

Chapter 7

DEGRADABLE SYNTHETIC BIOPOLYMERS IN MEDICINE

Timur PAÇACI¹

1.INTRODUCTION

Biopolymers can be examined in three parts as natural, semi-natural and synthetic. Rather than natural and semi-natural polymers, in this section, synthetic degradable biopolymers in medicine and their properties are tried to be explained.

Biopolymers was founded in 1963 and gain importance in the medicine industry with their low toxicity, different physical and chemical properties. Biopolymers are divided into two as degradable and non-degradable polymers in nature. Degradable and non-degradable polymers are preferred depending on the usage life and place in metabolism. Their common point is that they are compatible with living metabolism. Biocompatibility, growth ability, production in large quantities and specific areas of use have given importance to synthetic biopolymers in the health sector. In this section, the properties of degradable synthetic biopolymers and their use in the field of medicine are emphasized.

2.DEGRADABLE SYNTHETIC BIOPOLYMERS IN MEDICINE

Today, studies are carried out on 10 different biodegradable synthetic polymers: polylactic acid, polyglycolic acid, polycaprolactone, polyhydroxybutyrate, polybutylene succinate, polyvinyl alcohol, polyethylene adipate, polyether sulfone, polyurethane and polyvinylpyrrolidone. Biodegradable polymers derived from these polymers are frequently emphasized by scientists due to their tensile and impact strength, low toxicity, ease of production on a large scale allowing the production of polymers with different physical properties with mixtures and their derivatives can be handled.

¹ Dr.; Institution: Tokat Science and Art School; e-mail: timurboun@gmail.com;
ORCID iD: 0009-0001-4378-8200

3.CONCLUSION

Biodegradable polymers, which have been produced for use in the medical field since the 1960s, are gaining more and more importance. This is evident in the number of publications. However, the fact that they do not have the desired level of physical and chemical resistance has made the development of biopolymers necessary. Biopolymers with biodegradable properties, on the other hand, are finding more and more medical uses with their easy dissolution in nature, high number of derivatives, biocompatibility, sufficient physical and chemical resistance. In this context, it is an inevitable fact that the newly developed biodegradable polymers will create more economic and wider usage areas thanks to their advanced physical and chemical properties. Oxygen concentration in polymers significantly affects their degradability and changes in chain length are important in the physical strength of polymers. The most striking disadvantage of degradable biopolymers is that they do not have sufficient physical strength depending on the place of use. The reasons for this are based on insufficient chain length or unsuitable crystal/amorphous region ratios (elasticity) in polymers.

In this section, the general properties of synthetic biodegradable polymers, which are frequently studied, and their usage areas in the medical sector are tried to be explained. Apart from these polymers, synthetic nondegradable biopolymers and natural biopolymers are also used in different ways in the medical field. It is promising for the future to make nondegradable polymers degradable by adding side groups or to develop semi-synthetic polymers with sufficient physical strength by changing the chain structures of natural polymers.

REFERENCES

1. Drumright RE, Gruber PR, Henton DE. Polylactic Acid Technology. *Advanced Materials*; 2000;12(23): 1841–1846. doi.org/10.1002/1521-4095(200012)12:23<1841::AID-ADMA1841>3.0.CO;2-E
2. Van de Velde K, Kiekens P. Biopolymers: overview of several properties and consequences on their applications, Department of Textiles, Ghent University; 2002;21: 433–442. doi.org/10.1016/S0142-9418(01)00107-6
3. Hartmann MH. High Molecular Weight Polylactic Acid Polymers. In: Kaplan, D.L. (eds) *Biopolymers from Renewable Resources*. Macromolecular Systems — Materials Approach. Springer, Berlin, Heidelberg, 1998.
4. Nijenhuis AJ, Du YJ, Van Aert HAM, Bastiaansen C. ABA Type Copolymers of Lactide with Poly(ethylene glycol). Kinetic, Mechanistic, and Model Studies. *Macromolecules*; 1995;28(7): 2124–2132. doi.org/10.1021/ma00111a004
5. Lasprilla AJR, Martinez GAR, Lunelli BH, Jardini AL, Filho RM. Poly-lactic acid synthesis for application in biomedical devices. A review. *Biotechnology Advances*; 2012;30(1):321–328. doi.org/10.1016/j.biotechadv.2011.06.019

6. Fukushima K, Kimura Y. Stereocomplexed polylactides (Neo-PLA) as high-performance bio-based polymers: their formation, properties, and application. *Polymer International*; 2006;55(6):626–642. <https://doi.org/10.1002/pi.2010>
7. Tracy MA, Ward KL. Firouzabadian, L.; Wang, Y.; Dong, N.; Qian, R.; Zhang, Y. Factors affecting the degradation rate of poly (lactide-co-glycolide) microspheres in vivo and in vitro. *Biomaterials*; 1999;20(11): 1057–1062. [https://doi.org/10.1016/S0142-9612\(99\)00002-2](https://doi.org/10.1016/S0142-9612(99)00002-2)
8. Liu L, Li S, Garreau H, Vert M. Selective Enzymatic Degradations of Poly(l-lactide) and poly(E-caprolactone) blend films. *Biomacromolecules*; 2000;1(3): 350–359. doi.org/10.1021/bm000046k
9. Soni S, Gupta H, Kumar N, Nishad D, Mittal G, Bhatnagar A. Biodegradable biomaterials; 2010;3(1): 30–40.
10. Nakafuku C, Yoshimura H. Melting parameters of poly(glycolic acid). *Polymer (Guildf)*; 2004;45(11): 3583–3585. doi.org/10.1016/j.polymer.2004.03.041
11. Polyglycolic Acid (PGA) Market Research Report - Global Forecast till 2030. <https://www.marketresearchfuture.com/reports/polyglycolic-acid-market-5749> (Accessed: August 2023)
12. Jahno VD. Síntese e caracterização do Poli (L-Ácido Láctico) para uso como biomaterial, 2005.
13. Nair LS, Laurencin CT. Biodegradable polymers as biomaterials. *Progress in Polymer Science*; 2007;32(8-9):762–98. doi.org/10.1016/j.progpolymsci.2007.05.017
14. Hayashi T. Biodegradable polymers for biomedical uses. *Progress in Polymer Science*; 1994;19(4): 663–702. [doi.org/10.1016/0079-6700\(94\)90030-2](https://doi.org/10.1016/0079-6700(94)90030-2)
15. Sinha VR, Bansal K, Kaushik R, Kumria R, Trehan A. Poly-ε-caprolactone microspheres and nanospheres: an overview. *International Journal of Pharmaceutics*; 2004;278(1): 1–23. doi.org/10.1016/j.ijpharm.2004.01.044
16. Woodruff MA, Huttmacher DW. The return of a forgotten polymer—Polycaprolactone in the 21st century. *Progress in Polymer Science*; 2010;35(10): 1217–1256. doi.org/10.1016/j.progpolymsci.2010.04.002
17. Lohmeijer BGG, Pratt RC, Leibfarth F, Logan JW, Long DA, Dove AP, Nederberg F, Choi Wade C, Waymouth RM, Hedrick JL. Guanidine and amidine organocatalysts for ring-opening polymerization of cyclic esters. *Macromolecules*; 2006;39(25): 8574–8583. doi.org/10.1021/ma0619381
18. Yuan M, Xiong C, Deng X. Ring-opening polymerization of ε-caprolactone initiated by cyclopentadienyl sodium. *Journal of Applied Polymer Science*; 1998;67(7): 1273–1276. [doi.org/10.1002/\(SICI\)1097-4628\(19980214\)67:7<1273::AID-APP17>3.0.CO;2-2](https://doi.org/10.1002/(SICI)1097-4628(19980214)67:7<1273::AID-APP17>3.0.CO;2-2)
19. Kowalski A, Duda A, Penczek S. Kinetics and Mechanism of Cyclic Esters Polymerization Initiated with Tin(II) Octoate. 3.† Polymerization of l,l-Dilactide. *Macromolecular Rapid Communications*; 2000;33(20) 77359-7370. doi.org/10.1021/ma000125o
20. Yamashita M, Takemoto Y, Ihara E, Yasuda H., Organolanthanide-Initiated Living Polymerizations of ε-Caprolactone, δ-Valerolactone, and β-Propiolactone. *Macromolecules*; 1996;29(5), 1798-1806. doi.org/10.1021/ma951400n
21. Williams MD, Rahn JA, Sherman DH. Production of a polyhydroxyalkanoate biopolymer in insect cells with a modified eucaryotic fatty acid synthase. *Applied and Environmental Microbiology*; 1996;62(7): 2540-2546. doi.org/10.1128/aem.62.7.2540-2546.1996

22. Patnaik P. “Intelligent” descriptions of microbial kinetics in finitely dispersed bioreactors: neural and cybernetic models for PHB biosynthesis by *Ralstonia eutropha*. *Microbial Cell Factories*; 2007;6: 23-25. doi.org/10.1186/1475-2859-6-23
23. Lee C, Song B, Jegal J, Kimura Y. Cell adhesion and surface chemistry of biodegradable aliphatic polyesters: discovery of particularly low cell adhesion behavior on poly (3-RS.-hydroxybutyrate. *Macromolecular Research*; 2003;21: 305–1313. doi.org/10.1007/s13233-013-1181-8
24. Simon F, Martin DP. Applications of PHAs in medicine and pharmacy. In: Y. Doi, A. Steinbüchel (eds) *Biopolymers*, 4th edn. Wiley-VCH, Weinham; 2002;91–103.
25. Yagmurlu MF, Korkusuz F, Gursel I, Korkusuz P, Ors U, Hasirci V. Sulbactam-cefoperazone polyhydroxybutyrate-co-hydroxyvalerate (PHBV) local antibiotic delivery system: in vivo effectiveness and biocompatibility in the treatment of implant-related experimental osteomyelitis. *Journal of Biomedical Materials Research*; 1999; 46(4): 494–503. doi.org/10.1002/(SICI)1097-4636(19990915)46:4<494::AID-JB-M7>3.0.CO;2-E
26. Xu J, Guo BH. Poly(butylene succinate) and its copolymers: Research, development and industrialization. *Biotechnology Journal*; 2010;5(11): 1149–1163. doi.org/10.1002/biot.201000136
27. Ishioka R, Kitakuni E, Ichikawa Y. Aliphatic polyesters: “Bionolle”. In: Doi, Y., Steinbüchel, A. (Eds.), *Biopolymers, Polyesters III Applications and Commercial Products*; 2002;4:275–297. doi.org/10.1002/3527600035.bpol4010
28. Platnieks O, Gaidukovs S, Thakur VK, Barkane A, Beluns S. Bio-based poly (butylene succinate): Recent progress, challenges and future opportunities. *European Polymer Journal*; 2021;161:110855. doi.org/10.1016/j.eurpolymj.2021.110855.
29. Mtibe A, Muniyasamy S, Mokhena TC, Ofosu O, Ojijo V, John M. Recent insight into the biomedical applications of polybutylene succinate and polybutylene succinate-based materials. *Express Polymer Letters*; 2023;17(1): 2–28. doi.org/10.3144/expresspolymlett.2023.2
30. Cicero L, Licciardi M, Cirincione R, Puleio R, Giammona G, Giglia G, Sardo P, Vigni GE, Cioffi A, Sanfilippo A, Cassata G. Polybutylene succinate artificial scaffold for peripheral nerve regeneration. *Journal of Biomedical Materials Research Part B Applied Biomaterials*; 2022;110(1): 125–134. doi.org/10.1002/jbm.b.34896
31. Mtibe A, Motloung MP, Bandyopadhyay J, Ray SS. Synthetic biopolymers and their composites: Advantages and limitations – An Overview. *Macromolecular Rapid Communications*; 2021;42(15): 2100130. doi.org/10.1002/marc.202100130
32. Das R, Kundu D. Structural and Transport Properties of Norbornene-Functionalized Poly(vinyl alcohol) “Click” Hydrogel: A Molecular Dynamics Study. *ACS Sustainable Chemistry & Engineering*; 2023;11(29): 10812-10824. doi.org/10.1021/acssuschemeng.3c01948
33. Hassan CM, Peppas NA. Structure and Applications of Poly(vinyl alcohol) Hydrogels Produced by Conventional Crosslinking or by Freezing/Thawing Methods. In: *Biopolymers · PVA Hydrogels, Anionic Polymerisation Nanocomposites*. *Advances in Polymer Science*; 2000;153:37-65. doi.org/10.1007/3-540-46414-X_2
34. Wan WK, Campbell G, Zhang ZF, Hui AJ, Boughner DR. Optimizing the tensile properties of polyvinyl alcohol hydrogel for the construction of a bioprosthetic heart valve stent. *Journal of Biomedical Materials Research*; 2002;63(6): 854-61. doi.org/10.1002/jbm.10333

35. Peppas NA, Benner RE. Proposed method of intracordal injection and gelation of poly (vinyl alcohol) solution in vocal cords: polymer considerations. *Biomaterials*; 1980;1(3):158-62. doi.org/10.1016/0142-9612(80)90039-3
36. Paul W, Sharma CP. Polyacrylonitrile-reinforced poly(vinyl alcohol) membranes: Mechanical and dialysis performance. *Journal of Applied Polymer Science*; 1995;57(12): 1447-1454. doi.org/10.1002/app.1995.070571204
37. <https://www.rxlist.com/> (access date:08.16.2023)
38. McNeil C, Basan S. Thermal degradation of blends of PVC with poly(ethylene adipate). *Polymer Degradation and Stability*; 1993;41(3): 311-17. doi.org/10.1016/0141-3910(93)90077-V
39. Monvisade P, Loungvanidprapa P. Synthesis of poly(ethylene adipate) and poly(ethylene adipate-co-terephthalate) via ring-opening polymerization. *European Polymer Journal*; 2007;43(8): 3408-3414. doi.org/10.1016/j.eurpolymj.2007.05.009
40. Chen L, Xu J, Xue W, Zeng Z. Mechanism and kinetics of esterification of adipic acid and ethylene glycol by tetrabutyl titanate catalyst. *Korean Journal of Chemical Engineering*; 2017;35(1): 82-88. doi.org/10.1007/s11814-017-0276-x
41. Jin HJ, Lee BY, Kim MN, Yoon JS. Properties and biodegradation of poly(ethylene adipate) and poly(butylene succinate) containing styrene glycol units. *European Polymer Journal*; 2000;36(12): 2693-2698. [https://doi.org/10.1016/S0014-3057\(00\)00057-4](https://doi.org/10.1016/S0014-3057(00)00057-4)
42. Atanase LI, Salhi S, Cucoveica O, Ponjavic M, Nikodinovic-Runic J, Delaite C. Biodegradability Assessment of Polyester Copolymers Based on Poly(ethylene adipate) and Poly(ϵ -caprolactone). *Polymers*; 2022;14(18): 3736. <https://doi.org/10.3390/polym14183736>
43. Atkins TW. Biodegradation of poly(ethylene adipate) microcapsules in physiological media. *Biomaterials*; 1998;19(1-3): 61-67. [https://doi.org/10.1016/S0142-9612\(97\)00156-7](https://doi.org/10.1016/S0142-9612(97)00156-7)
44. Anthierens T, Billiet L, Devlieghere F, Du Prez F. Poly(butylene adipate) functionalized with quaternary phosphonium groups as potential antimicrobial packaging material. *Innovative Food Science & Emerging Technologies*, 2012;15: 81-85. <https://doi.org/10.1016/j.ifset.2012.02.010>
45. Bikiaris D, Karavelidis V, Karavas E. Novel Biodegradable Polyesters. Synthesis and Application as Drug Carriers for the Preparation of Raloxifene HCl Loaded Nanoparticles. *Molecules*; 2009;14(7): 2410-2430. <https://doi.org/10.3390/molecules14072410>
46. Rahimpour A, Madaeni SS, Mehdipour-Ataei S. Synthesis of a novel poly (amide-imide)(PAI) and preparation and characterization of PAI blended polyethersulfone (PES) membranes. *Journal of Membrane Science*; 2008;311(1-2):349-359. <https://doi.org/10.1016/j.memsci.2007.12.038>
47. <https://omnexus.specialchem.com/selection-guide/polyethersulfone-pesthermoplastic/key-applications> (access date: 09.02.2023)
48. Burg KJL, Shalaby SW. PES and PEEK. *Encyclopedia of Materials: Science and Technology*; 2001;6837-6839. 10.1016/B0-08-043152-6/01212-2
49. Lelah MD, Cooper JL. *Polyurethanes in Medicine*. Boca Raton, FL: CRC Press, ISBN: 0849363071,1987.
50. Tatai L, Moore TG, Adhikari R, Malherbe F, Jayasekara R, Griffiths I, Gunatillake PA. Thermoplastic biodegradable polyurethanes: The effect of chain extender structure on properties and in-vitro degradation. *Biomaterials*; 2007;28(36): 5407-5417. doi.org/10.1016/j.biomaterials.2007.08.035

51. Cohn D, Lando G, Sosnik A, Garty S, Levi A. PEO-PPG-PEO-based poly(ether ester urethane)s as degradable reverse thermo-responsive multiblock copolymers. *Biomaterials*; 2006;27(9): 1718. <https://doi.org/10.1016/j.biomaterials.2005.10.035>
52. Loomis K, McNeeley K, Bellamkonda V. Nanoparticles with targeting, triggered release, and imaging functionality for cancer applications. *Soft Matter*; 2011;7(3): 839–856. DOI: 10.1039/c0sm00534g
53. Guelcher SA. Biodegradable Polyurethanes: Synthesis and Applications in Regenerative Medicine. *Tissue Engineering Part B: Reviews*, 2008;14(1): 3–17. doi.org/10.1089/teb.2007.0133
54. Santerre JP, Woodhouse K, Laroche G, Labow RS. Understanding the biodegradation of polyurethanes: From classical implants to tissue engineering materials. *Biomaterials*; 2005;26: 7457–7470. <https://doi.org/10.1016/j.biomaterials.2005.05.079>
55. Ding M, Li J, Tan H, Fu Q. Self-assembly of biodegradable polyurethanes for controlled delivery applications. *Soft Matter*; 2012;8(20): 5414. DOI: 10.1039/c2sm07402h
56. Kurakula M, Rao GSNK. Type of Article: REVIEW Pharmaceutical Assessment of Polyvinylpyrrolidone (PVP): As Excipient from Conventional to Controlled Delivery Systems with a Spotlight on COVID-19 Inhibition. *Journal of Drug Delivery Science and Technology*; 2020;60: 102046. <https://doi.org/10.1016/j.jddst.2020.102046>
57. Christensen M, Johansen P, Hau C. Storage of polyvinylpyrrolidone (PVP) in tissues following long-term treatment with a PVP-containing vasopressin preparation. *Acta Medica Scandinavica*; 1978;204(1-6): 295–298. <https://doi.org/10.1111/j.0954-6820.1978.tb08442.x>
58. Bühler V. Polyvinylpyrrolidone – Excipients for Pharmaceuticals: Povidone, Crospovidone and Copovidone, Illustrate, Springer-Verlag, Berlin Heidelberg, New York, 2005;67.