

BÖLÜM 92

ÇOCUKLUK ÇAĞI LÖKODİSTROFİLERİ

Gülcan AKYÜZ YÜCEL¹

Hakki AKBAYAZ²

Olcay ÜNVER³

GİRİŞ

Lökodistrofiler santral sinir sistemi (SSS) beyaz cevherini etkileyen herediter heterojen bir hastalık grubudur.¹ Son dönemde ileri görüntüleme yöntemleri ve ileri genetik testler (Whole Exom Sequencing (WES), Whole Genom Sequencing (WGS) sayesinde lökodistrofiler hakkında daha fazla bilgi sahibi olabilmekteyiz.

Manyetik rezonans görüntüleme (MRG) ve ileri görüntüleme teknikleri hastlığın tanısı, прогнозu ve tedaviye yanıtın değerlendirilmesinde oldukça yararlı bilgiler vermektedir. Bunun yanı sıra patoloji temelli tanı da teşhiste, прогнозu belirlemeye ve tedaviye yanımı değerlendirmede etkili bir yöntemdir.¹

Lökoensefalopati terimi SSS'de beyaz cevheri yaygın ve baskın olarak tutan tüm hastalıkları tanımlamak için kullanılmaktadır. Bu hastalıklar kalıtsal (herediter) ya da edinsel olabilir. Günümüzde yüzden fazla tanımlanmış kalıtsal lökoensefalopati mevcuttur. Lökoensefalopatiler, genetik lökoensefalopatiler ve lökodistrofiler olmak üzere iki gruba ayrılır.²

Lökodistrofiler primer olarak SSS'i etkileyen hastalıklar iken, lökoensefalopatiler sistemik hastalıklar olup, sekonder olarak SSS'i

beyaz cevher tutulumu olan hastalıklardır.^{3,4} Bunun yanı sıra son zamanlarda genetik lökoensefalopati yerine "lökodistrofi" terimi yaygın olarak kullanılmaktadır.⁵

Lökodistrofilerin tanısında MRG önemli bir tanı aracıdır. Bilgisayarlı tomografi (BT) beyaz cevherde hipointensiteyi gösterebilir ancak ayrıntıları belirlemeye oldukça yetersizdir. Beyaz cevher maturasyonu ve miyelinizasyonunu ilk 6 ayda T1 ağırlıklı serilerde, 6-18. yaşlarda T2 ağırlıklı daha iyi değerlendirilir.¹

Normal beyin dokusu miyelinizasyonu çocukların oldukça iyi bilinen belli bir sıra dahilinde yaklaşık ilk iki yaşta tamamlanır.^{6,7} Miyelinize olmuş beyin yapıları gri cevherle karşılaşıldığında T1 serilerde hiperintens (beyaz), T2 serilerde ise hipointens (siyah) görülür.^{6,7} Buna karşılık demiyelinize alanlar ise T1 ağırlıklı serilerde hipointens (siyah), T2 ağırlıklı serilerde hiperintens (beyaz) izlenir.

T2 ağırlıklı serilerde 1,5 yaşına gelmiş bir çocukta serebral beyaz cevher hala hiperintens (beyaz) görülyorsa alta yatan bir beyaz cevher hastalığı göz önünde bulundurulmalıdır.^{6,7} Buradaki patoloji miyelin yapımındaki yetersizlik ya da beyaz cevheri etkileyen myelini bozan başka bir durumdan kaynaklanabilir. (Resim 1.)

¹ Uzm. Dr., Sancaktepe Şehit Profesör Dr. İlhan Varank Eğitim ve Araştırma Hastanesi Çocuk Nörolojisi Kliniği
gulcan.akyuz@hotmail.com

² Arş. Gör., Marmara Üniversitesi Pendik Eğitim ve Araştırma Hastanesi Çocuk Nörolojisi Kliniği, hakkiakbeyaz@gmail.com

³ Doç. Dr., Marmara Üniversitesi Pendik Eğitim ve Araştırma Hastanesi Çocuk Nörolojisi Kliniği, olcaymd@hotmail.com

terozigot mutasyon sonucu oluşmaktadır. Klinik olarak geç çocukluk ya da erken adolasan dönemde başlayan progresif nörolojik disfonksiyon, serebellar ataksi, kognitif bozulma, tendon ksantomları, prematür ateroskleroz, katarakt, periferik nöropati, myopati bulguları izlenir. Kronik diyare ve uzamış yenidoğan sırlığı öyküde bulunabilir. Aşıl tendonu, beyin ve akciğer dokusunda kolestrol ve kolestrol esterlerinin birikimi söz konusudur. Teşhiste serum kolestanol düzeyleri ve safra alkollerinin yüksekliği önemlidir. Kranial MR'da serebral ve/veya serebellar atrofi yanısıra cerebellum, beyin sapı ve bazal ganglionlarda fokal hiperintens alanlar izlenmektedir. Tedavide oral kenodeoksikolik asit tedavisi hastalığın progressyonunu yavaşlatmada etkilidir.

L-2 Hidroksi Glutarik Asitüri

OR, geçişli L-2 OH glutarat dehidrogenez enzimini kodlayan L2HGDH geninde homozigot mutasyona bağlı oluşan bir hastalıktır. Kromozomun 14q21 bölgesinde yer alır. Hayatın ilk yılında psikomotor retardasyon ve epilepsi bulguları ile başlar, piramidal ve serebellar bulgular, spastisite, bilşsel kayıp ile ilerler. Hastalar makrosefaliktir. Hastalarda beyin tümörü insadiansı yüksektir.⁷³ Kranial MRG'de subkortikal beyaz cevher tutulumu, serebellar atrofi, putamen, globus pallidus ve dentat nükleus tutulumu ön planda izlenir. (Resim 7) İdrar organik asit incelemesinde hidroksiglutarik asit atılımı oldukça yüksektir. Mutasyonun gösterilmesi ile tanı doğrulanır. Prenatal tanı amniotik sıvıda L-2 hidroksi glutarik asit ölçümlü ile mümkündür. Spesifik bir tedavisi yoktur. Yavaş ilerleyen bir seyri vardır.

Klasik Konjenital Musküler Distrofi (Merozin Negatif)

Hipotoni, güçsüzlük ve serebral hemisferlerde yapısal bozukluk olmaksızın demiyelinizasyon ve kreatin kinaz (CK) enzim yüksekliği ile karakterizedir. Motor gelişim basamaklarını geç kazanır. Kromozom 6q22-23'te lokalize

LAMA2 geninde merozin (alfa2 laminin) olarak bilinen merozin eksikliği vardır. Kranial MR'da yaygın beyaz cevher tutulumu izlenir. PMD ayırcı tanısında düşünülmelidir, ancak bu hastalarda nistagmus izlenmez.

KAYNAKLAR

- Vanderver A, Prust M, Tonduti D, et al. Case definition and classification of leukodystrophies and leukoencephalopathies. Mol Genet Metab. 2015;114(4):494–500.
- Parikh S, Bernard G, Leventer RJ, et al. A clinical approach to the diagnosis of patients with leukodystrophies and genetic leukoencephalopathies. Mol Genet Metab. 2015;114(4):501–515.
- Kohlschütter A, Eichler F. Childhood leukodystrophies: a clinical perspective. Expert Rev Neurother. 2011;11(10):1485–1496.
- Ashrafi MR, Tavasoli AR. Childhood leukodystrophies: a literature review of updates on new definitions, classification, diagnostic approach and management. Brain Dev. 2017;39(5):369–385.
- Van der Knaap MS, Schiffmann R, Mochel F, et al. Diagnosis, prognosis, and treatment of leukodystrophies. Lancet Neurol. 2019;18 (10):962–972.
- Barkovich AJ, Kjos BO, Jackson DE, Norman D. Normal maturation of the neonatal and infant brain: MR imaging at 1.5 T. Radiology 1988;166:173–180.
- Barkovich AJ. Concepts of myelin and myelination in neuroradiology. AJNR Am J Neuroradiol 2000;21:1099– 1109.
- Van der Knaap MS, Naidu S, Pouwels PJ, et al. New syndrome characterized by hypomyelination with atrophy of the basal ganglia and cerebellum. AJNR Am J Neuroradiol 2002;23:1466–1474.
- Heim P, Claussen M, Hoffmann B, et al. Leukodystrophy incidence in Germany. Am J Med Genet A. 1997;71(4):475–478. 22.
- Ashrafi MR, Rezaei Z, Heidari M, et al. The first report of relative incidence of inherited white matter disorders in an Asian country based on an Iranian bioregistry system. J Child Neurol. 2018;33 (4):255–259.
- Bonkowski JL, Wilkes J, Bardsley T, et al. Association of diagnosis of leukodystrophy with race and ethnicity among pediatric and adolescent patients. JAMA Network Open. 2018;1(7):e185031–e185031.
- Kevelam SH, Steenweg ME, Srivastava S, et al. Update on leukodystrophies: a historical perspective and adapted definition. Neuropediatrics. 2016;47(06):349–354.
- Geren BB, Raskind J. Development of the fine structure of the myelin sheath in sciatic nerves of chick embryos. Proc Natl Acad Sci U S A. 1953;39(8):880
- van der Knaap MS, Bugiani M. Leukodystrophies: a proposed classification system based on pathological changes and pathogenetic mechanisms. Acta Neuropathol. 2017;134(3):351–382.
- Adang LA, Sherbini O, Ball L, et al. Revised consen-

- sus statement on the preventive and symptomatic care of patients with leukodystrophies. *Mol Genet Metab.* 2017;122(1–2):18–32.
16. Prust M, Wang J, Morizono H, et al. GFAP mutations, age at onset, and clinical subtypes in Alexander disease. *Neurology.* 2011;77(13):1287–1294.
 17. Schuster J, Sundblom J, Thuresson A-C, et al. Genomic duplications mediate overexpression of lamin B1 in adult-onset autosomal dominant leukodystrophy (ADLD) with autonomic symptoms. *Neurogenetics.* 2011;12(1):65–72.
 18. Köhler W, Curiel J, Vanderver A. Adulthood leukodystrophies. *Nat Rev Neurol.* 2018;14(2):94.
 19. Crow YJ, Chase DS, Lowenstein Schmidt J, et al. Characterization of human disease phenotypes associated with mutations in TREX1, RNASEH2A, RNASEH2B, RNASEH2C, SAMHD1, ADAR, and IFIH1. *Am J Med Genet Part A.* 2015;167(2):296–312.
 20. Cameron CL, Kang PB, Burns TM, et al. Multifocal slowing of nerve conduction in metachromatic leukodystrophy. *Muscle Nerve* 2004; 29:531.
 21. Datar R, Prasad AN, Tay KY, et al. Magnetic resonance imaging in the diagnosis of white matter signal abnormalities. *Neuroradiol J.* 2018;31(4):362–371.
 22. Koob M, Rousseau F, Laugel V, et al. Cockayne syndrome: a diffusion tensor imaging and volumetric study. *Br J Radiol.* 2016;89 (1067):20151033.
 23. Barreau P, Prust MJ, Crane J, et al. Focal central white matter lesions in Alexander disease. *J Child Neurol.* 2011;26(11):1422–1424.
 24. Schiffmann R, van der Knaap MS. Invited article: an MRI-based approach to the diagnosis of white matter disorders. *Neurology.* 2009;72 (8):750–759.
 25. Wolf NI, van Spaendonk RML, Hobson GM, et al. PLP1 disorders. In: GeneReviews, Adam MP, Arlinger HH, Pagon RA, et al. (Eds), University of Washington, Seattle 2019. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1182/> (Accessed on July 23, 2021).
 26. Garbern JY. Leukodystrophies. In: Neurogenetics: Clinical and Scientific Advances, Lynch DR (Ed), Taylor and Francis, New York 2005. p.469.
 27. Barkovich AJ. Magnetic resonance techniques in the assessment of myelin and myelination. *J Inherit Metab Dis* 2005; 28:311.
 28. Wilson MG, Towner JW, Forsman I, Siris E. Syndromes associated with deletion of the long arm of chromosome 18[del(18q)]. *Am J Med Genet* 1979; 3:155.
 29. van der Knaap MS, Naidu S, Pouwels PJ, et al. New syndrome characterized by hypomyelination with atrophy of the basal ganglia and cerebellum. *AJNR Am J Neuroradiol* 2002; 23:1466.
 30. Zara F, Biancheri R, Bruno C, et al. Deficiency of hyccin, a newly identified membrane protein, causes hypomyelination and congenital cataract. *Nat Genet* 2006; 38:1111.
 31. La Piana R, Tonduti D, Gordish Dressman H, et al. Brain magnetic resonance imaging (MRI) pattern recognition in Pol III-related leukodystrophies. *J Child Neurol* 2014; 29:214.
 32. Aula P, Autio S, Raivio KO, et al. "Salla disease": a new lysosomal storage disorder. *Arch Neurol* 1979; 36:88. Mohammad S, Murthy SP, Didonna A, et al. Giant axonal neuropathy-associated gigaxonin mutations impair intermediate filament protein degradation. *J Clin Invest.* 2013;123 (5):1964–1975.
 33. Gustavson KH, Hagberg B. The incidence and genetics of metachromatic leucodystrophy in northern Sweden. *Acta Paediatr Scand* 1971; 60:585.
 34. Jeworutzki E, López-Hernández T, Capdevila-Nortes X, et al. GlialCAM, a protein defective in a leukodystrophy, serves as a ClC-2 Cl⁻ channel auxiliary subunit. *Neuron.* 2012;73(5):951–961.
 35. Singh N, Bixby C, Etienne D, et al. Alexander's disease: reassessment of a neonatal form. *Childs Nerv Syst* 2012; 28:2029. Gazzero E, Baldassari S, Giacomin C, et al. Hyccin, the molecule mutated in the leukodystrophy hypomyelination and congenital cataract (HCC), is a neuronal protein. *PLoS One.* 2012;7(3):e32180.
 36. Martidis A, Yee RD, Azzarelli B, Biller J. Neuro-ophthalmic, radiographic, and pathologic manifestations of adult-onset Alexander disease. *Arch Ophthalmol* 1999; 117:265. van der Voorn JP, Kamphorst W, van der Knaap MS, et al. The leukoencephalopathy of infantile GM1 gangliosidosis: oligodendrocytic loss and axonal dysfunction. *Acta Neuropathol.* 2004;107(6):539–545.
 37. Garbern JY. Leukodystrophies. In: Neurogenetics: Clinical and Scientific Advances, Lynch DR (Ed), Taylor and Francis, New York 2005. p.469.
 38. Russo LS Jr, Aron A, Anderson PJ. Alexander's disease: a report and reappraisal. *Neurology* 1976; 26:607.
 39. Srivastava S, Waldman A, Naidu S. Alexander disease. 2002 Nov 15 [Updated 2020 Nov 12]. In: GeneReviews [Internet], Adam MP, Arlinger HH, Pagon RA, et al. (Eds), University of Washington, Seattle 2021. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1172/> (Accessed on July 26, 2021).
 40. van der Knaap MS, Naidu S, Breiter SN, et al. Alexander disease: diagnosis with MR imaging. *AJNR Am J Neuroradiol* 2001; 22:541.
 41. Brockmann K, Dechent P, Meins M, et al. Cerebral proton magnetic resonance spectroscopy in infantile Alexander disease. *J Neurol* 2003; 250:300.
 42. Orsini JJ, Escolar ML, Wasserstein MP, Caggana M. Krabbe disease. In: GeneReviews [Internet], Adam MP, Arlinger HH, Pagon RA, et al. (Eds), University of Washington, Seattle 2018. www.ncbi.nlm.nih.gov/books/NBK1238/ (Accessed on July 09, 2020).
 43. Duffner PK, Barczykowski A, Jalal K, et al. Early infantile Krabbe disease: results of the World-Wide Krabbe Registry. *Pediatr Neurol* 2011; 45:141.
 44. Bascou N, DeRenzo A, Poe MD, Escolar ML. A prospective natural history study of Krabbe disease in a patient cohort with onset between 6 months and 3 years of life. *Orphanet J Rare Dis* 2018; 13:126.
 45. Rafi MA, Luzi P, Chen YQ, Wenger DA. A large deletion together with a point mutation in the GALC gene is a common mutant allele in patients with

- infantile Krabbe disease. *Hum Mol Genet* 1995; 4:1285. Waldman AT. Leukodystrophies. *Continuum*. 2018;24(1):130–149.
46. Beslow LA, Schwartz ES, Bönnemann CG. Thickening and enhancement of multiple cranial nerves in conjunction with cystic white matter lesions in early infantile Krabbe disease. *Pediatr Radiol* 2008; 38:694.
47. Miller RG, Gutmann L, Lewis RA, Sumner AJ. Acquired versus familial demyelinative neuropathies in children. *Muscle Nerve* 1985; 8:205. Freeman SH, Hyman BT, Sims KB, et al. Adult onset leukodystrophy with neuroaxonal spheroids: clinical, neuroimaging and neuropathologic observations. *Brain Pathol*. 2009;19(1):39–47.
48. Komatsuzaki S, Zielonka M, Mountford WK, et al. Clinical characteristics of 248 patients with Krabbe disease: quantitative natural history modeling based on published cases. *Genet Med* 2019; 21:2208.
49. Moser HW, Moser AB. Peroxisomal disorders: overview. *Ann N Y Acad Sci* 1996; 804:427.
50. Mosser J, Douar AM, Sarde CO, et al. Putative X-linked adrenoleukodystrophy gene shares unexpected homology with ABC transporters. *Nature* 1993; 361:726.
51. van Roermund CW, Visser WF, Ijlst L, et al. The human peroxisomal ABC half transporter ALDP functions as a homodimer and accepts acyl-CoA esters. *FASEB J* 2008; 22:4201.
52. Raymond GV, Moser AB, Fatemi, A. X-linked adrenoleukodystrophy. In GeneReviews (Last updated February 15, 2018). Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1315/> (Accessed on June 20, 2018).
53. van Geel BM, Bezman L, Loes DJ, et al. Evolution of phenotypes in adult male patients with X-linked adrenoleukodystrophy. *Ann Neurol* 2001; 49:186. Koob M, Rousseau F, Laugel V, et al. Cockayne syndrome: a diffusion tensor imaging and volumetric study. *Br J Radiol*. 2016;89 (1067):20151033.
54. Moser HW, Moser AB, Smith KD, et al. Adrenoleukodystrophy: phenotypic variability and implications for therapy. *J Inherit Metab Dis* 1992; 15:645.
55. Dubey P, Fatemi A, Barker PB, et al. Spectroscopic evidence of cerebral axonopathy in patients with “pure” adrenomyeloneuropathy. *Neurology* 2005; 64:304.
56. Mahmood A, Dubey P, Moser HW, Moser A. X-linked adrenoleukodystrophy: therapeutic approaches to distinct phenotypes. *Pediatr Transplant* 2005; 9 Suppl 7:55.
57. Peters C, Charnas LR, Tan Y, et al. Cerebral X-linked adrenoleukodystrophy: the international hematopoietic cell transplantation experience from 1982 to 1999. *Blood* 2004; 104:881.
58. Kemp S, Huffnagel IC, Linthorst GE, et al. Adrenoleukodystrophy - neuroendocrine pathogenesis and redefinition of natural history. *Nat Rev Endocrinol* 2016; 12:606.
59. Cosma MP, Pepe S, Annunziata I, et al. (2003) The multiple sulphatase deficiency gene encodes an essential and limiting factor for the activity of sulphatases. *Cell* 113: 445–56
60. Victoria San Antonio Arce JCP, Alexis Arzimanoglou and Robert Ouvrier. Aicardi's Diseases of the Nervous System in Childhood. Arzimanoglou A, editor: Mac Keith Press; 2018
61. Kaul R, Gao GP, Balamurugan K, Matalon R (1993) Cloning of the human aspartoacylase cDNA and a common missense mutation in Canavan disease. *Nat Genet* 5: 118–23
62. Grodd W, Krageloh-Mann I, Petersen D, et al. (1990) In vivo assessment of N-acetylaspartate in brain in spongy degeneration (Canavan's disease) by proton spectroscopy. *Lancet* 336: 437–8.
63. Topcu M, Saatci L, Topcuoglu MA, et al. (1998) Megalencephaly and leukodystrophy with mild clinical course: a report of 12 new cases. *Brain Dev* 20: 142–53.
64. Bomont P, Cavalier L, Blondeau F, et al. (2000) The gene encoding gigaxonin, a new member of the cytoskeletal BTB/kelch repeat family, is mutated in giant axonal neuropathy. *Nat Genet* 26: 370–4.
65. Tazir M, Nouioua S, Magy L, et al. (2009) Phenotypic variability in giant axonal neuropathy. *Neuromusc Disord* 19: 270–4
66. Lebon P, Badoval J, Ponsot G, et al. (1988) Intrathecal synthesis of interferon-alpha in infants with progressive familial encephalopathy. *J Neurol Sciences* 84: 201–8.
67. Crow YJ, Hayward BE, Parmar R, et al. (2006a) Mutations in the gene encoding the 3-prime-5-prime DNA exonuclease TREX1 cause Aicardi-Goutières syndrome at the AGS1 locus. *Nat Genet* 38: 917–20.
68. Crow YJ, Leitch A, Hayward BE, et al. (2006b) Mutations in genes encoding ribonuclease H2 subunits cause Aicardi-Goutières syndrome and mimic congenital viral brain infection. *Nat Genet* 38: 910–16.
69. Haaxma CA, Crow YJ, van Steensel MA, et al. (2010) A de novo p.Asp18Asn mutation in TREX1 in a patient with Aicardi-Goutières syndrome. *Am J Med Genet* 152A: 2612–17.
70. Rice GI, Bond J, Asipu A, et al. (2009) Mutations involved in Aicardi-Goutières syndrome implicate SAMHD1 as regulator of the innate immune response. *Nat Genet* 41: 829–32
71. Rice GI, Kasher PR, Forte GM, et al. (2012) Mutations in ADAR1 cause Aicardi-Goutières syndrome associated with a type I interferon signature. *Nat Genet* 44: 1243–8.
72. Rice GI, del Toro Duany Y, Jenkinson EM, et al. (2014) Gain-of-function mutations in IFIH1 cause a spectrum of human disease phenotypes associated with upregulated type I interferon signaling. *Nat Genet* 46: 503–9.
73. Aghili M, Zahedi F, Rafiee E (2009) Hydroxyglutaric aciduria and malignant brain tumor: a case report and literature review. *J Neurooncol* 91: 233–6.