

17. BÖLÜM

NON-INVAZİV PRENATAL TARAMA TEKNOLOJİSİ VE GELECEĞİ

Aydeniz AYDIN GÜMÜŞ¹
Ertaç GÜMÜŞ²

GİRİŞ

Non-invazif prenatal tarama (NIPT), gebelik sırasında maternal kandan fetüse ait genetik anormallikleri belirlemeye yönelik yapılan bir testtir. NIPT, maternal plazma içinde dolaşan hücre içermeyen serbest fetal DNA'dan (circulating cell free fetal-cffDNA) fetüse ait bir takım genetik anormallikleri öngörmeye yönelik bir testtir(1). Pek çok ülkede, trizomi 21 (Down sendromu), 18 (Edward sendromu) ve 13 (Patau sendromu) için tarama rutin doğum öncesi takiplerin bir parçası olarak sunulmaktadır. Bu, kombine tarama (ense kalınlığı, gebelikle ilişkili plazma protein A (PAPP-A), insan koryonik gonadotropin β (β HCG) ve anne yaşı) kullanılarak gebeliğin ilk tremesterinde veya dörtlü tarama (β HCG, konjuge olmayan östriol, α -fetoprotein ve inhibin A) kullanılarak ikinci tremesterde yapılmaktadır. Bu testlerde yüksek risk saptanan gebelerde koryonik villus örneklemesi veya amniyosentez invaziv testlerinin yapılması önerilmektedir. Bu invaziv prosedürlerin düşüğe yol açma riski bulunmaktadır(2). NIPT bu riskleri ortadan kaldırmaya yönelik geliştirilmekte olan en yeni tarama aracı olarak güncel prosedürler arasında yerini almaya başlamıştır.

SİRKÜLE(SERBEST) FETAL DNA

Gebelikte, gelişmekte olan fetüse ait hücre dışı serbest DNA (cffDNA), erkek fetüs taşıyan gebelerin plazmasında Y kromozoma ait DNA'nın tespit edilmesi ile bulunmuştur(1) cffDNA'nın kaynağı plasentanın sinsityotroblast tabakasıdır(3). Bir yıl sonra Lo ve ark., fetal DNA konsantrasyonunun maternal plaz-

¹ Uzm. Dr., Başakşehir Çam ve Sakura Şehir Hastanesi, Tıbbi Genetik, aydenizaydingumus@gmail.com

² Uzm. Dr., Manisa Şehir Hastanesi, Kadın Hastalıkları ve Doğum, ertacgumus@gmail.com

tespitini zorlaştıran homolog psödojenlere sahip genler, en önemlisi CYP21A2 ile ilişkili konjenital adrenal hiperplazide(95) RMD yaklaşımının aksine, RHD klinik olarak uygulanmaktadır ve Duchenne musküler distrofi(96), spinal musküler atrofi(97) ve kistik fibrozis(98) hastalıkları için NIPT hizmetleri artık Birleşik Krallık Sağlık Sisteminde mevcuttur.

Son zamanlarda, dPCR ve NGS gibi gelişmiş teknolojiler yüksek hassasiyetleri ve maternal plazma DNA'sından tüm fetal genomu açığa çıkarma olanakları nedeniyle NIPT'nin klinik uygulamasına izin vermişlerdir. Gelecekte, bu yöntemlerin anneden miras alınan mutasyonları da tespit edebilmesi beklenmektedir(99,100).cffDNA'da tüm fetal genom izlenebilmektedir ve genom sekanslama metodolojileri, ticari sağlayıcıların, cinsiyet kromozom anormallikleri, NOT'lar ve CNV sendromları dahil olmak üzere daha geniş bir fetal genetik anormallik yelpazesini rapor etmesine izin vermiştir. Yeni gelişen teknoloji ile NIPT ile babadan kalıtılanlar dahil olmak üzere monogenik hastalıkların tümüne yönelik ön tarama imkanının ailelere sunulması gelecek vizyonları içinde yerini almaya başlamıştır. Küçük CNV'lerin tespiti ve yorumlanmasıının açıklığa kavuşması ile tüm genom sekanslama tabanlı NIPT yaklaşımının prenatal takiplerde hekim ve danışan arasındaki belirsizlikleri en aza indirmesi umulmaktadır.

KAYNAKÇA

1. Dennis Lo YM, Corbetta N, Chamberlain PF, Rai V, Sargent IL, Redman CWG, et al. Presence of fetal DNA in maternal plasma and serum. *Lancet*. 1997;350(9076):485–7.
2. Salomon LJ, Sotiriadis A, Wulff CB, Odibo A, Akolekar R. Risk of miscarriage following amniocentesis or chorionic villus sampling: systematic review of literature and updated meta-analysis. *Ultrasound Obstet Gynecol*. 2019;54(4):442–51.
3. Flori E, Doray B, Gautier E, Kohler M, Ernault P, Flori J, et al. Circulating cell-free fetal DNA in maternal serum appears to originate from cyto- and syncytiotrophoblastic cells. Case report. *Hum Reprod*. 2004;19(3):723–4.
4. Lo YMD, Tein MSC, Lau TK, Haines CJ, Leung TN, Poon PMK, et al. Quantitative analysis of fetal DNA in maternal plasma and serum: Implications for noninvasive prenatal diagnosis. *Am J Hum Genet*. 1998;62(4):768–75.
5. Bianchi DW, Flint AF, Pizzimenti MF, Knoll JHM, Latt SA. Isolation of fetal DNA from nucleated erythrocytes in maternal blood. *Proc Natl Acad Sci U S A*. 1990;87(9):3279–83.
6. Lo YMD. Fetal DNA in maternal plasma. *Ann NY Acad Sci*. 2000; 906: 141–7.
7. Sekizawa A, Samura O, Zhen D, Falco V, Farina A, Bianchi DW. Apoptosis in fetal nucleated erythrocytes circulating in maternal blood. *Prenat Diagn*. 2000;20(11):886–9.
8. Sekizawa A, Yokokawa K, Sugito Y, Iwasaki M, Yukimoto Y, Ichizuka K, et al. Evaluation of bidirectional transfer of plasma DNA through placenta. *Hum Genet*. 2003;113(4):307–10.
9. Jackson L. Fetal cells and DNA in maternal blood. *Prenat Diagn*. 2003;23(10):837–46.
10. Alberry M, Maddocks D, Jones M, Abdel Hadi M, Abdel-Fattah S, Avent N, et al. Free fetal DNA in maternal plasma in anembryonic pregnancies: confirmation that the origin is the trophoblast. *Int Soc Prenat Diagnosis*. 2007;27(5):415–8.

11. Sandovici I, Hoelle K, Angiolini E, Constâncio M. Placental adaptations to the maternal-fetal environment: Implications for fetal growth and developmental programming. *Reprod Biomed Online.* 2012;25(1):68–89.
12. Lun FMF, Chiu RWK, Chan KCA, Tak YL, Tze KL, Lo YMD. Microfluidics digital PCR reveals a higher than expected fraction of fetal DNA in maternal plasma. *Clin Chem.* 2008;54(10):1664–72.
13. Illanes S, Denbow M, Kailasam C, Finning K, Soothill PW. Early detection of cell-free fetal DNA in maternal plasma. *Early Hum Dev.* 2007;83(9):563–6.
14. Zhou Y, Zhu Z, Gao Y, Yuan Y, Guo Y, Zhou L, et al. Effects of Maternal and Fetal Characteristics on Cell-Free Fetal DNA Fraction in Maternal Plasma. *Reprod Sci.* 2015;22(11):1429–35.
15. Vora NL, Johnson KL, Basu S, Catalano PM, Hauguel-De Mouzon S, Bianchi DW. A multifactorial relationship exists between total circulating cell-free DNA levels and maternal BMI. Vol. 32, *Prenatal Diagnosis.* 2012. p. 912–4.
16. Attilakos G, Maddocks DG, Davies T, Hunt LP, Avent ND, Soothill PW, et al. Quantification of free fetal DNA in multiple pregnancies and relationship with chorionicity. *Prenat Diagn.* 2011;31(10):967–72.
17. Bischoff FZ, Lewis DE, Simpson JL. Cell-free fetal DNA in maternal blood: Kinetics, source and structure . Vol. 11, *Human Reproduction Update.* 2005. p. 59–67.
18. Chan KA, Zhang J, Hui AB, Wong N, Lau TK, Leung TN, et al. Size distributions of maternal and fetal DNA in maternal plasma. *Clin Chem.* 2004;50(1):88–92.
19. Li Y, Zimmermann B, Rusterholz C, Kang A, Holzgreve W, Hahn S. Size separation of circulatory DNA in maternal plasma permits ready detection of fetal DNA polymorphisms. *Clin Chem.* 2004;50(6):1002–11.
20. Angert RM, LeShane ES, Lo YMD, Chan LYS, Delli-Bovi LC, Bianchi DW. Fetal cell-free plasma DNA concentrations in maternal blood are stable 24 hours after collection: Analysis of first- and third-trimester samples. *Clin Chem.* 2003;49(1):195–8.
21. Hui L, Vaughan JI, Nelson M. Effect of labor on postpartum clearance of cell-free fetal DNA from the maternal circulation. *Prenat Diagn.* 2008;28(4):304–8.
22. Ordoñez E, Rueda L, Cañadas MP, Fuster C, Cirigliano V. Evaluation of Sample Stability and Automated DNA Extraction for Fetal Sex Determination Using Cell-Free Fetal DNA in Maternal Plasma. Kamboh MI, editor. *Biomed Res Int.* 2013;2013
23. Hidestrand M, Stokowski R, Song K, Oliphant A, Deavers J, Goetsch M, et al. Influence of temperature during transportation on cell-free DNA analysis. *Fetal Diagn Ther.* 2012;31(2):122–8.
24. Zhang Y, Li Q, Hui N, Fei M, Hu Z, Sun S. Effect of formaldehyde treatment on the recovery of cell-free fetal DNA from maternal plasma at different processing times. *Clin Chim Acta.* 2008;397(1–2):60–4.
25. Li J, Makrigiorgos GM. COLD-PCR: A new platform for highly improved mutation detection in cancer and genetic testing. *Biochem Soc Trans.* 2009;37(2):427–32.
26. Papageorgiou EA, Fiegler H, Rakyan V, Beck S, Hulten M, Lammissou K, et al. Sites of differential DNA methylation between placenta and peripheral blood: Molecular markers for noninvasive prenatal diagnosis of aneuploidies. *Am J Pathol.* 2009;174(5):1609–18.
27. Tong YK, Jin S, Chiu RWK, Ding C, Chan KCA, Leung TY, et al. Noninvasive prenatal detection of trisomy 21 by an epigenetic-genetic chromosome-dosage approach. *Clin Chem.* 2010;56(1):90–8.
28. Ioannides M, Papageorgiou EA, Keravnou A, Tsaliki E, Spyrou C, Hadjidanuel M, et al. Inter-individual methylation variability in differentially methylated regions between maternal whole blood and first trimester CVS. *Mol Cytogenet.* 2014;7(1):1–8.
29. Pohl G, Shih LM. Principle and applications of digital PCR. *Expert Rev Mol Diagn.* 2004;4(1):41–7.
30. Majumdar N, Banerjee S, Pallas M, Wessel T, Hegerich P. Poisson Plus Quantification for

- Digital PCR Systems. *Sci Rep.* 2017;7(1):1–10.
- 31. Behjati S, Tarpey PS. What is next generation sequencing? *Arch Dis Child Educ Pract Ed.* 2013;98(6):236–8.
 - 32. Mardis ER. Next-generation DNA sequencing methods. *Annu Rev Genomics Hum Genet.* 2008;9:387–402.
 - 33. McCarthy A. Third generation DNA sequencing: Pacific biosciences' single molecule real time technology. *Chem Biol.* 2010;17(7):675–6.
 - 34. Manegold-Brauer G, Hahn S, Lapaire O. What does next-generation sequencing mean for prenatal diagnosis? *Biomark Med.* 2014;8(4):499–508.
 - 35. Skrzypek H, Hui L. Noninvasive prenatal testing for fetal aneuploidy and single gene disorders. *Best Pract Res Clin Obstet Gynaecol.* 2017;42:26–38.
 - 36. Ferrari M, Carrera P, Lampasona V, Galbiati S. New trend in non-invasive prenatal diagnosis. *Clin Chim Acta* 2015;451:9–13.
 - 37. Bianchi DW. From prenatal genomic diagnosis to fetal personalized medicine: Progress and challenges. *Nat Med.* 2012;18(7):1041–51.
 - 38. Lee SY, Kim SJ, Han SH, Park JS, Choi HJ, Ahn JJ, et al. A new approach of digital PCR system for non-invasive prenatal screening of trisomy 21. *Clin Chim Acta [Internet].* 2018;476(November 2017):75–80. Available from: <https://doi.org/10.1016/j.cca.2017.11.015>
 - 39. Norton ME, Jacobsson B, Swamy GK, Laurent LC, Ranzini AC, Brar H, et al. Cell-Free DNA Analysis for Noninvasive Examination of Trisomy. Vol. 70, *Obstetrical and Gynecological Survey.* 2015. p. 483–4.
 - 40. Gil MM, Accurti V, Santacruz B, Plana MN, Nicolaides KH. Analysis of cell-free DNA in maternal blood in screening for aneuploidies: updated meta-analysis. *Ultrasound Obstet Gynecol.* 2017;50(3):302–14.
 - 41. Petersen AK, Cheung SW, Smith JL, Bi W, Ward PA, Peacock S, et al. Positive predictive value estimates for cell-free noninvasive prenatal screening from data of a large referral genetic diagnostic laboratory. *Am J Obstet Gynecol.* 2017;217(6):691.e1–691.e6.
 - 42. Grati FR, Ferreira J, Benn P, Izzi C, Verdi F, Vercellotti E, et al. Outcomes in pregnancies with a confined placental mosaicism and implications for prenatal screening using cell-free DNA. *Genet Med.* 2020;22(2):309–16.
 - 43. Chatron N, Till M, Abel C, Bardel C, Ramond F, Sanlaville D, et al. Detection of rare autosomal trisomies through non-invasive prenatal testing: benefits for pregnancy management. *Ultrasound Obstet Gynecol.* 2019;53(1):129–30.
 - 44. Van Opstal D, Van Maarle MC, Lichtenbelt K, Weiss MM, Schuring-Blom H, Bhola SL, et al. Origin and clinical relevance of chromosomal aberrations other than the common trisomies detected by genome-wide NIPS: Results of the TRIDENT study. *Genet Med.* 2018;20(5):480–5.
 - 45. Pertile MD, Halks-Miller M, Flowers N, Barbacioru C, Kinnings SL, Vavrek D, et al. Rare autosomal trisomies, revealed by maternal plasma DNA sequencing, suggest increased risk of feto-placental disease. *Sci Transl Med.* 2017;9(405).
 - 46. Benn P, Malvestiti F, Grimi B, Maggi F, Simoni G, Grati FR. Rare autosomal trisomies: comparison of detection through cell-free DNA analysis and direct chromosome preparation of chorionic villus samples. *Ultrasound Obstet Gynecol.* 2019;54(4):458–67.
 - 47. Scott F, Bonifacio M, Sandow R, Ellis K, Smet ME, McLennan A. Rare autosomal trisomies: Important and not so rare. *Prenat Diagn.* 2018;38(10):765–71.
 - 48. Reiss RE, Discenza M, Foster J, Dobson L, Wilkins-Haug L. Sex chromosome aneuploidy detection by noninvasive prenatal testing: helpful or hazardous? *Prenat Diagn.* 2017;37(5):515–20.
 - 49. Zhang B, Zhou Q, Chen Y, Shi Y, Zheng F, Liu J, et al. High false-positive non-invasive prenatal screening results for sex chromosome abnormalities: Are maternal factors the culprit? *Prenat Diagn.* 2020;40(4):463–9.
 - 50. Hyett JA, Gardener G, Stojilkovic-Mikic T, Finning KM, Martin PG, Rodeck CH, et al. Re-

- duction in diagnostic and therapeutic interventions by non-invasive determination of fetal sex in early pregnancy. *Prenat Diagn.* 2005;25(12):1111–6.
- 51. Forest MG, Morel Y, David M. Prenatal treatment of congenital adrenal hyperplasia. *Trends Endocrinol Metab.* 1998;9(7):284–9.
 - 52. Zhong XY, Holzgreve W, Hahn S. The levels of circulatory cell free fetal DNA in maternal plasma are elevated prior to the onset of preeclampsia. *Hypertens Pregnancy.* 2002;21(1):77–83.
 - 53. Shah VC, Smart V. Human chromosome Y and SRY. *Cell Biol Int.* 1996;20(1):3–6.
 - 54. Lo YD, Patel P, Sampietro M, Gillmer MDG, Fleming KA, Wainscoat JS. Detection of single-copy fetal DNA sequence from maternal blood. *The Lancet.* 1990;335(8703):1464–5.
 - 55. Stanghellini I, Bertorelli R, Capone L, Mazza V, Neri C, Percesepe A, et al. Quantitation of fetal DNA in maternal serum during the first trimester of pregnancy by the use of a DAZ repetitive probe. *Mol Hum Reprod.* 2006;12(9):587–91.
 - 56. Chim SSC, Tong YK, Chiu RWK, Lau TK, Leung TN, Chan LYS, et al. Detection of the placental epigenetic signature of the maspin gene in maternal plasma. *Proc Natl Acad Sci U S A* 2005;102(41):14753–8.
 - 57. Chan KCA, Ding C, Gerovassili A, Yeung SW, Chiu RWK, Leung TN, et al. Hypermethylated RASSF1A in maternal plasma: A universal fetal DNA marker that improves the reliability of noninvasive prenatal diagnosis. *Clin Chem.* 2006;52(12):2211–8.
 - 58. Bellido ML, Radpour R, Lapaire O, De Bie I, Hösl I, Bitzer J, et al. MALDI-TOF mass array analysis of RASSF1A and SERPINB5 methylation patterns in human placenta and plasma. *Biol Reprod.* 2010;82(4):745–50.
 - 59. Tang NLS, Leung TN, Zhang J, Lau TK, Lo YMD. Detection of fetal-derived paternally inherited X-chromosome polymorphisms in maternal plasma. *Clin Chem.* 1999;45(11):2033–5.
 - 60. Tsui NBY, Kadir RA, Chan KCA, Chi C, Mellars G, Tuddenham EG, et al. Noninvasive prenatal diagnosis of hemophilia by microfluidics digital PCR analysis of maternal plasma DNA. *Blood.* 2011;117(13):3684–91.
 - 61. Jenkins LA, Deans ZC, Lewis C, Allen S. Delivering an accredited non-invasive prenatal diagnosis service for monogenic disorders and recommendations for best practice. *Prenat Diagn.* 2018;38(1):44–51.
 - 62. Hill M, Twiss P, Verhoef TI, Drury S, McKay F, Mason S, et al. Non-invasive prenatal diagnosis for cystic fibrosis: Detection of paternal mutations, exploration of patient preferences and cost analysis. *Prenat Diagn.* 2015;35(10):950–8.
 - 63. Xiong L, Barrett AN, Hua R, Tan TZ, Ho SSY, Chan JKY, et al. Non-invasive prenatal diagnostic testing for β-thalassaemia using cell-free fetal DNA and next generation sequencing. *Prenat Diagn.* 2015;35(3):258–65.
 - 64. Verhoef TI, Hill M, Drury S, Mason S, Jenkins L, Morris S, et al. Non-invasive prenatal diagnosis (NIPD) for single gene disorders: cost analysis of NIPD and invasive testing pathways. *Prenat Diagn.* 2016;36(7):636–42.
 - 65. Traeger-Synodinos J. Real-time PCR for prenatal and preimplantation genetic diagnosis of monogenic diseases. *Mol Aspects Med.* 2006;27(2–3):176–91.
 - 66. Xiong L, Barrett AN, Hua R, Ho SSY, Jun L, Chan KCA, et al. Non-invasive prenatal testing for fetal inheritance of maternal β-thalassaemia mutations using targeted sequencing and relative mutation dosage: a feasibility study. *BJOG An Int J Obstet Gynaecol.* 2018;125(4):461–8.
 - 67. Tsao DS, Silas S, Landry BP, Itzep NP, Nguyen AB, Greenberg S, et al. A novel high-throughput molecular counting method with single base-pair resolution enables accurate single-gene NIPT. *Sci Rep.* 2019;9(1):1–14.
 - 68. Cutts A, Vavoulis D V, Petrou M, Smith F, Clark B, Henderson S, et al. A method for noninvasive prenatal diagnosis of monogenic autosomal recessive disorders. *Blood.* 2019;134(14):1190–3.

69. Lo YMD, Chan KCA, Sun H, Chen EZ, Jiang P, Lun FMF, et al. Maternal plasma DNA sequencing reveals the genome-wide genetic and mutational profile of the fetus. *Sci Transl Med.* 2010;2(61).
70. Vermeulen C, Geeven G, de Wit E, Verstegen MJAM, Jansen RPM, van Kranenburg M, et al. Sensitive Monogenic Noninvasive Prenatal Diagnosis by Targeted Haplotyping. *Am J Hum Genet.* 2017;101(3):326–39.
71. Hui WWI, Jiang P, Tong YK, Lee WS, Cheng YKY, New MI, et al. Universal haplotype-based noninvasive prenatal testing for single gene diseases. Vol. 63, *Clinical Chemistry.* 2017. p. 513–24.
72. Shaw J, Scotchman E, Chandler N, Chitty LS. Non-invasive prenatal testing for aneuploidy, copy-number variants and single-gene disorders. *Reproduction.* 2020;160(5): A1–11.
73. Lo YD, Hjelm NM, Fidler C, Sargent IL, Murphy MF, Chamberlain PF, et al. Prenatal diagnosis of fetal RhD status by molecular analysis of maternal plasma. In: *New England Journal of Medicine.* 1998. p. 1734–8.
74. Clausen FB, Damkjær MB, Dziegieł MH. Noninvasive fetal RhD genotyping. *Transfus Appl Sci* 2014;50(2):154–62.
75. Scheffer PG, Van Der Schoot CE, Page-Christiaens GCML, De Haas M. Noninvasive fetal blood group genotyping of rhesus D, c, e and of K in alloimmunised pregnant women: Evaluation of a 7-year clinical experience. *BJOG An Int J Obstet Gynaecol.* 2011;118(11):1340–8.
76. Chang HE, Hwang SM, Hong YJ, Han M, Park JS, Park KU. Genotyping for RhD and Rh-CEcE Antigens Using Free Circulating Nucleic Acids in Plasma and Serum. *Korean J Blood Transfus.* 2014;25(3):249–59.
77. Tax MGHM, Van der Schoot CE, Van Doorn R, Douglas-Berger L, Van Rhenen DJ, Maaskant-Van Wijk PAM. RHC and RHc genotyping in different ethnic groups. *Transfusion.* 2002;42(5):634–44.
78. Clausen FB. Lessons learned from the implementation of non-invasive fetal RHD screening. *Expert Rev Mol Diagn.* 2018;18(5):423–31.
79. Advani H V, Barrett AN, Evans MI, Choolani M. Challenges in non-invasive prenatal screening for sub-chromosomal copy number variations using cell-free DNA. *Prenat Diagn.* 2017;37(11):1067–75.
80. Grati FR, Molina Gomes D, Ferreira JCPB, Dupont C, Alesi V, Gouas L, et al. Prevalence of recurrent pathogenic microdeletions and microduplications in over 9500 pregnancies. *Prenat Diagn.* 2015;35(8):801–9.
81. Peters D, Chu T, Yatsenko SA, Hendrix N, Hogge WA, Surti U, et al. Noninvasive prenatal diagnosis of a fetal microdeletion syndrome. *N Engl J Med.* 2011;365(19):1847.
82. Devaney SA, Palomaki GE, Scott JA, Bianchi DW. Noninvasive fetal sex determination using cell-free fetal DNA: a systematic review and meta-analysis. *Jama,* 2011;306(6):627–36.
83. Chitty LS, Hudgins L, Norton ME. Current controversies in prenatal diagnosis 2: Cell-free DNA prenatal screening should be used to identify all chromosome abnormalities. *Prenat Diagn.* 2018;38(3):160–5.
84. Helgeson J, Wardrop J, Boomer T, Almasri E, Paxton WB, Saldivar JS, et al. Clinical outcome of subchromosomal events detected by whole-genome noninvasive prenatal testing. *Prenat Diagn.* 2015;35(10):999–1004.
85. Grati FR, Gross SJ. Noninvasive screening by cell-free DNA for 22q11.2 deletion: Benefits, limitations, and challenges. *Prenat Diagn.* 2019;39(2):70–80.
86. Sun K, Jiang P, Wong AIC, Cheng YKY, Cheng SH, Zhang H, et al. Size-tagged preferred ends in maternal plasma DNA shed light on the production mechanism and show utility in noninvasive prenatal testing. *Proc Natl Acad Sci U S A.* 2018;115(22):E5106–14.
87. Canick JA, Palomaki GE, Kloza EM, Lambert-Messerlian GM, Haddow JE. The impact of maternal plasma DNA fetal fraction on next generation sequencing tests for common fetal aneuploidies,. *Prenat Diagn.* 2013;(33):667–674.

88. Yao H, Zhang L, Zhang H, Jiang F, Hu H, Chen F, et al. Noninvasive prenatal genetic testing for fetal aneuploidy detects maternal trisomy X. *Prenat Diagn.* 2012;32(11):1114–6.
89. Hartwig TS, Ambye L, Sørensen S, Jørgensen FS. Discordant non-invasive prenatal testing (NIPT)—a systematic review. *Prenat Diagn.* 2017;37(6):527–39.
90. Bianchi DW, Wilkins-Haug LE, Enders AC, Hay ED. Origin of extraembryonic mesoderm in experimental animals: Relevance to chorionic mosaicism in humans. *Am J Med Genet.* 1993;(46):542–550.
91. Curnow KJ, Wilkins-Haug L, Ryan A, Kirkizlar E, Stosic M, Hall MP, et al. Detection of triploid, molar, and vanishing twin pregnancies by a single-nucleotide polymorphism-based noninvasive prenatal test. *Am J Obstet Gynecol.* 2015;212(1):79–81.
92. Osborne CM, Hardisty E, Devers P, K. K, Hayden MA, Goodnight W, et al. Discordant noninvasive prenatal testing results in a patient subsequently diagnosed with metastatic disease. *Prenat Diagn.* 2013;33(6):609–11.
93. Zhang J, Li J, Saucier JB, Feng Y, Jiang Y, Sinson J, et al. Non-invasive prenatal sequencing for multiple Mendelian monogenic disorders using circulating cell-free fetal DNA. *Nat Med.* 2019;25(3):439–47.
94. Hudecova I, Jiang P, Davies J, Lo YMD, Kadir RA, Chiu RWK. Noninvasive detection of F8 int22h-related inversions and sequence variants in maternal plasma of hemophilia carriers. *Blood.* 2017;130(3):340–7.
95. New MI, Tong YK, Yuen T, Jiang P, Pina C, Chan KA, et al. Noninvasive prenatal diagnosis of congenital adrenal hyperplasia using cell-free fetal DNA in maternal plasma. *J Clin Endocrinol Metab.* 2014;99(6): E1022–30.
96. Chen M, Chen C, Huang X, Sun J, Jiang L, Li Y, et al. Noninvasive prenatal diagnosis for Duchenne muscular dystrophy based on the direct haplotype phasing. *Prenat Diagn.* 2020;40(8):918–24.
97. Parks M, Court S, Bowns B, Cleary S, Clokie S, Hewitt J, et al. Non-invasive prenatal diagnosis of spinal muscular atrophy by relative haplotype dosage. *Eur J Hum Genet.* 2017;25(4):416–22.
98. Chandler NJ, Ahlfors H, Drury S, Mellis R, Hill M, McKay FJ, et al. Noninvasive prenatal diagnosis for cystic fibrosis: Implementation, uptake, outcome, and implications. *Clin Chem.* 2020;66(1):207–16.
99. Perlado S, Bustamante-Aragonés A, Donas M, Lorda-Sánchez I, Plaza J, De Alba MR. Fetal genotyping in maternal blood by digital PCR: Towards NIPD of monogenic disorders independently of parental origin. *PLoS One.* 2016;11(4).
100. Camunas-Soler J, Lee H, Hudgins L, Hintz SR, Blumenfeld YJ, El-Sayed YY, et al. Noninvasive Prenatal Diagnosis of Single-Gene Disorders by Use of Droplet Digital PCR. 2017