

# BÖLÜM 97

## SEREBELLUM VE BEYİN SAPININ HEREDODEJENERATİF HASTALIKLARI

Elif ACAR ARSLAN<sup>1</sup>

### HEREDİTER ATAKSİLER

#### Giriş

Ataksi, serebellum ve yollarının bozukluğunun sonucu olarak motor inkoordinasyon sonucunda oluşur. <sup>1</sup> Ataksiler ya, doğuştan metabolik bozukluklardan veya doğuştan metabolik hastalık kaynaklı olmayan, ilerleyici dejeneratif ataksi nedeniyle oluşur. İlerleyici dejeneratif ataksiler, otozomal dominant, otozomal resesif, X'e bağlı ve mitokondrial hastalıklar sonucu olan ataksilerdir.

**Otozomal Dominant Ataksiler:** Bu grupta otozomal dominant ataksiler grubundan olan spinoserebellar ataksiler bulunmaktadır. Bugüne kadar 40'dan fazla tipi tanımlanmıştır. Bir kısmı tirnükleotid tekrar sayısından kaynaklanır. Bu tekrarlar gerek somatik gerek ise germline hücrelerde kararsızlığa yol açarak, hastalık oluştururlar. Bir sonraki jenerasyonlarda 'anti-sipasyon' özelliği nedeniyle, hastalığın daha erken yaşlarda olan, klinik yansımalarına neden olabilirler. Çocukluk yaş grubundan ziyade, daha çok erişkin yaş grubunda görülmektedir. Bu nedenle, bu yazı daha çok otozomal resesif ataksiler üzerine yoğunlaşmıştır.

**Otozomal Resesif Ataksiler:** Çocukluk çağı başlangıçlı olup, en sık bilinen alt grupları, Friedreich ataksisi, ataksi-telanjektazi, ataksi okulomotor apraksi tip 1 ve tip 2'dir. Dört otozomal resesif ataksi, E vitamin eksikliği ile beraber olan ataksi, Refsum hastalığı, Co enzim Q10 eksikliğine bağlı ataksi ve serebrotendinömatöz ksantamatozis de otozomal resesif hastalıklar olup, tedavi şansları olan grupta oldukları için atlanılmaması gereken hastalıklardır.<sup>2</sup> Herediter atakilere genellikle serebellar atrofi eşlik eder. Bu grupta en sık olarak Friedreich ataksisi görülmektedir.

#### Friedreich Ataksisi

Herediter ataksiler, heterojen bir grup hastalık olup, en sık görüleni, Friedreich ataksisi olarak bilinmektedir. Frataxin geni 9. kromozomun uzun kolunda kodlanmaktadır. Hastaların çoğunda, frataxin geninin her iki alelinin intron 1'de GAA tekrar sayısının "ekspansiyonu" görülmektedir. GAA tekrarları, 66-100 arasındadır. <sup>3,4</sup> Normal alelde bu sayı genellikle 7-34 arasındadır.

Friedreich ataksisinin kliniğe yansımada, GAA "ekspansiyonun" sayısına göre değişmektedir. Büyük GAA "ekspansiyonları",

<sup>1</sup> Doç. Dr., Karadeniz Teknik Üniversitesi Tıp Fakültesi, Çocuk Nörolojisi Kliniği, elifacararslan@gmail.com

**Epizodik ataksi tip 6:** SLC1A3 genindeki heterozigot mutasyondan kaynaklanır. Epizodik ataksi, migren, alternan hemipleji, nöbet gibi klinik olabilir.<sup>49</sup>

**Epizodik ataksi tip 7** ise oldukça nadirdir.

## TEDAVİ EDİLEBİLİR ATAKSİLER

**Vitamin E eksikliği ile giden ataksi:** Otozomal resesif olup, alfa tokoferol transfer protein genindeki patolojiden kaynaklanır. Yavaş ve ilerleyici ataksi, nöropati ile Friedreich ataksisine benzer. Bazı olgularda, retinitis pigmentosa da görülmektedir.<sup>50,51</sup> Malabsorbsiyona yol açan hastalıklarda da görülebilir. Yüksek doz E vitamini, nörolojik bulgulara düzelmeye sağlar.

**Serebrotendinöz ksantamatozis:** Otozomal resesif, ilerleyici nadir bir hastalıktır. Mitokondrial sterol 27 hidrosilaz genindeki, safra asidi sentezindeki bloğa neden olan bir mutasyondan kaynaklanır. Ataksi, nöropati, katarakt, aşil tendondaki ksantomlar, ateroskleroz ile karakterizedir. Kolestanol, hastalığı, nörolojik toksisiteden sorumludur.

**Refsum hastalığı:** Refsum hastalığı, otozomal resesif ataksi grubunda olup, ihtiyozis, retinitis pigmentosa, nöropati ve ataksi ile karakterizedir. PHYH genindeki, mutasyondan kaynaklanır. Fitanik asidin yıkılmasındaki sorundan kaynaklanır. Diyetle, fitanik asidin alımındaki kısıtlama, tedavide kullanılır.

**Özet olarak,** herediter ataksiler, otozomal dominant, otozomal resesif, X'e bağlı, mitokondrial ataksiler şeklinde sınıflandırılabilir. Otozomal dominant ataksiler, çocuklarda görece nadir olup, "antisipasyon" özelliği, dolaşımı ile de, bir sonraki jenerasyonlarda, daha erken yaşlarda görülebilir. Otozomal resesif ataksilerinde, en sık görüleni, Friedreich ataksisi'dir. Derin tendon reflekslerinin alınmadığı olgularda, periferik nöropati, ekstremiteler ve yürüyüş ataksisi varlığında, kardiyolojik incelemede hipertrofik kardiyomyopati sap-

tanmasında, tanı, Friedreich ataksisi lehine değerlendirilmelidir. Frataxin geninde GAA üçlü tekrar "ekspansiyonu" ile olgunun genetik tanısı konulur.<sup>52-53</sup>

## KAYNAKLAR

1. Brusse E, Maat-Kievit JA, van Swieten JC. Diagnosis and management of early- and late-onset cerebellar ataxia. *Clin Genet.* 2007; 71:12.
2. Jayadev, S., Bird, T. Hereditary ataxias: overview. *Genet Med.* 2013;15, 673–683.
3. Dürr A, Cossee M, Agid Y, et al. Clinical and genetic abnormalities in patients with Friedreich's ataxia. *N Engl J Med.* 1996; 335:1169.
4. Epplen C, Epplen JT, Frank G, et al. Differential stability of the (GAA)<sub>n</sub> tract in the Friedreich ataxia (STM7) gene. *Hum Genet.* 1997; 99:834.
5. Cossée M, Schmitt M, Campuzano V, et al. Evolution of the Friedreich's ataxia trinucleotide repeat expansion: founder effect and premutations. *Proc Natl Acad Sci U S A.* 1997; 94:7452.
6. Filla A, De Michele G, Cavalcanti F, et al. The relationship between trinucleotide (GAA) repeat length and clinical features in Friedreich ataxia. *Am J Hum Genet.* 1996; 59:554.
7. Harding AE. Friedreich's ataxia: a clinical and genetic study of 90 families with an analysis of early diagnostic criteria and intrafamilial clustering of clinical features. *Brain.* 1981; 104:589.
8. Koeppen AH. Friedreich's ataxia: pathology, pathogenesis, and molecular genetics. *J Neurol Sci.* 2011; 303:1.
9. Schulz JB, Dehmer T, Schöls L, et al. Oxidative stress in patients with Friedreich ataxia. *Neurology* 2000; 55:1719.
10. Sherer T, Greenamyre JT. A therapeutic target and biomarker in Friedreich's ataxia. *Neurology* 2000; 55:1600.
11. Campuzano V, Montermini L, Lutz Y, et al. Frataxin is reduced in Friedreich ataxia patients and is associated with mitochondrial membranes. *Hum Mol Genet.* 1997; 6:1771.
12. Priller J, Scherzer CR, Faber PW, et al. Frataxin gene of Friedreich's ataxia is targeted to mitochondria. *Ann Neurol.* 1997; 42:265.
13. Babcock M, de Silva D, Oaks R, et al. Regulation of mitochondrial iron accumulation by Yfh1p, a putative homolog of frataxin. *Science.* 1997; 276:1709.
14. Tan G, Chen LS, Lonnerdal B, et al. Frataxin expression rescues mitochondrial dysfunctions in FRDA cells. *Hum Mol Genet.* 2001; 10:2099.
15. Beal MF. Mitochondria take center stage in aging and neurodegeneration. *Ann Neurol* 2005; 58:495.
16. Delatycki MB, Corben LA. Clinical features of Friedreich ataxia. *J Child Neurol* 2012; 27:1133.
17. Collins A. Clinical neurogenetics: friedreich ataxia. *Neurol Clin.* 2013; 31:1095.

18. Ribai P, Pousset F, Tanguy ML, et al. Neurological, cardiological, and oculomotor progression in 104 patients with Friedreich ataxia during long-term follow-up. *Arch Neurol.* 2007; 64:558.
19. Delatycki MB, Paris DB, Gardner RJ, et al. Clinical and genetic study of Friedreich ataxia in an Australian population. *Am J Med Genet.* 1999; 87:168.
20. Wüllner U, Klockgether T, Petersen D, et al. Magnetic resonance imaging in hereditary and idiopathic ataxia. *Neurology.* 1993; 43:318.
21. Schulz JB, Boesch S, Bürk K, et al. Diagnosis and treatment of Friedreich ataxia: a European perspective. *Nat Rev Neurol.* 2009; 5:222.
22. Child JS, Perloff JK, Bach PM, et al. Cardiac involvement in Friedreich's ataxia: a clinical study of 75 patients. *J Am Coll Cardiol.* 1986; 7:1370.
23. Payne RM, Wagner GR. Cardiomyopathy in Friedreich ataxia: clinical findings and research. *J Child Neurol.* 2012; 27:1179.
24. Mejia E, Lynch A, Hearle P, et al. Ectopic Burden via Holter Monitors in Friedreich Ataxia. *Pediatr Neurol.* 2021; 117:29.
25. Lodi R, Hart PE, Rajagopalan B, et al. Antioxidant treatment improves in vivo cardiac and skeletal muscle bioenergetics in patients with Friedreich's ataxia. *Ann Neurol.* 2001; 49:590.
26. Hart PE, Lodi R, Rajagopalan B, et al. Antioxidant treatment of patients with Friedreich ataxia: four-year follow-up. *Arch Neurol.* 2005; 62:621.
27. Pandolfo M, Arpa J, Delatycki MB, et al. Deferiprone in Friedreich ataxia: a 6-month randomized controlled trial. *Ann Neurol.* 2014; 76:509.
28. Fogel BL, Perlman S. Clinical features and molecular genetics of autosomal recessive cerebellar ataxias. *Lancet Neurol.* 2007; 6:245.
29. Lutz R, Bodensteiner J, Schaefer B, Gay C. X-linked olivopontocerebellar atrophy. *Clin Genet.* 1989; 35:417.
30. Spira PJ, McLeod JG, Evans WA. A spinocerebellar degeneration with X-linked inheritance. *Brain.* 1979; 102:27.
31. Kremer H, Hamel BC, van den Helm B, et al. Localization of the gene (or genes) for a syndrome with X-linked mental retardation, ataxia, weakness, hearing impairment, loss of vision and a fatal course in early childhood. *Hum Genet.* 1996; 98:513.
32. Synofzik M, Puccio H, Mochel F, Schöls L. Autosomal Recessive Cerebellar Ataxias: Paving the Way toward Targeted Molecular Therapies. *Neuron.* 2019; 101:560.
33. Batshaw ML, Msall M, Beaudet AL, Trojak J. Risk of serious illness in heterozygotes for ornithine transcarbamylase deficiency. *J Pediatr.* 1986; 108:236.
34. Yudkoff M, Daikhin Y, Nissim I, et al. In vivo nitrogen metabolism in ornithine transcarbamylase deficiency. *J Clin Invest.* 1996; 98:2167.
35. Gaspari R, Arcangeli A, Mensi S, et al. Late-onset presentation of ornithine transcarbamylase deficiency in a young woman with hyperammonemic coma. *Ann Emerg Med.* 2003; 41:104.
36. Seow HF, Bröer S, Bröer A, et al. Hartnup disorder is caused by mutations in the gene encoding the neutral amino acid transporter SLC6A19. *Nat Genet.* 2004; 36:1003.
37. Hasan SM, D'Adamo MC. GeneReviews®, Adam MP, Ardinger HH, Pagon RA, et al. (Eds), University of Washington, Seattle, Seattle (WA) 1993. [www.ncbi.nlm.nih.gov/books/NBK25442/](http://www.ncbi.nlm.nih.gov/books/NBK25442/) (December 11, 2018).
38. Steckley JL, Ebers GC, Cader MZ, McLachlan RS. An autosomal dominant disorder with episodic ataxia, vertigo, and tinnitus. *Neurology.* 2001; 57:1499.
39. Tomlinson SE, Rajakulendran S, Tan SV, et al. Clinical, genetic, neurophysiological and functional study of new mutations in episodic ataxia type 1. *J Neurol Neurosurg Psychiatry.* 2013; 84:1107.
40. Graves TD, Cha YH, Hahn AF, et al. Episodic ataxia type 1: clinical characterization, quality of life and genotype-phenotype correlation. *Brain.* 2014; 137:1009.
41. Scheffer H, Brunt ER, Mol GJ, et al. Three novel KCNA1 mutations in episodic ataxia type I families. *Hum Genet.* 1998; 102:464.
42. Tacik P, Guthrie KJ, Strongosky AJ, et al. Whole-exome sequencing as a diagnostic tool in a family with episodic ataxia type 1. *Mayo Clin Proc.* 2015; 90:366.
43. Zsorin NL, Baloh RW, Myers LB. Acetazolamide-responsive episodic ataxia syndrome. *Neurology.* 1983; 33:1212.
44. Spacey S. Episodic ataxia type 2. In: *GeneReviews* 2011.
45. Yue Q, Jen JC, Thwe MM, et al. De novo mutation in CACNA1A caused acetazolamide-responsive episodic ataxia. *Am J Med Genet.* 1998; 77:298.
46. Cader MZ, Steckley JL, Dymont DA, et al. A genome-wide screen and linkage mapping for a large pedigree with episodic ataxia. *Neurology.* 2005; 65:156.
47. Herrmann A, Braathen GJ, Russell MB. [Episodic ataxias]. *Tidsskr Nor Laegeforen.* 2005; 125:2005.
48. Kono N, Ohto U, Hiramatsu T, et al. Impaired  $\alpha$ -TTP-PIPs interaction underlies familial vitamin E deficiency. *Science.* 2013; 340:1106.
49. Yokota T, Shiojiri T, Gotoda T, et al. Friedreich-like ataxia with retinitis pigmentosa caused by the His101Gln mutation of the alpha-tocopherol transfer protein gene. *Ann Neurol* 1997; 41:826.
50. <https://www.uptodate.com/contents/overview-of-the-hereditary-ataxias>.
51. <https://www.uptodate.com/contents/friedreich-ataxia>.

olabilir. Saf ve komplike HSP olarak iki grupta olabilir. Pigmenter retinopati açısından, göz konsültasyonu önemlidir. Otozomal dominant HSP'lerin en sık bilinenleri HSP4 ve HSP3'ürdür. Otozomal dominant HSP'ler, genellikle saf HSP'ye neden olurlar. Tedavide, fizik tedavi ve rehabilitasyon, iş-uğraşı terapisi önemlidir. Germe egzersizleri, spastisiteyi azaltmada ve denge ve güç kazanımı açısından önerilir.<sup>13</sup>

## KAYNAKLAR

1. Fink JK. Hereditary spastic paraplegia: clinico-pathologic features and emerging molecular mechanisms. *Acta Neuropathol.* 2013; 126:307.
2. Denora PS, Santorelli FM, Bertini E. Hereditary spastic paraplegias: one disease for many genes, and still counting. *Handb Clin Neurol.* 2013; 113:1899.
3. Coutinho P, Ruano L, Loureiro JL, et al. Hereditary ataxia and spastic paraplegia in Portugal: a population-based prevalence study. *JAMA Neurol.* 2013; 70:746.
4. Loureiro JL, Brandão E, Ruano L, et al. Autosomal dominant spastic paraplegias: a review of 89 families resulting from a portuguese survey. *JAMA Neurol.* 2013; 70:481.
5. McDermott CJ, Burness CE, Kirby J, et al. Clinical features of hereditary spastic paraplegia due to spastin mutation. *Neurology.* 2006; 67:45.
6. Depienne C, Stevanin G, Brice A, Durr A. Hereditary spastic paraplegias: an update. *Curr Opin Neurol.* 2007; 20:674.
7. Züchner S. The genetics of hereditary spastic paraplegia and implications for drug therapy. *Expert Opin Pharmacother.* 2007; 8:1433.
8. Zhao X, Alvarado D, Rainier S, et al. Mutations in a newly identified GTPase gene cause autosomal dominant hereditary spastic paraplegia. *Nat Genet.* 2001; 29:326.
9. Warnecke T, Duning T, Schirmacher A, et al. A novel splice site mutation in the SPG7 gene causing widespread fiber damage in homozygous and heterozygous subjects. *Mov Disord.* 2010; 25:413.
10. Klebe S, Depienne C, Gerber S, et al. Spastic paraplegia gene 7 in patients with spasticity and/or optic neuropathy. *Brain.* 2012; 135:2980.
11. van Gassen KL, van der Heijden CD, de Bot ST, et al. Genotype-phenotype correlations in spastic paraplegia type 7: a study in a large Dutch cohort. *Brain.* 2012; 135:2994.
12. Orthmann-Murphy JL, Salsano E, Abrams CK, et al. Hereditary spastic paraplegia is a novel phenotype for GJA12/GJC2 mutations. *Brain.* 2009; 132:426.
13. <https://www.uptodate.com/contents/hereditary-spastic-paraplegia>

hidrosefali, jeneralize beyaz cevher değişiklikleri ve diffüz kortikal anormallikler görülür.<sup>41</sup> Diğer nedenleri arasında ise, CASK ilişkili pontoserebellar hipoplazi, pontin tegmental 'cap' displazisi, serebellar agenezi ve prematüriteye sekonder hasar ile olan serebellar yıkım gelmektedir.<sup>1</sup>

Özet olarak; Serebellar hipoplazi, şekli normal olan serebellumdaki serebellar volümün azalmasına verilen isimdir.<sup>2,3</sup> Serebellar hipoplazi, diğer beyin malformasyonları ile birlikte olabilir. Çocuklar, genellikle, global gelişim geriliği ve hipotoni ile kliniğe gelirler.<sup>4</sup> Serebellar bulgular, sonradan gelişir. Nörolojik bulgular arasında, trunkal ataksi, hipotoni, oküler hareketteki bozukluklar, disartri, intansiyonel tremor ve mikrosefali gelir. Genetik nedenler yanı sıra, konjenital enfeksiyonlar, metabolik bozukluklar, kromozomal anormallikler ile beraber görülebilir. Santral sinir sistemi gelişim malformasyonları, konjenital glikozilasyon defektleri, alfa distroglikanopatiler ile birlikte görülebilir. Diğer beyin sapı hipoplazileri ile beraber görülebilceği gibi, Dandy-Walker malformasyonları gibi sendromların komponentleri şeklinde de olabilir.

## KAYNAKLAR

1. Poretti A, Boltshauser E, Doherty D. Cerebellar hypoplasia: differential diagnosis and diagnostic approach. *Am J Med Genet C Semin Med Genet.* 2014;166C(2):211-26.
2. Boltshauser E. Cerebellum-small brain but large confusion: A review of selected cerebellar malformations and disruptions. *Am J Med Genet A.* 2004;126A:376-385.
3. Boltshauser E, Poretti A. Nonprogressive congenital ataxia. In: Boltshauser E, Schmähmann JD, editors. *Cerebellar disorders in children.* London: Mac Keith Press. 2012;135-139.
4. Bolduc ME, Limperopoulos C. Neurodevelopmental outcomes in children with cerebellar malformations: A systematic review. *Dev Med Child Neurol.* 2009;51: 256-267.
5. Wassmer E, Davies P, Whitehouse WP, Green SH. Clinical spectrum associated with cerebellar hypoplasia. *Pediatr Neurol.* 2003;28: 347-351.
6. Ventura P, Presicci A, Perniola T, Campa MG, Margari L. Mental retardation and epilepsy in patients with isolated cerebellar hypoplasia. *J Child Neurol.* 2006. 21:776-781.
7. Schmähmann JD, Sherman JC. The cerebellar cognitive affective syndrome. *Brain.* 1998. 121:561-579.
8. Poretti A, Meoded A, Rossi A, Raybaud C, Huisman TA. Diffusion tensor imaging and fiber tractography in brain malformations. *Pediatr Radiol.* 2013; 43:28.
9. Bosemani T, Poretti A, Huisman TA. Susceptibility-weighted imaging in pediatric neuroimaging. *J Magn Reson Imaging.* 2014;40(3):530-44.
10. Poretti A, Wolf NI, Boltshauser E. Differential diagnosis of cerebellar atrophy in childhood. *Eur J Paediatr Neurol.* 2008; 12:155- 167.
11. Vermeulen RJ, Peeters-Scholte C, Van Vugt JJ, et al. Fetal origin of brain damage in 2 infants with a CO-L4A1 mutation: Fetal and neonatal MRI. *Neuropediatrics.* 2011;42:1-3.
12. Oza VS, Wang E, Berenstein A, et al. PHACES association: A neuroradiologic review of 17 patients. *AJNR Am J Neuroradiol.* 2008;29:807-813.
13. Hess CP, Fullerton HJ, Metry DW, et al. Cervical and intracranial arterial anomalies in 70 patients with PHACE syndrome. *AJNR Am J Neuroradiol.* 2010;31: 1980-1986.
14. Laine CM, Joeng KS, Campeau PM, et al. WNT1 mutations in early-onset osteoporosis and osteogenesis imperfecta. *N Engl J Med.* 2013;368:1809-1816.
15. Pyott SM, Tran TT, Leistriz DF, et al. WNT1 mutations in families affected by moderately severe and progressive recessive osteogenesis imperfecta. *Am J Hum Genet.* 2013;92:590-597.
16. Poretti A, Huisman TA, Scheer I, Boltshauser E. Joubert syndrome and related disorders: Spectrum of neuroimaging findings in 75 patients. *AJNR Am J Neuroradiol.* 2011;32:1459-1463.
17. Parisi MA, Dobyns WB. Human malformations of the midbrain and hindbrain: Review and proposal classification scheme. *Mol Genet Metab.* 2003;80:36-53.
18. Liao C, Fu F, Li R, Yang X, Xu Q, Li D. Prenatal diagnosis and molecular characterization of a novel locus for Dandy-Walker malformation on chromosome 7p21.3q. *Eur J Med Genet.* 2012. 55:472-475.
19. Doherty D. Joubert syndrome: Insights into brain development, cilium biology, and complex disease. *Semin Pediatr Neurol.* 2009;16: 143-154.
20. Romani M, Micalizzi A, Valente EM. Joubert syndrome: Congenital cerebellar ataxia with the molar tooth. *Lancet Neurol.* 2013; 12:894-905.
21. Halbritter J, Bizet AA, Schmidts M, Porath JD, Braun DA, Gee HY et al. Defects in the IFT-B component IFT172 cause Jeune and Mainzer-Saldino syndromes in humans. *Am J Hum Genet.* 2013. 93:915-925.
22. Thomas S, Wright KJ, Le Corre S, et al. A homozygous PDE6D mutation in Joubert syndrome impairs targeting of farnesylated INPP5E protein to the primary cilium. *Hum Mutat.* 2014; 35:137-146.
23. Tuz K, Bachmann-Gagescu R, O'Day DR, et al. Mu-



- tations in CSPP1 cause primary cilia abnormalities and Joubert syndrome with or without Jeune asphyxiating thoracic dystrophy. *Am J Hum Genet.* 2014; 94:62–72.
24. Poretti A, Alber FD, Burki S, Toelle SP, Boltshauser E. Cognitive outcome in children with rhombencephalosynapsis. *Eur J Paediatr Neurol.* 2009; 13:28–33.
  25. Ishak GE, Dempsey JC, Shaw DW, et al. Rhombencephalosynapsis: A hindbrain malformation associated with incomplete separation of midbrain and forebrain, hydrocephalus and a broad spectrum of severity. *Brain.* 2012; 135:1370–1386.
  26. Steinlin ML, Nadal D, Eich GF, Martin E, Boltshauser EJ. Late intrauterine cytomegalovirus infection: Clinical and neuroimaging findings. *Pediatr Neurol.* 1996; 15: 249–253.
  27. Rosati P, Guariglia L. Cerebellar hypoplasia: could it be a sonographic finding of abnormal fetal karyotype in early pregnancy? *Fetal Diagn Ther.* 1999; 14:365–367.
  28. Lin HY, Lin SP, Chen YJ, et al. Clinical characteristics and survival of trisomy 18 in a medical center in Taipei, 1988– 2004. *Am J Med Genet A.* 2006; 140:945–951.
  29. Ulgiati F, Nicita F, Papetti L, et al. Posterior fossa malformations and sex chromosomes anomalies. Report of a case with XYY syndrome and overview of known associations. *Eur J Pediatr.* 2013; 172:1267–1270.
  30. Leonardi ML, Pai GS, Wilkes B, Lebel RR. Ritscher-Schinzel cranio-cerebello-cardiac (3C) syndrome: Report of four new cases and review. *Am J Med Genet.* 2001; 102:237–242.
  31. Elliott AM, Simard LR, Coghlan G, Chudley AE, Chodirker BN, Greenberg CR et al. A novel mutation in KLAA0196: Identification of a gene involved in Ritscher–Schinzel/3C syndrome in a First Nations cohort. *J Med Genet.* 2013; 50:819–822.
  32. Sznajer Y, Baumann C, David A, et al. Further delineation of the congenital form of X-linked dyskeratosis congenita (Hoyeraal-Hreidarsson syndrome). *Eur J Pediatr.* 2003; 162:863–868.
  33. Ozgen HM, Overweg-Plandsoen WC, Blee-Pelk J, Besselaar PP, Hennekam RC. Cerebellar hypoplasia-endosteal sclerosis: A long term follow-up. *Am J Med Genet A.* 2005;134A:215–219.
  34. Pisano T, Barkovich AJ, Leventer RJ, et al. Peritrigonal and temporo-occipital heterotopia with corpus callosum and cerebellar dysgenesis. *Neurology.* 2012. 79:1244–1251.
  35. Adachi Y, Mochida G, Walsh C, Barkovich J. Posterior fossa in primary microcephaly: Relationships between forebrain and mid-hindbrain size in 110 patients. *Neuropediatrics.* 2014; 45:93–101.
  36. Boycott KM, Flavelle S, Bureau A, et al. Homozygous deletion of the very low density lipoprotein receptor gene causes autosomal recessive cerebellar hypoplasia with cerebral gyral simplification. *Am J Hum Genet.* 2005; 77:477–483.
  37. Glass HC, Boycott KM, Adams C, et al. Autosomal recessive cerebellar hypoplasia in the Hutterite population. *Dev Med Child Neurol.* 2005;47: 691–695.
  38. Feraco P, Mirabelli-Badenier M, Severino M, et al. The shrunken, bright cerebellum: A characteristic MRI finding in congenital disorders of glycosylation type 1a. *AJNR Am J Neuroradiol.* 2012; 33:2062–2067.
  39. Freeze HH, Chong JX, Bamshad MJ, Ng BG. Solving glycosylation disorders: Fundamental approaches reveal complicated pathways. *Am J Hum Genet.* 2014; 94:161–175.
  40. Bonnemann CG, Wang CH, Quijano-Roy S, et al. Diagnostic approach to the congenital muscular dystrophies. *Neuromuscul Disord.* 2014; 24:289–311.
  41. Clement E, Mercuri E, Godfrey C, et al. Brain involvement in muscular dystrophies with defective dystroglycan glycosylation. *Ann Neurol.* 2008; 64:573–582.

**PCH11:** Yedi aile bildirilmiştir.<sup>11,12</sup> TBC1D23 geni sorumludur. PCH11 de, tıpkı PCH8 gibi, dejeneratif olmayan formudur. Ciddi gelişimsel gerilik, mikrosefali ve hipotonisite ile karakterizedir. Bazı olgular, bağımsız yürüme yetisini de kazanmışlardır. Olguların yarısında, ataksi gibi serebellar bulgular tanımlanmıştır. Beyin MRG'de, ponsda, ilerleyici olmayan hipoplazi ve korpus kallosum hipoplazisi ve serebellum hipoplazisi ile karakterizedir.<sup>1,11,12</sup>

**Özet olarak,** pontoserebellar hipoplaziler, serebellum ve ponsu ilgilendiren bir grup nörodejeneratif hastalıktır.<sup>1,2</sup> Kalıtları, otozomal resesiftir. Pontoserebellar hipoplazi tip 1, bulbar ve spinal motor nöronlarda dejenerasyon ile karakterize olup, bu yönüyle spinal musküler atrofiye benzer. PCH2, ise jeneralize klonus, yutma bozuklukları, kore, distoni ve ilerleyici mikrosefali ile karakterizedir. PCH4, PCH2'nin daha ciddi olan klinik formu olup, uzamış neonatal klonus, hipertonisite, primer hipoventilasyon, polihidramnios ve konjenital kontraktürler gibi şiddetli klinik bulgular ile gider. PCH'ların günümüze kadar 11 alt tipi tanımlanmıştır.

## KAYNAKLAR

1. van Dijk, T., Baas, F., Barth, P.G. et al. What's new in pontocerebellar hypoplasia? An update on genes and subtypes. *Orphanet J Rare Dis.* 2018;13, 92.
2. Brun R. Zur Kenntnis der Bildungsfehler des Kleinhirns. *Epikritische Bemerkungen zur Entwicklungspathologie, Morphologie und Klinik der Umschriebenen Entwicklungshemmungen des Neocerebellums.* *Schweiz Arch Neurol Psychiatr.* 1917;1:48-105.
3. Goutieres F, Aicardi J, Farkas E. Anterior horn cell disease associated with pontocerebellar hypoplasia in infants. *J Neurol Neurosurg Psychiatry.* 1977; 40:370-378.,
4. Barth PG. Pontocerebellar hypoplasias. An overview of a group of inherited neurodegenerative disorders with fetal onset. *Brain Dev.* 1993;15(6):411-22.
5. Eggers VR, Barth PG, J-MF N, et al. EXOSC3 mutations in pontocerebellar hypoplasia type 1: novel mutations and genotype-phenotype correlations. *Orphanet J Rare Dis.* 2014; 9:23.
6. Rudnik-Schöneborn S, Senderek J, Jen JC, et al. Pontocerebellar hypoplasia type 1: clinical spectrum and relevance of EXOSC3 mutations. *Neurology.* 2013;80(5):438-46.
7. Edvardson S, Shaag A, Kolesnikova O, et al. Deleterious mutation in the mitochondrial Arginyl-transfer RNA Synthetase gene is associated with pontocerebellar hypoplasia. *Am J Hum Genet.* 2007;81(4):857-862.
8. Mochida GH, Ganesh VS, de Michelena MI, et al. CHMP1A encodes an essential regulator of BMI1-INK4A in cerebellar development. *Nat Genet.* 2012;44(11):1260-1264.
9. Schaffer AE, Eggers VRC, Caglayan AO, et al. CLP1 founder mutation links tRNA splicing and maturation to cerebellar development and neurodegeneration. *Cell.* 2014;157(3):651-653.
10. Karaca E, Weitzer S, Pehlivan D, et al. Human CLP1 mutations alter tRNA biogenesis, affecting both peripheral and central nervous system function. *Cell.* 2014;157(3):636-650.
11. Ivanova EL, Mau-Them FT, Riazuddin S, et al. Homozygous truncating variants in TBC1D23 cause pontocerebellar hypoplasia and Alter cortical development. *Am J Hum Genet.* 2017;101(3):428-440.
12. Grosso S, Mostadini R, Cioni M, Galluzzi P, Morgese G, Balestri P. Pontocerebellar hypoplasia type 2: Further clinical characterization and evidence of positive response of dyskinesia to levodopa. *J Neurol.* 2002; 249(5):596-600.

**KIAA0226 İlişkili Salih Ataksisi/SCAR15**

Erken başlangıçlı serebellar ataksi, motor gelişimde gecikme, dismetri, anormal göz hareketleri ve hiporefleksi ile karakterizedir.<sup>30</sup> Kranial MR görüntülemelerde hafif derecede serebellar atrofi ile karakterizedir.

**İyonotrofik Glutamat Reseptör Delta-2 İlişkili Serebellar Ataksi/SCAR18**

GRID2 geninin kodlamış olduğu glutamat reseptör kanal delta 2 subünitesi, özellikle serebellar purkinje hücrelerinde eksprese olur.<sup>31</sup> Nistagmus, hipotoni, psikomotor gelişimde gecikme, ataksi, disartri, okülomotor apraksi, dismetri ve disdiadokinezi, canlı derin tendon refleksleri, ektansör plantar yanıt ile karakterizedir. Kranial MR görüntülemelerinde serebral ve serebellar atrofi görülmektedir.<sup>1,31</sup>

**ATG5 İlişkili İlerleyici Olmayan Serebellar Ataksi/SCAR25**

Psikomotor gelişimde gecikme, trunkal ataksi, dismetri, nistagmus ve bilişsel gerilik ile karakterizedir.<sup>32,33</sup>

**Bilişsel Gerilik, Optik Atrofi ve Deri Anormallikleri ile Beraber Görülen WDR73 İlişkili Serebellar Ataksi (CAMOS)**

Ciddi gelişimsel gerilik, psikomotor gerilik, orantılı boy kısalığı, mikrosefali, optik atrofi, konuşma bozukluğu, serebellar atrofi ile karakterizedir.<sup>1,28,34</sup>

**Mental Retardasyon ile Birlikte, İlerleyici Olmayan, CAMTA1 İlişkili Serebellar Ataksi (CANPMR)**

CAMTA1, geninden kaynaklanan bu hastalıkta, hafif derecede bilişsel gerilik, erken başlangıçlı ataksi, konuşma gecikmesi görülür.<sup>1,35,36</sup> Şaşılık, infantil hipotoni, konuşma gecikmesi, miyoklonik nöbetler ve dismorfik bulgular bildirilmiştir.<sup>1,36</sup>

**ATCAY İlişkili Serebellar Ataksi, CAYMAN Tipi.**

İlk kez 1978'de Johnson ve ark. tarafından, Büyük Cayman Ada'larındaki izole bir popülasyonda tanımlanmıştır. Belirgin hipotoni, psikomotor gerilik, belirgin ve ilerleyici olmayan serebellar disfonksiyon, nistagmus, intansiyonel tremor, disartri ve geniş tabanlı yürüyüş ile karakterizedir.

**KCNJ10-İlişkili Ataksi ve EAST/SESAME Sendromu**

İnfant döneminde jeneralize nöbetler, psikomotor gelişimde gecikme, ataksi, mental retardasyon ve elektrolit bozukluğu ile karakterizedir.<sup>37</sup> Bilişsel gerilik bazen tabloya katılmayabilir.<sup>38</sup> Ataksi, sensoryal işitme kaybı, hipokalemik metabolik asidoz ve hipomahnezemi ile karakterize formları tanımlanmıştır. Kranial MR görüntülemelerde, kaudat nukleusda, T2'de hiperintensite görülebilir.<sup>39</sup>

**Özetle,** Birçok nadir görülen otozomal dominant veya resesif olarak kalıtılan, ilerleyici olmayan, konjenital ataksiler tanımlanmıştır. Özellikle, CACNA1A, GRID2, CAMTA1 mutasyonları erken başlangıçlı ataksi ile birlikte görülür. Bu gruptaki hastalıklarda, bilişsel gerilik, spastisite ve erken başlangıçlı nöbetler gibi eşlik eden bulgular görülebilir.<sup>1</sup>

**KAYNAKLAR**

1. Bertini E, Zanni G, Boltshauser E. Nonprogressive congenital ataxias. *Handb Clin Neurol.* 2018; 155:91-103.
2. Travaglini L, Nardella M, Bellacchio E et al. Missense mutations of CACNA1A are a frequent cause of autosomal dominant nonprogressive congenital ataxia. *Eur J Ped Neurol.* 2017;21: 450-456.
3. Zambonin JL, Bellomo A, Ben-Pazi H et al. Spinocerebellar ataxia type 29 due to mutations in ITPR1: a case series and review of this emerging congenital ataxia. *Orphanet J Rare Dis.* 2017;12: 121.
4. Steinlin M, Zanger B, Boltshauser E. Non-progressive congenital ataxia with or without cerebellar hypoplasia: a review of 34 subjects. *Dev Med Child Neurol.* 1998;40: 148-154.
5. Parmeggiani A, Posar A, Scaduto MC et al. Epilepsy, intelligence, and psychiatric disorders in patients with cerebellar hypoplasia. *J Child Neurol.* 2003;18: 1-4.
6. Wassmer E, Davies P, Whitehouse WP et al. Clinical spectrum associated with cerebellar hypoplasia. *Pediatr Neurol.* 2003; 28: 347-351.



7. Ventura P, Presicci A, Perniola T et al. Mental retardation and epilepsy in patients with isolated cerebellar hypoplasia. *J Child Neurol.* 2006;21: 776–781.
8. Lamonica DA, Maximino LP, Abramides DV et al. Nonprogressive congenital cerebellar ataxia, iris heterochromia, mental retardation and language impairment in two brothers. *Clin Dysmorphol.* 2010; 19: 76–78.
9. Schmahmann JD, Sherman JC. The cerebellar cognitive affective syndrome. *Brain.* 1998;121: 561–579.
10. Tavano A, Borgatti R. Evidence for a link among cognition, language and emotion in cerebellar malformations. *Cortex.* 2010;46: 907–918.
11. Pietrobon D, Striessnig J. Neurobiology of migraine. *Nat Rev Neurosci.* 2003;4: 386e98.
12. Jodice C, Mantuano E, Veneziano L et al. Episodic ataxia type 2 (EA2) and spinocerebellar ataxia type 6 (SCA6) due to CAG repeat expansion in the CACNA1A gene on chromosome 19p. *Hum Mol Genet.* 1997; 6: 1973e8.
13. Haan J, Kors EE, Vanmolkot KR et al. Migraine genetics: an update. *Curr Pain Headache Rep.* 2005. 9: 213e20.
14. Hans M, Luvisetto S, Williams ME et al. Functional consequences of mutations in the human alpha1A calcium channel subunit linked to familial hemiplegic migraine. *J Neurosci.* 1999; 19: 1610e9.
15. Reinson K, Oiglane-Shlik E, Talvik I et al. Biallelic CACNA1A mutations cause early onset epileptic encephalopathy with progressive cerebral, cerebellar, and optic nerve atrophy. *Am J Med Genet A.* 2016;170: 2173e6.
16. Zhang Y, Kaczmarek LK. Kv3.3 potassium channels and spinocerebellar ataxia. *J Physiol.* 2016; 594: 4677–4684.
17. Yamada N, Makino Y, Clark RA et al. Human inositol 1,4,5- trisphosphate type-1 receptor, InsP3R1: structure, function, regulation of expression and chromosomal localization. *Biochem J.* 1994; 302: 781–790.
18. Sugawara T, Hisatsune C, Le TD et al. Type 1 inositol trisphosphate receptor regulates cerebellar circuits by maintaining the spine morphology of purkinje cells in adult mice. *J Neurosci.* 2003;33: 12186–12196.
19. van Dijk T, Barth P, Reneman L et al. A de novo missense mutation in the inositol 1,4,5-triphosphate receptor type 1 gene causing severe pontine and cerebellar hypoplasia: expanding the phenotype of ITPR1-related spinocerebellar ataxias. *Am J Med Genet A.* 2017; 173:207–212.
20. Dixon-Salazar TJ, Silhavy JL, Udpa N et al. Exome sequencing can improve diagnosis and alter patient management. *Sci Transl Med.* 2012;4: 138ra78.
21. Kaya N, Aldhalaan H, Al-Younes B et al. Phenotypic spectrum of cerebellar ataxia associated with a novel mutation in the CA8 gene, encoding carbonic anhydrase (CA) VIII. *Am J Med Genet B Neuropsychiatr Genet.* 2011;156B: 826–834.
22. Halleck MS, Lawler JF, Blackshaw S et al. Differential expression of putative transbilayer amphipath transporters. *Physiol Genomics.* 1999;1: 139–150.
23. Cacciagli P, Haddad MR, Mignon-Ravix C et al. Disruption of the ATP8A2 gene in a patient with a t(10;13) de novo balanced translocation and a severe neurological phenotype. *Eur J Hum Genet.* 2010;18: 1360–1363.
24. Onat OE, Gulsuner S, Bilguvar K et al. Missense mutation in the ATPase, aminophospholipid transporter protein ATP8A2 is associated with cerebellar atrophy and quadrupedal locomotion. *Eur J Hum Genet.* 2013; 21:281–285.
25. GribaaM, SalihM, Anheim M et al. A new form of childhood onset, autosomal recessive spinocerebellar ataxia and epilepsy is localized at 16q21-q23. *Brain.* 2007;130: 1921–1928.
26. Guergueltcheva V, Azmanov DN, Angelicheva D et al. Autosomal-recessive congenital cerebellar ataxia is caused by mutations in metabotropic glutamate receptor 1. *Am J Hum Genet.* 2012;91: 553–564.
27. Gakh O, Cavadini P, Isaya G. Mitochondrial processing peptidases. *Biochim Biophys Acta.* 2002; 1592:63–77.
28. Megarbane A, Delague V, Salem N et al. Autosomal recessive congenital cerebellar hypoplasia and short stature in a large inbred family. *Am J Med Genet.* 1999; 87: 88–90.
29. Kvistad PH, Dahl A, Skre H. Autosomal recessive nonprogressive ataxia with an early childhood debut. *Acta Neurol Scand.* 1985; 71: 295–302.
30. Assoum M, Salih MA, Drouot N et al. Rundataxin, a novel protein with RUN and diacylglycerol binding domains, is mutant in a new recessive ataxia. *Brain.* 2010;133: 2439–2447.
31. Utine GE, Haliloğlu G, Salanci B. A homozygous deletion in GRID2 causes a human phenotype with cerebellar ataxia and atrophy. *J Child Neurol.* 2013;28 (7): 926–932.
32. Kim M, Sandford E, Gatica D et al. Mutation in ATG5 reduces autophagy and leads to ataxia with developmental delay. *eLife.* 2016;5: e12245.
33. Yapici Z, Eraksoy M. Non-progressive congenital ataxia with cerebellar hypoplasia in three families. *Acta Paediatr.* 2005;94: 248–253.
34. Delague V, Bareil C, Bouvagnet P et al. A new autosomal recessive non-progressive congenital cerebellar ataxia associated with mental retardation, optic atrophy, and skin abnormalities (CAMOS) maps to chromosome 15q24-q26 in a large consanguineous Lebanese Druze family. *Neurogenetics.* 2002; 4: 23–27.
35. Bas-Orth C, Tan YW, Oliveira AM et al. The calmodulin-binding transcription activator CAMTA1 is required for long-term memory formation in mice. *Learn Mem.* 2016;23: 313–321.
36. Thevenon J, Lopez E, Keren B et al. Intragenic CAMTA1 rearrangements cause non-progressive congenital ataxia with or without intellectual disability. *J Med Genet.* 2012;49: 400–408.
37. Scholl UI, Choi M, Liu T et al. Seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance (SeSAME syndrome) caused by mutations in KCNJ10. *Proc Natl Acad Sci U S A.* 2009;106: 5842–5847.
38. Bockenbauer D, Feather S, Stanescu HC et al. Epilepsy, ataxia, sensorineural deafness, tubulopathy, and KCNJ10 mutations. *N Engl J Med.* 2009;360: 1960–1970.
39. Freudenthal B, Kulaveerasingam D, Lingappa L et al. KCNJ10 mutations disrupt function in patients with EAST syndrome. *Nephron Physiol.* 2011;119 (3): 40–48.