. Stewart, Glenn F. Evans, Victoria L. Peek, Jennifer R. Stephens, Xi Lin, Seema Iyer, Beverly L. Falcon, Sudhakar Chintharlapalli, Sau-Chi Betty Yan, Anthony S. Fischl. Characterization of the anti-angiogenic properties of merestinib (LY2801653), an onco-kinase inhibitor [abstract]. In: Proceedings of the American Association for Cancer Research Annual Meeting 2017; 2017 Apr 1-5; Washington, DC. Philadelphia (PA): AACR; *Cancer Res* 2017;77(13 Suppl):Abstract nr 1823. doi:10.1158/1538-7445.AM2017-1823
76. Bruce W. Konicek, Steve M. Bray, Andrew R. Capen, John N. Calley, Kelly M. Credille, Philip J. Ebert, Gary Heady, Bharvin K. Patel, Victoria L. Peek, Jennifer R. Stephens, Suzane L. Um, Melinda D. Willard, Isabella H. Wulur, Yi Zeng, Richard A. Wargren, Sau-Chi Betty Yan. Merestinib (LY2801653), targeting several onco-kinases including NTRK1/2/3, shows potent anti-tumor effect in colorectal cell line- and patient-derived xenograft (PDX) model bearing TPM3-NTRK1 fusion. [abstract]. In: Proceedings of the 107th Annual Meeting of the American Association for Cancer Research; 2016 Apr 16-20; New Orleans, LA. Philadelphia (PA): AACR; *Cancer Res* 2016;76(14 Suppl):Abstract nr 2647.
77. Zhu C, Shi H, Wu M, Wei X. A dual MET/AXL small-molecule inhibitor exerts efficacy against gastric carcinoma through killing cancer cells as well as modulating tumor microenvironment. *MedComm.* 2020; 1: 103–118. <https://doi.org/10.1002/mco2.11>

78. Kollmannsberger C, Hurwitz H, Bazhenova L, Cho BC, Hong D, Park K, Reckamp KL, Sharma S, Der-Torossian H, Christensen JG, Faltaos D, Potvin D, Tassell V, Chao R, Shapiro GI. Phase I Study Evaluating Glesatinib (MGCD265), An Inhibitor of MET and AXL, in Patients with Non-small Cell Lung Cancer and Other Advanced Solid Tumors. *Target Oncol.* 2023 Jan;18(1):105-118. doi: 10.1007/s11523-022-00931-9. Epub 2022 Dec 2.

Bölüm 6

PROSTAT KANSERİ BİYOBELİRTEÇLERİ

Gamze GÖK¹

PROSTAT KANSERİ

Kanser, hücrelerdeki kontrolsüz bölünme ve çoğalma neticesinde oluşur. Çevresel ve genetik unsurlardan etkilenir. Kanserün günümüzde 100'den fazla türü tanımlanmıştır. Bazı kanser türlerine yaklaşım için standartlar oluşturulmasına rağmen, kanserli her olgu özgün değerlendirilmelidir (1).

Erkeklerde ikinci en fazla görülen kanser türü olan prostat kanserinin tüm kanserler içerisinde görülme oranı yaklaşık olarak %15'tir. Prostat kanseri teşhis edildiğinde lokalize ise kürün tam sağlanabilmesi mümkündür (2). Kaynağını prostatik bezden ya da kanal asinilerinden alan prostat kanseri en çok mortaliteye sebep olan kanserler içinde ilk beşte yer almaktadır (3). Prostat kanserine yaklaşım gelişme göstermektedir. Prostat kanseri özellikle 50 yaş üstü erkeklerin sağlıkları için ciddi risk içerir. Bu hastalığın klinik çeşitlilikleri farklı tedavi seçeneklerini beraberinde getirir (4).

Coğrafik ve etnik nedenlerden dolayı prostat kanserinin görülme sıklığı bölgeler arasında değişmektedir (5). Prostat kanseri görülme oranları Kuzey Amerika, Avustralya ve Avrupada yüksek iken; Asyada düşüktür (6). Prostat kanserinin Türkiye'de görülme sıklığı Amerika'dan daha düşük olmasına rağmen artış görülmektedir. Bu artış; ülkemizde yaşam tarzında meydana gelen değişiklikler, nüfusun yaşlanması ve prostat kanseri için yapılan taramaların yaygınlaşması nedenleri ile olabilir (7).

Aile öyküsü, ileri yaş, genetik faktörler ve ırk prostat kanseri risk faktörlerindedir. Obezite, diyet, yeterli fiziksel hareketin olmaması, kan glukozunda yükseklik, enfeksiyonlar ve enflamasyon prostat kanseri ile pozitif ilişki gösteren etmenlerdir (8).

Prostat kanseri genellikle yavaş ilerler. Periferik zonda oluşan odaklar küçüktür bu nedenle prostat kanserleri erken evrede genellikle semptom göstermezler (4).

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TMPRSS2-ERG

2005 yılında, prostat kanserinde v-ets erythroblastosis virus E26 onkogen homologu (ERG) mRNA'nın aşırı eksprese olduğu bildirilmiştir (32). ERG'yi aktive eden etken transmembran proteaz serin 2 (TMPRSS2)'dir. Androjenler, TMPRSS2 vasıtasıyla ERG'nin ekspresyonunu indükler. Bu durumun sonucunda protoonkogen ERG aşırı eksprese olur (33). ERG in vitro ve in vivo prostat kanserinde hem kanserin onkogenezi hem de metastazını uyararak başlatır (34). TMPRSS2-ERG füzyonu etnik köken ile ilişkilidir. Etnik grupları farklı olan bireyler arasında %7 ile %83 aralığında farklılık mevcuttur (35). Prostat kanseri ile teşhis edilen Kafkas hastaların yaklaşık yarısında TMPRSS2-ERG füzyonu vardır (13).

SelectMDx

SelectMDx testi, idrar prostat kanseri belirteçlerinden biri olarak geliştirilmiştir. DLX1 geni, HOXC6 geni ve TDRD1 geninin kombinasyonu, Gleason ≥ 7 olan yüksek dereceli prostat kanserlerinin doğru tespit edilmesinde yüksek doğruluk içermektedir. Ticari kiti mevcuttur (13).

SONUÇ

Prostat kanserinin tanısı için yeni biyobelirteçlerin ortaya çıkarılması, biyobelirteçlerin ölçümlerinin hassas, doğru, ulaşılabilir olması tedavi oranlarında iyileştirilme sağlayabilir. Yalnızca prostat kanserinde artış gösterip, diğer benign hastalıklarda ortamda bulunmayan ve böylece hastaları gereksinim dışı biyopsilerden kurtaran biyobelirteçlere ihtiyaç devam etmektedir.

KAYNAKÇA

1. Baykara O. Kanser Tedavisinde Güncel Yaklaşımlar. Balıkesir Sağlık Bilimleri Dergisi. 2016;5(3):154-65.
2. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. International journal of cancer. 2015;136(5):E359-E86.
3. Akbiyik I. Yaşlı kanser hastalarında kemoterapi öncesi kırılabilirlik ve diğer klinik etkenlerin erken dönem kemoterapi toksisitesi ile ilişkisi.
4. Yencilek F, Koca O, Kuru M. Prostat kanserinde tanı. Nucl Med Semin. 2018;4(3):163-73.
5. van den Ouden D, Kranse R, Hop WC, van der Kwast TH, Schröder FH. Microvascular invasion in prostate cancer: prognostic significance in patients treated by radical prostatectomy for clinically localized carcinoma. Urologia internationalis. 1998;60(1):17-24.

6. Kumar V, Majumder P. Prostate gland: structure, functions and regulation. *International urology and nephrology*. 1995;27:231-43.
7. KOÇAK T, Nilüfer A. Prostat Kanseri Etiyoloji ve Tedavisinde Beslenmenin Rolü. *Gümüşhane Üniversitesi Sağlık Bilimleri Dergisi*. 2022;11(3):1247-56.
8. Wolk A. Diet, lifestyle and risk of prostate cancer. *Acta Oncologica*. 2005;44(3):277-81.
9. TOSUNÖZ İK, DOĞAN SD. ERKEKLERİN PROSTAT KANSERİ TARAMALARINA İLİŞKİN TUTUMLARI VE BİLGİ DÜZEYLERİ. *Izmir Democracy University Health Sciences Journal*.5(3):651-63.
10. Bayçelebi G, AYDIN F, Gökosmanoğlu F, Tat TS, VARIM C. Trabzon'da kanser tarama testleri farkındalığı. *Journal of Human Rhythm*. 2015;1(3):90-4.
11. Musalli ZF, Alobaid MM, Aljahani AM, Alqahtani MA, Alshehri SS, Altulaihi BA, Altulaihi B. Knowledge, attitude, and practice toward prostate cancer and its screening methods among primary care patients in King Abdulaziz Medical City, Riyadh, Saudi Arabia. *Cureus*. 2021;13(4).
12. Duffy M. Clinical uses of tumor markers: a critical review. *Critical reviews in clinical laboratory sciences*. 2001;38(3):225-62.
13. Bolla M, van Poppel H. *Management of prostate cancer*: Springer; 2017.
14. Wang M, Valenzuela L, Murphy G, Chu T. Purification of a human prostate specific antigen. *Investigative urology*. 1979;17(2):159-63.
15. Papsidero LD, Wang MC, Valenzuela LA, Murphy GP, Chu TM. A prostate antigen in sera of prostatic cancer patients. *Cancer research*. 1980;40(7):2428-32.
16. Lilja H, Abrahamsson PA. Three predominant proteins secreted by the human prostate gland. *The Prostate*. 1988;12(1):29-38.
17. Stamey TA, Yang N, Hay AR, McNeal JE, Freiha FS, Redwine E. Prostate-specific antigen as a serum marker for adenocarcinoma of the prostate. *New England Journal of Medicine*. 1987;317(15):909-16.
18. Ulmert D, Becker C, Nilsson J-A, Piironen T, Bjork T, Hugosson J, et al. Reproducibility and accuracy of measurements of free and total prostate-specific antigen in serum vs plasma after long-term storage at -20 C. *Clinical chemistry*. 2006;52(2):235-9.
19. Loeb S. Guideline of guidelines: prostate cancer screening. *BJU international*. 2014;114(3):323-5.
20. Bradford TJ, Tomlins SA, Wang X, Chinnaiyan AM, editors. *Molecular markers of prostate cancer*. *Urologic Oncology: Seminars and Original Investigations*; 2006: Elsevier.
21. Borer JG, Sherman J, Solomon MC, Plawker MW, Macchia RJ. Age specific prostate specific antigen reference ranges: population specific. *The Journal of urology*. 1998;159(2):444-8.
22. Catalona WJ, Partin AW, Slawin KM, Brawer MK, Flanigan RC, Patel A, et al. Use of the percentage of free prostate-specific antigen to enhance differentiation of prostate cancer from benign prostatic disease: a prospective multicenter clinical trial. *Jama*. 1998;279(19):1542-7.
23. He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nature reviews genetics*. 2004;5(7):522-31.
24. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *cell*. 2004;116(2):281-97.

25. Miller MC, Doyle GV, Terstappen LW. Significance of circulating tumor cells detected by the CellSearch system in patients with metastatic breast colorectal and prostate cancer. *Journal of oncology*. 2010;2010.
26. Kulaç İ. Sıvı Biyopsi: Dolaşımdaki Tümör Hücreleri Kavramı ve Prostat Kanseri Hastalarının Takip/Tedavisindeki Önemi. *Üroonkoloji Bülteni*. 2014;13(4).
27. Hayes DF, Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Miller MC, et al. Circulating tumor cells at each follow-up time point during therapy of metastatic breast cancer patients predict progression-free and overall survival. *Clinical Cancer Research*. 2006;12(14):4218-24.
28. Ma X, Xiao Z, Li X, Wang F, Zhang J, Zhou R, et al. Prognostic role of circulating tumor cells and disseminated tumor cells in patients with prostate cancer: a systematic review and meta-analysis. *Tumor Biology*. 2014;35:5551-60.
29. Perk H, Ergün O. Prostat kanser teşhisinde PCA3.
30. Marks LS, Bostwick DG. Prostate cancer specificity of PCA3 gene testing: examples from clinical practice. *Reviews in urology*. 2008;10(3):175.
31. Loeb S, Bruinsma SM, Nicholson J, Briganti A, Pickles T, Kakehi Y, et al. Active surveillance for prostate cancer: a systematic review of clinicopathologic variables and biomarkers for risk stratification. *European urology*. 2015;67(4):619-26.
32. Petrovics G, Liu A, Shaheduzzaman S, Furasato B, Sun C, Chen Y, et al. Frequent overexpression of ETS-related gene-1 (ERG1) in prostate cancer transcriptome. *Oncogene*. 2005;24(23):3847-52.
33. Perner S, Demichelis F, Beroukheim R, Schmidt FH, Mosquera J-M, Setlur S, et al. TMPRSS2: ERG fusion-associated deletions provide insight into the heterogeneity of prostate cancer. *Cancer research*. 2006;66(17):8337-41.
34. Zong Y, Xin L, Goldstein AS, Lawson DA, Teitell MA, Witte ON. ETS family transcription factors collaborate with alternative signaling pathways to induce carcinoma from adult murine prostate cells. *Proceedings of the National Academy of Sciences*. 2009;106(30):12465-70.
35. Zhou CK, Young D, Yeboah ED, Coburn SB, Tettey Y, Biritwum RB, et al. TMPRSS2: ERG gene fusions in prostate cancer of West African men and a meta-analysis of racial differences. *American journal of epidemiology*. 2017;186(12):1352-61.

Bölüm 7

OTOİMMÜN TİROİD HASTALIKLARI VE TİROİD FONKSİYON TESTLERİ

Yavuz ELBAŞ¹
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GİRİŞ

Tiroid bezi embriyonik büyüme, organ gelişimi, nörolojik gelişim, enerji metabolizması, üreme, termoregülasyon, glukoz alımının düzenlenmesi, redoks homeostazisi ve iyodür alımı gibi fizyolojik süreçlerde rol oynayan endokrin bir organdır. Bu görevleri tiroksin (T4) ve triiyodotironin (T3) adlı tiroid hormonlarını (TH) üreterek gerçekleştirir. (1,2).

TH tiroid bezinde üretilen ve iyot içeren özel bileşiklerdir. T4 bir prohormondur. T3 ise dolaşımda aktif hormon olarak bulunmaktadır. T4 dolaşımda nispeten daha yüksek konsantrasyonda bulunmaktadır. TH'ler vücuttaki çoğu dokuya etki eder ve T3 üretiminin büyük kısmı dokularda T4'ün enzimatik deiyodinasyonu yoluyla gerçekleşir. Ayrıca TH'lerin üretimi TSH tarafından düzenlenir (3).

Hashimoto tiroiditi ve Graves hastalığı olarak adlandırılan otoimmün tiroid hastalıkları bağışıklık sisteminin tiroid bezine saldırısı ve anormal lenfosit aktivitesi ile karakterize bir grup hastalıktır. Tiroid antijenlerine karşı reaktif T ve B hücrelerinin tiroide infiltrasyonu, tiroid otoantikörlerinin üretimi ve anormal tiroid fonksiyonu ile oluşmaktadır. Otoimmün tiroid hastalıkları heterojen bir hastalık grubudur. Hastalar subklinik biyokimyasal anormalliklerden şiddetli hipertiroidizme veya şiddetli hipotiroidizme kadar geniş bir yelpazede yer alabilmektedir. (4,5,6,7).

T4, başta tiroid bağlayıcı proteine (TBG) (%60-75) olmak üzere, transtiretin (TTR/TBPA) (%15-30) ve albümin (~%10) olmak üzere %99,97 oranında plazma proteinlerine bağlı olarak dolaşır. T3'ün ise, yaklaşık %99,7'si TBG'ye bağlanır. Total tiroid hormonlarının (tT4 ve tT3), toplam (serbest + proteine

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Gebelerde %5-10 oranında görülen postpartum tiroiditin olasılığını tahmin etmek için, hamileliğin erken döneminde Anti-TPO testi kullanılabilir. Anti-TPO pozitif olan gebelerin yaklaşık %50'sinde tiroidit gelişir. Postpartum tiroidit genellikle geçicidir. Bu hastaların %67'sinde klinik semptomlar görülürken, %33'ünde TSH anormal olsa bile semptomlar subklinik kalır. Anti-TPO testi infertilitenin değerlendirilmesinde de yararlı olabilir. Çünkü yüksek Anti-TPO düzeyleri düşük yapma riskinin artmasıyla ve başarısız in vitro fertilizasyon tedavisiyle ilişkilendirilmiştir (8,59).

KAYNAKÇA

1. Marelli F, Rurale G., Persani L. From endoderm to progenitors: an update on the early steps of thyroid morphogenesis in the zebrafish. *Front Endocrinol.* 2021;12. doi:10.3389/fendo.2021.664557
2. Sulaiman A., Luaibi N., Qassim H. Effects of silver nanoparticles on thyroid gland structure and function in female rats. *Asian J Pharm Clin Res.* 2018;11(11): 509. doi:10.22159/ajpcr.2018.v11i11.29383
3. Analytical Science Advances. *Analysis of free, unbound thyroid hormones by liquid chromatography tandem mass spectrometry: A mini review of the medical rationale and analytical methods 2023.* (31/03/2024 tarihinde <https://chemistry-europe.onlinelibrary.wiley.com/doi/epdf/10.1002/ansa.202200067> adresinden ulaşılmıştır).
4. Califaretti E., Dall'armellina S., Rovera G. The role of PET/CT in thyroid autoimmune diseases. *Q J Nucl Med Mol Imaging.* 2022;66(3): 218–228. doi.org/10.23736/S1824-4785.22.03464-1
5. Liu Y., Liu X., Wu N. *Int G Jen Med.* 2023;16: 2355–2363. doi.org/10.2147/IJGM.S410640
6. Yoo W. and Chung H. *Endocrinol Metab.* 2016;31(3): 379. doi.org/10.3803/enm.2016.31.3.379
7. Ferrari S., Ragusa F., Elia G. *Front Pharmacol.* 2021;12. doi.org/10.3389/fphar.2021.750380
8. Spencer C-A, Feingold K-R., Anawalt B. Assay of Thyroid Hormones and Related Substances. Endotext [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000.2017 Feb 20. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK279113/>
9. Kapoor N., Kurian M. Interpretation of thyroid function tests. *Current Medical Issues.* 2018;16(2): 34. doi.org/10.4103/cmi.cmi_17_18
10. Gulnara Dashdamirova. R-A., Rahimova Samira Baghirova., Ulviya Azizova. Pathogenic Mechanisms of Autoimmune Thyroid Disease. *Int J Med Sci.* 2022;6(2): 26-33. doi.org/10.51505/ijmshr.2022.6203
11. Kumata S., Hashimoto Y., Okada H. Pulmonary hypertension associated with hyperthyroidism: a case report. *Cureus.* 2022. doi.org/10.7759/cureus.30567
12. Salah R., Kacem F., Soomauro S. Autoimmune thyroiditis associated with autoimmune diseases. *Electron J Gen Med.* 2022;19(6): 409. doi.org/10.29333/ejgm/12399
13. Albadrani A. Altobje M-A. Conducting Hormonal, Biochemical and Serological Tests in Autoimmune Thyroid Patients. *South Asian J. Res. Microbiol.* 2022;12(4): 6–14. doi.org/10.9734/sajrm/2022/v12i430281

14. Vurgun E., Baltan E., Serin H. Tiroid Hormonları Referans Aralıkları Reference Intervals of Thyroid Hormones Tiroid Hormonları Referans Aralıklarının Belirlenmesi: Merkezi Laboratuvar Verilerinden Determination of Reference Intervals of Thyroid Hormones: from a Central Laboratory Data. *Turkish Journal of Biochemistry*. 2022;20(1): 29–36.
15. Lewandowski K., Gaşior-Perczak D., Kowalska A. Prevalence of macroprolactinaemia in regularly menstruating women with non-toxic goitre or autoimmune thyroid disease. *Thyroid Research*. 2012;5(1). doi.org/10.1186/1756-6614-5-20
16. Fu J., Wu A., Wang X. Concurrent graves' disease and tsh secreting pituitary adenoma presenting suppressed thyrotropin levels: a case report and review of the literature. *Front. Endocrinol*. 2020;11. doi.org/10.3389/fendo.2020.00523
17. Oueslati I., Salhi S., Yazidi A case of hashimoto's thyroiditis following graves' disease. *Clin. Case Rep*. 2022;10(10). doi.org/10.1002/ccr3.6466
18. Marcocci C., Leo M., Altea M. Oxidative stress in graves disease. *Eur. Thyroid J*. 2012;1(2): 80-87. doi.org/10.1159/000337976
19. Nguyen J. and Joseph D. Graves' disease in an adolescent presenting with increased intracranial pressure and bilateral papilledema. *JCEM Case Rep*. 2022. doi.org/10.1530/edm-22-0240
20. Gursu T., Cevik H., Desteli G-A. Diagnostic value of shear wave velocity in polycystic ovarian syndrome. *J Ultrason*. 2021. doi:10.15557/JoU.2021.0047
21. Taylor P., Zhang L., Lee R. New insights into the pathogenesis and nonsurgical management of graves orbitopathy. *Nat Rev Endocrinol*. 2019;16(2): 104-116. doi.org/10.1038/s41574-019-0305-4
22. Kotwal N., Singh Y., Menon A. Thymic hyperplasia in graves disease. *Indian J Endocrinol Metab*. 2013;17(3): 521. doi.org/10.4103/2230-8210.111676
23. Kumata S., Hashimoto Y., Okada H. Pulmonary hypertension associated with hyperthyroidism: a case report. *Cureus*. 2022. doi.org/10.7759/cureus.30567
24. Elenius H., Cesa M., Suárez C. Thyrotoxic periodic paralysis causing back pain and leg weakness: an unusual presentation of hyperthyroidism. *Case Rep Endocrinol*. 2021;1-4. doi.org/10.1155/2021/6622658
25. TEMD. Tiroid Hastalıkları Tanı ve Tedavi Kılavuzu,2023. [Online] https://file.temd.org.tr/Uploads/publications/guides/documents/202305120904-2023tbl_kilavuz.pdf [Accessed: 20th March 2024]
26. Macovei M., Azis Ü., Gheorghie A., A systematic review of euthyroid graves' disease (review). *Exp Ther Med*. 2021;22(5). doi.org/10.3892/etm.2021.10781
27. Shahbaz A., Aziz K., Umair M. Graves' disease presenting as painful goiter: a case report and review of the literature. *Cureus*. 2018. doi.org/10.7759/cureus.2765
28. Wang S., Wang C., Tien K. Thyroid-stimulating hormone receptor antibodies during follow-up as remission markers in childhood-onset graves' disease treated with antithyroid drugs. *The Kaohsiung J Med Sci*. 2019;36(4): 281-286. doi.org/10.1002/kjm2.12167
29. Gallo D., Piantanida E., Gallazzi M. Immunological Drivers in Graves' Disease: NK Cells as a Master Switcher. *Front Endocrinol*. 2020;11: 406. doi.org/10.3389/fendo.2020.00406
30. Hiruma M., Watanabe N., Mitsumatsu T. Clinical features of moyamoya disease with graves' disease: a retrospective study of 394,422 patients with thyroid disease. *Endocr J*. 2023;70(2): 141-148. doi.org/10.1507/endocrj.ej22-0319

31. Azeez T., Egbu A. Clinical profiles of thyroid dermopathy: a dermato-endocrinology minireview. *Endocrinol Metab Int J.* 2020;8(6): 125-127. doi.org/10.15406/emij.2020.08.00294
32. Kaewdech A., Aiempanakit K., Sangmala S. Elephantiasic pretibial myxedema: a rare manifestation in graves' disease. *J Health Sci Med Res.* 2019. doi.org/10.31584/jhs-mr.201942
33. Piltcher-da-Silva R., Morillos M., Bertão S. Severe cholestatic syndrome secondary to graves disease. *Clin.Biomed. Res.* 2019;39(1): 101-103. doi.org/10.4322/2357-9730.87476
34. Lee C., Chen S., Yang Y. Association between graves' disease and risk of incident systemic lupus erythematosus: a nationwide population-based cohort study. *Int J Rheu Dis.* 2020;24(2): 240-245. doi.org/10.1111/1756-185x.14027
35. TEMD. Tiroid Hastalıkları Tanı ve Tedavi Kılavuzu,2020. [Online] https://file.temd.org.tr/Uploads/publications/guides/documents/20200929134733-2020tbl_kilavuzf527c34496.pdf?a=1 [Accessed: 20th March 2024]
36. Kızıllkan M., Kanbur N., Akgül S. An adolescent boy with comorbid anorexia nervosa and hashimoto thyroiditis. *J Clin Res Ped Endocrinol.* 2016;8(1): 92-95. doi.org/10.4274/jcrpe.2297
37. Dong L., Sun X., Cheng X. Hashimoto's thyroiditis and papillary carcinoma in an adolescent girl: a case report. *Mol Clin Oncol.* 2016;5(1): 129-131. doi.org/10.3892/mco.2016.895
38. Klubo-Gwiedzinska J., Wartofsky L. Hashimoto thyroiditis: an evidence-based guide: etiology, diagnosis and treatment. *Pol Arch Int Med.* 2022. doi.org/10.20452/pamw.16222
39. Casler K. Laboratory Policy and Regulations. Gawlik K. (Ed.) *Laboratory Screening and Diagnostic Evaluation: An Evidence Based Approach* içinde. New York: Springer Publishing Company; 2022. p. 25-30
40. Koulouri O., Gurnell M. How to interpret thyroid function tests. *Clin Med.* 2013;13(3): 282-286. doi.org/10.7861/clinmedicine.13-3-282
41. Thienpont L., Uytfanghe K., Grande L. Harmonization of serum thyroid-stimulating hormone measurements paves the way for the adoption of a more uniform reference interval. *Clin Chem.* 2017;63(7): 1248-1260. doi.org/10.1373/clinchem.2016.269456
42. Peters C., Trotsenburg A., Schoenmakers N. Diagnosis of endocrine disease: congenital hypothyroidism: update and perspectives. *Acta Endocrinol.* 2018;179(6): 297-317. doi.org/10.1530/eje-18-0383
43. Abdi H., Davoodi S., Gharibzadeh S. Association between thyroid function and body mass index: a 10-year follow-up. *Ann Nutr Metab.* 2017;70(4): 338-345. doi.org/10.1159/000477497
44. Hoermann R., Eckl W., Hoermann C. Complex relationship between free thyroxine and tsh in the regulation of thyroid function. *Acta Endocrinol.* 2010;162(6): 1123-1129. doi.org/10.1530/eje-10-0106
45. Shemesh R., Simon G., Zloto O. The role of thyroid antibodies in thyroid eye disease. *Int Ophthalmol Clin.* 2023;63(3): 225-231. doi.org/10.1097/iio.0000000000000451
46. Wanjia X., Jiajun Z. TSH in the Upper Limits Of The Normal Range Is Associated With An Adverse Lipid Profile in Euthyroid Non-Diabetics With Newly Diagnosed Asymptomatic Coronary Heart Disease. *Heart.* 2012;98: 161-162. doi.org/10.1136/heartjnl-2012-302920j.9

47. Meng F, Jonklaas J, Leow M. Interconversion of plasma free thyroxine values from assay platforms with different reference intervals using linear transformation methods. *Biology*. 2021;10(1): 45. doi.org/10.3390/biology10010045
48. Padoan A., Clerico A., Zaninotto M. Percentile transformation and recalibration functions allow harmonization of thyroid-stimulating hormone (tsh) immunoassay results. *Clin. Chem Lab Med*. 2020;58(10): 1663-1672. doi.org/10.1515/cclm-2019-1167
49. Grande L., Uytfanghe K., Reynders D. Standardization of free thyroxine measurements allows the adoption of a more uniform reference interval. *Clin Chem*. 2017;63(10): 1642-1652. doi.org/10.1373/clinchem.2017.274407
50. Marras, V., Casini, M., Pilia, S. Thyroid function in obese children and adolescents. *Horm Res Paediatr*. 2010;73(3): 193-197. doi.org/10.1159/000284361
51. Wolters B., Lass N., Reinehr T. Tsh and free triiodothyronine concentrations are associated with weight loss in a lifestyle intervention and weight regain afterwards in obese children. *Acta Endocrinol*. 2013;168(3): 323-329. doi.org/10.1530/eje-12-0981
52. Torun E., Özgen İ., Gökçe S. Thyroid hormone levels in obese children and adolescents with non-alcoholic fatty liver disease. *J Clin Res Pediatr Endocrinol*. 2014;6(1): 34-39. doi.org/10.4274/jcrpe.1155
53. Chu C., Lam H., Lee J. Hyperthyroidism-associated insulin resistance is not mediated by adiponectin levels. *J Thyroid Res*. 2011;1-5. doi.org/10.4061/2011/194721
54. Singh S., Lamsal M., Baral N. Correlation of iodine content of mother's milk and urine with their child's tsh level. *Asian J Med Sci*. 2015;7(1): 40-48. doi.org/10.3126/ajms.v7i1.12577
55. Clerico A., Trenti T., Aloe R. A multicenter study for the evaluation of the reference interval for tsh in Italy. *Clin Chem Lab Med*. 2018;57(2): 259-267. doi.org/10.1515/cclm-2018-0541
56. Witte T., Völzke H., Lerch M. Association between serum thyroid-stimulating hormone levels and visceral adipose tissue: a population-based study in northeast germany. *Eur Thyroid J*. 2016;6(1): 12-19. doi.org/10.1159/000450977
57. Miao Wang, Jing Li, Youyuan Huang. Analytical validation of the LiCA® high-sensitivity human thyroid stimulating hormone assay. *Clin Biochem*. 2022;101: 42-49. doi:10.1016/j.clinbiochem.2021.11.018
58. Bikle D-D. The Free Hormone Hypothesis: When, Why, and How to Measure the Free Hormone Levels to Assess Vitamin D, Thyroid, Sex Hormone, and Cortisol Status. *JBMR Plus*. 2020;11(2); 5. doi.org/10.1002/jbm4.10418
59. Carvalho G-A-D., Perez C-L-S., Ward L-S. The clinical use of thyroid function tests. *Arq Bras Endocrinol Metab*. 2013;57(3); 193-204. doi.org/10.1590/s0004-27302013000300005

Bölüm 8

PARAOKSONAZ ENZİM AİLESİ ENZİMLERİNDEN HOMOSİSTEİN TİYOLAKTONAZ, PARAOKSONAZ VE ARİL ESTERAZ AKTİVİTELERİNE GENEL BİR BAKIŞ

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GİRİŞ

Homosistein sülfür içeren doğal vaskülotoksik ve trombojenik bir amino asittir. Yapılan çalışmalar, yüksek plazma homosistein ve homosistein metabolizması sonucu oluşan homosistein tiyolakton seviyelerinin doğrudan toksisite ve vasküler endotel hasarına neden olabileceğini göstermektedir (1). Hiperhomosisteinemi homosisteinilasyon ve tiyolasyon yoluyla reaktif oksijen türlerinin (ROS) birikmesine ve proteinlerin posttranslasyonel modifikasyonlarına yol açarak zararlı etkilere sebep olur (2). Hatalı posttranslasyonel değişikliklere uğrayan bu toksik proteinler apoptoza, kazanılmış immün cevaba (LDL oksidasyonuna ve buna bağlı makrofaj cevabındaki artışa) ve tromboza sebep olur (3). Yakın dönemdeki çalışmalarda ise homosistein tiyolaktonun özellikle kromatin organizasyonu, tek karbon metabolizması ve lipit metabolizmasıyla ilgili gen ekspresyonunu da indüklediği gösterilmiştir (4). İnsan paraoksonaz enzimi (PON-1) ise paraoksonaz (PON), homosistein tiyolaktonaz (HTLaz) ve arilesteraz (ARE) enzim aktiviteleri sayesinde hatalı posttranslasyonel modifikasyonlara uğramış zararlı proteinleri inaktive eder. Bu etkiler aracılığıyla PON-1 enziminin vasküloprotektif, antienflamatuvar ve antioksidan özellikleri ortaya çıkar. Bu

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edilmesi, homosistein düzeylerinin düzenlenmesi ve oksidatif stresin azaltılması gibi birçok biyolojik süreçte kritik bir rol oynar.

KAYNAKÇA

1. Austin RC, Lentz SR, Werstuck GH. Role of hyperhomocysteinemia in endothelial dysfunction and atherothrombotic disease. *Cell Death Differ.* 2004;11(Suppl 1):S56–S64. doi:10.1038/sj.cdd.4401451
2. Lehotský J, Tothová B, Kovalská M, Dobrota D, Beňová A, Kalenská D, Kaplán P. Role of homocysteine in the ischemic stroke and development of ischemic tolerance. *Front Neurosci.* 2016;10:538. doi:10.3389/fnins.2016.00538
3. Jakubowski H. The pathophysiological hypothesis of homocysteine thiolactone-mediated vascular disease. *J Physiol Pharmacol.* 2008;59(Suppl 9):155–167.
4. Gurda D, Handschuh L, Kotkowiak W, Jakubowski H. Homocysteine thiolactone and N-homocysteinylated protein induce pro-atherogenic changes in gene expression in human vascular endothelial cells. *Amino Acids.* 2015;47(7):1319–1339. doi:10.1007/s00726-015-1956-7
5. Braekke K, Ueland PM, Harsem NK, Karlsen A, Blomhoff R, Staff AC. Homocysteine, cysteine, and related metabolites in maternal and fetal plasma in preeclampsia. *Pediatr Res.* 2007;62(3):319–324. doi: 10.1203/PDR.0b013e318123fba2
6. Jakubowski H. Homocysteine thiolactone: metabolic origin and protein homocysteinylated in humans. *J Nutr.* 2000;130(2S Suppl):377S–381S. doi:10.1093/jn/130.2.377S
7. Jakubowski H, Zhang L, Bardeguet A, Aviv A. Homocysteine thiolactone and protein homocysteinylated in human endothelial cells: implications for atherosclerosis. *Circ Res.* 2000;87(1):45–51. doi:10.1161/01.res.87.1.45
8. Jakubowski H. Metabolism of homocysteine thiolactone in human cell cultures. Possible mechanism for pathological consequences of elevated homocysteine levels. *J Biol Chem.* 1997;272(3):1935–1942.
9. Yilmaz N. Relationship between paraoxonase and homocysteine: crossroads of oxidative diseases. *Arch Med Sci.* 2012;8(1):138–153. doi:10.5114/aoms.2012.27294
10. Undas A, Jankowski M, Twardowska M, Padjas A, Jakubowski H, Szczeklik A. Antibodies to N-Homocysteinylated Albumin as a Marker for Early-Onset Coronary Artery Disease in Men. *Thromb Haemost.* 2005;93(2):346–50.
11. Refsum H, Ueland PM. Homocysteine and homocysteine thiolactone in human health and disease. *J Inherit Metab Dis.* 1985;8(S1):53–61.
12. Mazur A. An enzyme in animal tissues capable of hydrolysing the phosphorus-fluorine bond of alkyl fluorophosphates. *J Biol Chem.* 1946;164:271–289.
13. Aldridge WN. Serum Esterases. I. Two Types of Esterase (a and B) Hydrolysing P-Nitrophenyl Acetate, Propionate and Butyrate, and a Method for Their Determination. *Biochem J.* 1953;53(1):110–7.
14. Uriel J. [Characterization of Cholinesterase and Other Carboxylic Esterases after Electrophoresis and Immunoelectrophoresis on Agar. I. Application to the Study of Esterases of Normal Human Serum]. *Ann Inst Pasteur (Paris).* 1961;101:104–19.
15. Levy D, Reichert CO, Bydlowski SP. Paraoxonases Activities and Polymorphisms in Elderly and Old-Age Diseases: An Overview. *Antioxidants.* 2019;8(5).

16. Li WF, Costa LG, Furlong CE. Serum Paraoxonase Status: A Major Factor in Determining Resistance to Organophosphates. *J Toxicol Environ Health*. 1993;40(2-3):337-46.
17. Rajkovic MG, Rumora L, Barisic K. The paraoxonase 1, 2 and 3 in humans. *Biochem Med*. 2011;21(2):122–130. doi:10.11613/bm.2011.020
18. Vitarius JA, Sultatos LG. The role of calcium in the hydrolysis of the organophosphate paraoxon by human serum A-esterase. *Life Sci*. 1995;56(2):125–134. doi:10.1016/0024-3205(94)00422-o
19. Pasdar A, Adams HR, Cumming A, Cheung J, Whalley L, St Clair D, MacLeod MJ. Paraoxonase Gene Polymorphisms and Haplotype Analysis in a Stroke Population. *BMC Med Genet*. 2006;7(28):1-6.
20. Mackness M and Sozmen EY. A critical review on human serum paraoxonase-1 in the literature: truths and misconception. *Turk J Biochem* 2021;46(1):3-8
21. Blatter Garin MC, James RW, Dussoix P, et al. Paraoxonase polymorphism Met-Leu54 is associated with modified serum concentrations of the enzyme. A possible link between the paraoxonase gene and increased risk of cardiovascular disease in diabetes. *J Clin Invest*. 1997;99(1):62–66.
22. Dounousi E, Bouba I, Spoto B, Pappas K, Tripepi G, Georgiou I, Tselepis A, Elisaf M, Tsakiris D, Zoccali C, Siamopoulos K. A Genetic Biomarker of Oxidative Stress, the Paraoxonase-1 Q192R Gene Variant, Associates with Cardiomyopathy in CKD: A Longitudinal Study. *Oxid Med Cell Longev*. 2016;2016:1507270. doi:10.1155/2016/1507270
23. Mackness B, Turkie W, Mackness M. Paraoxonase-1 (PON1) promoter region polymorphisms, serum PON1 status and coronary heart disease. *Arch Med Sci*. 2013;9(1):8–13. doi:10.5114/aoms.2013.33189
24. Eckerson HW, Wyte CM, La Du BN. The human serum paraoxonase/arylesterase polymorphism. *Am J Hum Genet*. 1983;35(6):1126–1138.
25. Jakubowski H, Ambrosius WT, Pratt JH. Genetic determinants of homocysteine thiolactonase activity in humans: implications for atherosclerosis. *FEBS Lett*. 2001;491(1-2):35–39. doi:10.1016/s0014-5793(01)02143-3
26. Jakubowski H, Zhang L, Bardeguet A, Aviv A. Homocysteine thiolactone and protein homocysteinylation in human endothelial cells: implications for atherosclerosis. *Circ Res*. 2000;87(1):45–51. doi:10.1161/01.res.87.1.45
27. Eckerson HW, Wyte CM, La Du BN. The human serum paraoxonase/arylesterase polymorphism. *Am J Hum Genet*. 1983;35(6):1126–1138.
28. Zargari M, Sharafeddin F, Mahrooz A, Alizadeh A, Masoumi P. The common variant Q192R at the paraoxonase 1 (PON1) gene and its activity are responsible for a portion of the altered antioxidant status in type 2 diabetes. *Exp Biol Med*. 2016;241(14):1489–1496.