

BÖLÜM 44

HAYVAN DENEYLERİNDE GÖZ MODELLERİ

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Oftalmolojik hastalıklara neden olan risk faktörlerinin belirlenmesi, fizyopatolojide yer alan mekanizmaların aydınlatılması ve belirli bir tedavi protokolünün insanlar için güvenli ve etkili olup olmayacağını anlamak için, çeşitli hayvan modelleri kullanılarak birçok deneysel çalışma yapılmaktadır (1). Planlanan deneysel çalışmaların doğru sonuçlar vermesi ve klinik uygulamalarda yol gösterici olması için üzerinde durulması gereken en önemli nokta, kullanılacak modelin istenen amaca uygun olmasıdır. Bu nedenle deneysel hayvan çalışmaları planlanırken insan gözünün anatomisi ve fizyolojisine en yakın benzerliğe sahip hayvanların denek olarak seçilmesine dikkat edilmelidir (2). Üzerinde çalışılan hastalığın doğal olarak geliştiği hayvanların denek olarak kullanılması, fizyopatolojik sürecin daha iyi anlaşılmasına neden olur. Ancak bu koşulu sağlamak her zaman mümkün değildir.

İnsan göz hastalıklarının incelenmesi için çeşitli kemirgenler (fareler, ratlar ve kobaylar), memeliler (tavşanlar, köpekler ve primatlar), omurgasızlar (meyve sinekleri ve nematodlar) ve zebra balığı modelleri geliştirilmiştir. Bu modellerden en yaygın olarak kullanılanı faredir. Çünkü fare gözü yapısal olarak insan gözüne benzer ve insanlardaki birçok oküler bozukluk farelerde de gözlenir. Ayrıca insan ve fare genomları, kodlama bölgelerinde önemli bir deoksiribonükleik asit (DNA) dizi homolojisi sergiler. Böylece glokom ve yaşa bağlı makula dejenerasyonu

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fare sınıfı II moleküllerinin silindiđi ve insan muadilleriyle deđiřtirildiđi HLA sınıf II transgenik farelerde retina antijeni ile ařılama yoluyla da indüklenebilir. HLA-DR3, HLA-DR4, HLA-DQ6 veya HLA-DQ8 transgenik fareler, bu sınıf II molekülleri taşıyan insanlar tarafından tanınabilecek antijenik epitoplar sunar ve bunlara yanıt verir (85). HLA-Tg fareleri, yalnızca IRBP ile deđil, S-Ag ile ařıldıklarında da řiddetli üveit geliřtirir. MHC sınıf II molekülleri antijen tanımada yer aldıđından, HLA sınıf II Tg fareleri, insan hastalıđına karıřan retinal antijenlerin kritik bölgelerini belirlemeye yardımcı olabilir ve insan üveiti için antijene özđü tedaviler bulmak için önemli olabilir (81,85).

Prematüre Retinopatisi

Yenidođanlarda prematüre retinopatisini (ROP) incelemek için geliřtirilen “oksijenin neden olduđu retinopati modeli” ilk olarak Smith laboratuvarı tarafından kullanılmıřtır (88). Bu modelde, dođum sonrası 7. gün hayvanlar (P7) 5 gün boyunca yüksek kısmi oksijen basıncına (~ %75 oksijen) maruz bırakılır, bunun ardından normal retina damar geliřimi durur ve VEGF azalmasına bađlı olarak merkezi retinada bulunan intraretinal kılcal damarlar dejenere olur. Hayvanlar daha sonra yüksek oksijen ortamından normal atmosferik kořullara (% 21 oksijen) çıkarılır, ardından yetersiz kan ve oksijen kaynađı nedeniyle retina řiddetli iskemiye maruz kalır ve retina öncesi neovaskularizasyon meydana gelir (88). Retinal hipoksi, PDR gibi iskemik retinopatilerin bir özelliđidir ve oksijene bađlı retinopati modeli, bu durumu makulada oldukça belirgin bir řekilde sađlar (65). Retinada glia ve nöronların oksijen yoksunluđu, oksijene bađlı retinopatide pro-anjiyogenik büyüme faktörü ekspresyonunu yönlendirir ve ortaya çıkan neovasküler model vazojenik ajanların deđerlendirilmesinde yaygın olarak kullanılmaktadır (65) .

KAYNAKLAR

1. Chan C-C, ed. Animal models of ophthalmic diseases. Essentials in Ophthalmology. Cham, Switzerland: Springer International Publishing; 2016:1-152.
2. Bahçeci UA, Özdek ř, Konuk O, ve ark. Oftalmolojide Kullanılan Deneysel Hayvan Modelleri. T Klin Oftalmoloji 2004, 13:109-116.
3. Xu F, Schillinger JA, Sternberg MR, et al. Seroprevalence and coinfection with herpes simplex virus type 1 and type 2 in the United States, 1988–1994. J Infect Dis 2002, 185:1019–24
4. Wald A, Corey L. Persistence in the population: epidemiology, transmission. Human herpesviruses: biology, therapy, and immunoprophylaxis. Cambridge University Press, Cambridge; 2007

5. Hendricks RL, Yun H, Rowe AM, et al. Animal Models of Herpes Keratitis. In: Chan C-C, ed. *Animal Models of Ophthalmic Diseases*. Cham, Switzerland: Springer; 2016: 1–10.
6. Santos C, Briones O, Dawson CR Peripheral adrenergic stimulation and indomethacin in experimental ocular shedding of HSV. *Curr Eye Res* 1987; 6:111–118
7. Knickelbein JE, Khanna KM, Ye MB, et al. Noncytotoxic lytic granule-mediated CD8+ T cell inhibition of HSV-1 reactivation from neuronal latency. *Science* 2008 Oct 10;322(5899):268-71.
8. Holland EJ, Schwartz GS, Neff KD (2011) Herpes simplex keratitis. In: Krachmer JH, Mannis MJ, Holland EJ (eds) *Cornea*, 3rd edn. Mosby Year Book Publishers, St Louis
9. Fenton RR, Molesworth-Kenyon S, et al. Linkage of IL-6 with neutrophil chemoattractant expression in virus-induced ocular inflammation. *Invest Ophthalmol Vis Sci* 2002 Mar;43(3):737-43.
10. WHO Visual impairment and blindness. <http://www.who.int/mediacentre/factsheets/fs282/en/index>
11. West-Mays J, Bowman S. Animal Models of Cataracts. In: Chan C-C, ed. *Animal Models of Ophthalmic Diseases*. Cham, Switzerland: Springer; 2016: 11–29.
12. Talbot WS, Hopkins N Zebrafish mutations and functional analysis of the vertebrate genome. *Genes Dev* 2000;14(7):755–762
13. Glass AS, Dahm R The zebrafish as a model organism for eye development. *Ophthalm Res* 2004;36(1):4–24
14. Hejtmancik JF Congenital cataracts and their molecular genetics. *Seminars Cell Dev Biol* 2008;19(2):134–149.
15. Obrosova IG, Chung SS, Kador PF Diabetic cataracts: mechanisms and management. *Diabetes/Metabolism Res Rev* 2010;26(3):172–180.
16. Merriam JC, Lofgren S, Michael R, et al. An action spectrum for UV-B radiation and the rat lens. *Invest Ophthalmol Vis Sci* 2000;41(9):2642–2647
17. Lassen N, Bateman JB, Estey T, et al. Multiple and additive functions of ALDH3A1 and ALDH1A1: cataract phenotype and ocular oxidative damage in *Aldh3a1*($-/-$)/*Aldh1a1*($-/-$) knock-out mice. *J Biol Chem* 2007;282(35):25668–25676.
18. Meyer LM, Lofgren S, Ho YS, et al. Absence of glutaredoxin1 increases lens susceptibility to oxidative stress induced by UVR-B. *Exp Eye Res* 2009;89(6):833–839.
19. Zhang J, Yan H, Lofgren S, et al. Ultraviolet radiation-induced cataract in mice: the effect of age and the potential biochemical mechanism. *Invest Ophthalmol Vis Sci* 2012;53(11):7276–7285
20. Galichanin K, Lofgren S, Bergmanson J, et al. Evolution of damage in the lens after in vivo close to threshold exposure to UV-B radiation: cytomorphological study of apoptosis. *Exp Eye Res* 2010;91(3):369–377.
21. Giblin FJ, Lin LR, Simpanya MF, et al. A Class I UV-blocking (senofilcon A) soft contact lens prevents UVA-induced yellow fluorescence and NADH loss in the rabbit lens nucleus in vivo. *Exp Eye Res* 2012;102:17–27.
22. Lyu J, Kim JA, Chung SK, et al. Alteration of cadherin in dexamethasone-induced cataract organ-cultured rat lens. *Invest Ophthalmol Vis Sci* 2003;44(5):2034–2040
23. Wang C, Dawes LJ, Liu Y, et al. Dexamethasone influences FGF-induced responses in lens epithelial explants and promotes the posterior capsule coverage that is a feature of gluco- corticoid-induced cataract. *Exp Eye Res* 2013;111:79–87.

24. Gwon A. The rabbit in cataract/IOL surgery. In: Tsonis P (ed) *Animal models in eye research*. Elsevier, New York (2008)
25. Manthey AL, Terrell AM, Wang Y, et al. The Zeb proteins deltaEF1 and Sip1 may have distinct functions in lens cells following cataract surgery. *Invest Ophthalmol Vis Sci* 2014;55(8):5445–5455.
26. Stamper RL LM, Drake MV. Primary open angle glaucoma. In: Stamper RL LM, Drake MV, editor. *Becker-Shaffer's Diagnosis and Therapy of the Glaucomas*. Missouri: Mosby; 2009. p. p.239-65.
27. Tham YC, Li X, Wong TY, et al. Global prevalence of glaucoma and projections of glaucoma burden through 2040: asystematic review and meta-analysis. *Ophthalmology* 2014;121:2081–2090
28. Johnson TV, Tomarev SI *Animal Models of Glaucoma*. In: Chan C-C, ed. *Animal Models of Ophthalmic Diseases*. Cham, Switzerland: Springer; 2016: 31–50
29. Pease ME, Cone FE, Gelman S, et al. Calibration of the TonoLab tonometer in mice with spontaneous or experimental glaucoma. *Invest Ophthalmol Vis Sci* 2011; 52:858–864
30. Morrison JC, Moore CG, Deppmeier LM, et al. A rat model of chronic pressure-induced optic nerve damage. *Exp Eye Res* 1997;64:85–96
31. Shareef SR, Garcia-Valenzuela E, Salierno A, et al. Chronic ocular hypertension following episcleral venous occlusion in rats. *Exp Eye Res* 1995;61:379–382
32. Johnson TV, Tomarev SI. Rodent models of glaucoma. *Brain Res Bull* 2010;81:349–358
33. Levkovitch-Verbin H, Quigley HA, Martin KR, et al. Translimbal laser photocoagulation to the trabecular meshwork as a model of glaucoma in rats. *Invest Ophthalmol Vis Sci* 2002;43:402–410
34. Feng L, Chen H, Suyeoka G, et al. A laser- induced mouse model of chronic ocular hypertension to characterize visual defects. *J Vis Exp* 2013;(78):50440.
35. Mc Kinnon SJ, Pease ME, Wolde Mussie E et al. Comparison of three models of rat glaucoma caused by chronic intraocular pressure elevation . *Invest Ophthalmol Vis Sci* 1999;40:787.
36. Belforte N, Sande PH, de Zavalía N, et al. (2012) Therapeutic benefit of radial optic neurotomy in a rat model of glaucoma. *PloS ONE* 7:e34574
37. Urcola JH, Hernandez M, Vecino E. Three experimental glaucoma models in rats: comparison of the effects of intraocular pressure elevation on retinal ganglion cell size and death. *Exp Eye Res* 2006;83:429–437
38. Buie LK, Karim MZ, Smith MH, et al. Development of a model of elevated intraocular pressure in rats by gene transfer of bone morphogenetic protein 2. *Invest Ophthalmol Vis Sci* 2013;54:5441–5455
39. Giovingo M, Nolan M, McCarty R, et al. sCD44 overexpression increases intra-ocular pressure and aqueous outflow resistance. *Mol Vis* 2013;19:2151–2164
40. Shepard AR, Millar JC, Pang IH, et al. Adenoviral gene transfer of active human transforming growth factor- β 2 elevates intraocular pressure and reduces outflow facility in rodent eyes. *Invest Ophthalmol Vis Sci* 2010;51:2067–2076.
41. Zode GS, Sharma AB, Lin X, et al. Ocular-specific ER stress reduction rescues glaucoma in murine glucocorticoid-induced glaucoma. *J Clin Invest* 2014;124:1956–1965

42. Overby DR, Bertrand J, Tektas OY, et al. Ultrastructural changes associated with dexamethasone induced ocular hypertension in mice. *Invest Ophthalmol Vis Sci* 2014;55:4922–4933
43. Stone EM, Fingert JH, Alward WL, et al. Identification of a gene that causes primary open angle glaucoma. *Science* 1997;275:668–670
44. Junglas B, Kuespert S, Seleem AA, et al. Connective tissue growth factor causes glaucoma by modifying the actin cyto- skeleton of the trabecular meshwork. *Am J Pathol* 2012;180:2386–2403
45. Browne JG, Ho SL, Kane R, et al. Connective tissue growth factor is increased in pseudoexfoliation glaucoma. *Invest Ophthalmol Vis Sci* 2011;52:3660–3666
46. Libby RT, Anderson MG, Pang IH, et al. Inherited glaucoma in DBA/2J mice: pertinent disease features for studying the neurodegeneration. *Vis Neurosci* 2005;22:637–648
47. Pang JH, Cantu-Crouch D, Savinova OV et al. Age-dependent changes in ocular morphology of a spontaneous ocular hypertensive mouse strain . *Invest Ophthalmol Vis. Sci* 1999;40:671.
48. Bayer AU, Neuhardt T, May AC et al. Retinal morphology and ERG response in the DBA/2NNIA mouse model of angle -closure glaucoma . *Invest Ophthalmol Vis. Sci* 2001;42:1258-65.
49. Aung T, Rezaie T, Okada K, et al. Clinical features and course of patients with glaucoma with the E50K mutation in the optineurin gene. *Invest Ophthalmol Vis Sci* 2005;46:2816–2822
50. Hauser MA, Allingham RR, Linkroum K, et al. Distribution of WDR36 DNA sequence variants in patients with primary open-angle glaucoma. *Invest Ophthalmol Vis Sci* 2006;47:2542–2546
51. Harada T, Harada C, Nakamura K, et al. The potential role of glutamate transporters in the pathogenesis of normal tension glaucoma. *J Clin Invest* 2007;117:1763–1770
52. Rastoin O, Pagès G, Dufies M. Experimental Models in Neovascular Age Related Macular Degeneration. *Int J Mol Sci.* 2020;21(13):4627.
53. Sennlaub F., *Animal Models of Age-Related Macular Degeneration.* In: Chan C-C, ed. *Animal Models of Ophthalmic Diseases.* Cham, Switzerland: Springer; 2016: 51-65.
54. Swaroop A, Branham KE, Chen W, et al. Genetic susceptibility to age-related macular degeneration: a paradigm for dissecting complex disease traits. *Human Mol Genet* 2007;16 (Spec No. 2):R174–R182
55. Chakravarthy U, Wong TY, Fletcher A, et al. Clinical risk factors for age- related macular degeneration: a systematic review and meta-analysis. *BMC Ophthalmol* 2010;10:31
56. Calippe B, Guillonneau X, Sennlaub F. Complement factor H and related proteins in age-related macular degeneration. *CR Biol* 2014;337(3):178–184
57. Ufret-Vincenty RL, Aredo B, Liu X, et al. Transgenic mice expressing variants of complement factor H develop AMD- like retinal findings. *Invest Ophthalmol Vis Sci* 2012;51(11):5878–5887
58. Liao S-M, Crowley M, Louie S, et al. HtrA1 regulates the subretinal infiltration of microglia cells in response to bacterial lipopolysaccharides (LPS) and aging in mice. *Invest Ophthalmol Vis Sci* 2013;54(6):3666

59. Xu H, Chen M, Manivannan A, et al. Age-dependent accumulation of lipofuscin in perivascular and subretinal microglia in experimental mice. *Aging Cell* 2008;7(1):58–68
60. Ng TF, Streilein JW Light-induced migration of retinal microglia into the subretinal space. *Invest Ophthalmol Vis Sci* 2001;42(13):3301–3310
61. Zhao Z, Chen Y, Wang J et al. Age-related retinopathy in NRF2-deficient mice. *PLoS ONE* 2011, 6, e19456.
62. Shah RS, Soetikno BT, Lajko M et al. A mouse model for laser-induced choroidal neovascularization. *JoVE* 2015, 53502.
63. Antonetti DA, Klein R, Gardner TW Diabetic retinopathy. *N Engl J Med* 2012;366(13):1227–39
64. Stitt AW, Lois N, Medina RJ et al. Advances in our understanding of diabetic retinopathy. *Clin Sci (Lond)* 2013;125(1):1–17
65. Chen M, Stitt A, Animal Models of Diabetic Retinopathy. In: Chan C-C, ed. *Animal Models of Ophthalmic Diseases*. Cham, Switzerland: Springer; 2016: 66-83.
66. Jousseaume AM, Smyth N, Niessen C. Pathophysiology of diabetic macular edema. *Dev Ophthalmol* 2007;39:1–12
67. Lai AK, Lo AC. Animal models of diabetic retinopathy: summary and comparison. *J Diabetes Res* 2013:106594
68. Zhang SX, Ma JX, Sima J et al. Genetic difference in susceptibility to the blood-retina barrier breakdown in diabetes and oxygen-induced retinopathy. *Am J Pathol* 2005;166(1):313–21
69. Kern TS, Miller CM, Tang J et al. Comparison of three strains of diabetic rats with respect to the rate at which retinopathy and tactile allodynia develop. *Mol Vis* 2010;16:1629–39
70. Robinson R, Barathi VA, Chaurasia SS et al. Update on animal models of diabetic retinopathy: from molecular approaches to mice and higher mammals. *Dis Model Mech* 2012;5(4):444–56
71. Cox O, Stitt AW, Simpson DA, Gardiner TA. Sources of PDGF expression in murine retina and the effect of short-term diabetes. *Mol Vis* 2003;10(9):665–72
72. McVicar CM, Hamilton R, Colhoun LM. Intervention with an erythropoietin-derived peptide protects against neuroglial and vascular degeneration during diabetic retinopathy. *Diabetes* 2011;60(11):2995–3005
73. Saidi T, Mbarek S, Omri S, et al. The sand rat, *Psammomysobesus*, develops type 2 diabetic retinopathy similar to humans. *Invest Ophthalmol Vis Sci* 2011;52(12):8993–9004
74. Tang J, Kern TS Inflammation in diabetic retinopathy. *Prog Retin Eye Res* 2011;30(5):343–58
75. Anderson HR, Stitt AW, Gardiner TA et al. Induction of alloxan/streptozotocin diabetes in dogs: a revised experimental technique. *Lab Anim* 1993;27(3):281–5
76. Kador PF, Takahashi Y, Sato S, Wyman M Amelioration of diabetes-like retinal changes in galactose-fed dogs. *Prev Med* 1994;23(5):717–21
77. Olsen AS, Sarras MP Jr, Intine RV Limb regeneration is impaired in an adult zebrafish model of diabetes mellitus. *Wound Repair Regen* 2010;18(5):532–42
78. Gleeson M, Connaughton V, Arneson LS Induction of hyperglycaemia in zebrafish (*Danio rerio*) leads to morphological changes in the retina. *Acta Diabetol* 2007;44(3):157–63

79. Srivastava A, Rajappa M, Kaur J Uveitis: Mechanisms and recent advances in therapy. *Clin Chim Acta* 2010;411(17–18):1165–1171
80. Lee RW, Nicholson LB, Sen HN et al. Autoimmune and autoinflammatory mechanisms in uveitis. *Semin Immunopathol* 2014;36:581–594
81. Kielczewski J.L., Caspi R.R. Animal Models of Autoimmune Uveitis. In: Chan C-C, ed. *Animal Models of Ophthalmic Diseases*. Cham, Switzerland: Springer; 2016: 84–100.
82. Agarwal RK, Silver PB, Caspi RR Rodent models of experimental autoimmune uveitis. *Methods Mol Biol* 2012;900:443–469
83. Levy RA, de Andrade FA, Foeldvari I. Cutting- edge issues in autoimmune uveitis. *Clin Rev Allergy Immunol* 2011;41(2):214–223
84. Zeiss CJ. Translational models of ocular disease. *Vet Ophthalmol* 2013;16(Suppl 1):15–33
85. Mattapallil MJ, Sahin A, Silver PB. Common genetic determinants of uveitis shared with other autoimmune disorders. *J Immunol* 2008;180(10):6751–6759
86. Wildner G, Diedrichs-Mohring M, Thureau SR. Rat models of autoimmune uveitis. *Ophthalm Res* 2008;40(3–4):141–144
87. Papotto PH, Marengo EB, Sardinha LR. Immunotherapeutic strategies in autoimmune uveitis. *Autoimmun Rev* 2014;13:909–916
88. Smith LE, Wesolowski E, McLellan A. Oxygen-induced retinopathy in the mouse. *Invest Ophthalmol Vis Sci* 1994;35(1):101–11