Nörolojik Hastalıkların Deneysel Hayvan Modelleri

Editörler
Aysel AĞAR
Güven AKÇAY

ISBN
978-625-8155-75-4

Kitap Adı
Nörolojik Hastalıkların Deneysel Hayvan Modelleri

Editörler
Aysel AĞAR
ORCID iD: 0000-0001-8693-5648
Güven AKÇAY
ORCID iD: 0000-0003-3418-8825

Yayın Koordinatörü
Yasin DİLMEN

Sayfa ve Kapak Tasarımı
Akademisyen Dizgi Ünitesi

Yayıncı Sertifika No
47518

Baskı ve Cilt
Vadi Matbaacılık

Bisac Code
MED056000

DOI
10.37609/akya.1774

GENEL DAĞITIM
Akademisyen Kitabevi A.Ş.

Halk Sokak 5 / A Yenişehir / Ankara
Tel: 0312 431 16 33
siparis@akademisyen.com

www.akademisyen.com

ÖNSÖZ

Geleceğin güvencesi eğitime, eğitim ise öğretmene dayalıdır. Öğretmen doğan güneşe benzer. Etrafını aydınlatarak karanlıklara meydan okur. Prof. Dr. Aysel AĞAR da bilim yolunda ışık olup pek çok öğrencinin ve eğitimcinin yolunu aydınlatmıştır. Bizler de bu mirası hocamızdan alıp gelecek nesillere aktarmak adına ve santral sinir sistemi hastalıkları üzerine çalışan veya çalışmak isteyen araştırmacılara yol göstermesi amacıyla bu kitabı hazırladık. Kitabımız, bölüm yazarları tarafından 04.03.2021 tarihinde emekliliğe ayrılmış olan değerli hocamız Prof. Dr. Aysel AĞAR'a ithaf edilmiştir.

Eserinin üzerinde imzası olmayan yegâne sanatkâr öğretmendir.

M. Kemal Atatürk

KİTAP HAKKINDA

Vücudun sinir sistemiyle ilgili hastalıklar, nörolojik hastalıklar olarak bilinmektedir. Beyin, omurilik ya da sinirlerin elektriksel yapısındaki anormallikler birçok soruna yol açtığı için nörolojik hastalıklar tüm vücut sistemlerini etkileyebilmektedir. Nörolojik hastalıklar, kalp hastalıklarından sonra ölümlerin ikinci önde gelen nedenidir ve dünya genelinde insidansı ve prevelansı oldukça yüksektir. Santral sinir sisteminin karmaşık yapısı ve kompleks sinyal yolakları, hastalıkların nörobiyolojisini ve patofizyolojisini anlamayı zorlaştırdığı gibi bu hastalıklara yönelik radikal tedavilerin geliştirilmesinin de önünü kapatmaktadır. Nörolojik hastalıkların mekanizmalarının anlaşılması, bu hastalıkların önlenmesi ve etkin tedavi yöntemlerinin uygulanmasında deneysel çalışmalar önemli rol oynamaktadır. Hayvan modelleri kullanılarak yapılan araştırmalar, bu hastalıkların mekanizmalarını aydınlatabilmek ve etkin tedavi stratejileri geliştirebilmek için hayati rol oynamaktadır. Bir tedavi yönteminin araştırılmasında, yeni tedavi protokollerinin geliştirilmesinde ve yeni ilaç moleküllerinin keşfedilmesinde kemirgenler (fare, sıçan, gerbil gibi) ve kemirgen olmayan memeliler (köpek, tavşan, kedi, domuz, şempanze gibi) sıklıkla kullanılmaktadır. Hayvan modellerinde hem normal hem de anormal beyin fonksiyonunun temel nöronal mekanizmaları araştırılmaktadır. Her modelin kendine özgü avantajları ve dezavantajları vardır. Bu nedenle araştırmacı, belirli bir model seçmeden önce bu avantajların ve kısıtlamaların farkında olarak çalışması için en uygun modeli tercih etmelidir. Nörolojik hastalıklar modellenirken büyük ölçüde kemirgen modelleri kullanılmaktadır. Çünkü; kemirgenlerin üretimi ve manipülasyonu kolaydır, boyutları küçüktür ve anatomileri insanlarınkine benzerdir. Bu kitapta Alzheimer, epilepsi, migren, nöropatik ağrı, Parkinson, serebral iskemi ve travmatik beyin hasarı deneysel modellerine ve bu modeller arasından çalışmalarda en sık kullanılan yöntemlere detaylı olarak yer verilmiştir. Kliniği yansıtan ve ilaç araştırmalarında en çok tercih edilen modellerin protokolleri de görseller ile zenginleştirilerek bölümler oluşturulmuştur. Kitap bölüm yazarları, daha önce bu alanda çalışmış olan bilgi ve tecrübe sahibi araştırmacılardan oluşmaktadır.

İÇİNDEKİLER

YAZARLAR

Prof. Dr. Ahmet HACIMÜFTÜOĞLU
Atatürk Üniversitesi Tıp Fakültesi, Tıbbi Farmakoloji AD.
0000-0002-9658-3313

Dr. Öğr. Üyesi Güven AKÇAY
Hitit Üniversitesi Tıp Fakültesi, Biyofizik AD.
0000-0003-3418-8825

Dr. Öğr. Üyesi Betül ÇİÇEK
Erzincan Binali Yıldırım Üniversitesi, Fizyoloji AD.
0000-0003-1395-1326

Dr. Öğr. Üyesi Dilara NEMUTLU SAMUR
Alanya Alaaddin Keykubat Üniversitesi Tıp Fakültesi, Tıbbi Farmakoloji AD.
0000-0003-2630-6182

Arş. Gör. Dr. Betül DANIŞMAN
Atatürk Üniversitesi, Biyofizik AD.
0000-0002-3812-9884

Arş Gör. Fatma YEŞİLYURT
Atatürk Üniversitesi Tıp Fakültesi, Tıbbi Farmakoloji AD.
0000-0002-1336-6322

Arş. Gör. Büşra CESUR
Akdeniz Üniversitesi Tıp Fakültesi Tıbbi Farmakoloji AD.
0000-0003-1644-7061

DENEYSEL ALZHEİMER HASTALIĞI MODELLERİ

BÖLÜM 1

Betül DANIŞMAN[1]
Betül ÇİÇEK[2]

Giriş

Demansın en yaygın şekli olarak da bilinen Alzheimer hastalığı, dünya çapında yaklaşık 50 milyon kişiyi etkiyen ilerleyici, nörodejeneratif bir hastalıktır. Yıllar içinde Alzheimer hastalığı için çeşitli tedaviler geliştirilmiş olsa da bu hastalık için mevcut iyileştirici tedaviler henüz bulunamamış, sadece semptomlar hafifletilmiştir. Alzheimer hastalığı, çok sayıda biyolojik yolu etkileyen, çeşitli etiyolojik faktörlere atfedilebilen karmaşık bir durumdur. Deneysel modeller hastalığın patogenezini daha iyi anlayabilmek ve klinik öncesi yeni tedavileri test edebilmek için önemlidir. Alzheimer hastalığının karmaşıklığını tamamen kapsayan bir araştırma modeli geliştirmek zor olsa da çeşitli yönleriyle ilgili bilgi edinmek için birçok model geliştirilmiştir.

Alzheimer hastalığının deneysel hayvan modelleri, hem transgenik hayvanları hem de doğal, transgenik olmayan Alzheimer hastalığı modellerini içerir. Tüm bu modeller, Alzheimer hastalığı patolojisinin altında yatan temel mekanizmaların araştırılması ve ayrıca bu hastalığa karşı hedeflenen yeni tedavilerin test edilmesi için değerlidir.

Her bir modelin belirli sınırlamaları olsa da araştırmacılar uygun deneysel modelli kullanarak, Alzheimer hastalığı hakkında önemli bilgilere ulaşabilirler. Bu bölümde, güncel modellerin nasıl kullanılacağına ve bu modellerin insan hastalıklarıyla benzerlik ve farklılıklarına odaklanılarak, mevcut deneysel Alzheimer hastalığı modelleri özetlenmektedir.

[1] Arş. Gör. Dr., Atatürk Üniversitesi, Biyofizik AD., brrhd02@gmail.com
[2] Dr. Öğr. Üyesi, Erzincan Binali Yıldırım Üniversitesi, Fizyoloji AD., bcicek@erzincan.edu.tr

delleri için ek bilgi sağlayarak hücre dışı ortamların doğru ve tekrarlanabilir kontrolünü sağlar. Tüm bu modeller, AH'nin patofizyolojisini araştırmak ve potansiyel tedavileri değerlendirmek için uygundur. Her model belirli avantajlar ve belirli sınırlamalar sunar. Deneysel bir modelin seçilmesi, hem araştırma hedeflerine hem de çalışmanın temel amaçlarına bağlıdır.

Bir bütün olarak, bu bölümde tartışılan deneysel AH modelleri AH'yi anlamamıza katkıda bulunmuştur. Bununla birlikte, bu modellerin hiçbiri, AH alt tiplerinin büyük çoğunluğunda hastalık ilerlemesinin tüm yönlerini yeniden üretemez. Bu nedenle, mevcut modeller, insan AH'sinin karmaşık koşullarını tam olarak çoğaltmak için ek modifikasyonlar gerektirir. Kuşkusuz, deneysel modeller gelecekteki AH araştırmalarında hayati bir rol oynamaya devam edecektir.

Kaynaklar

1. Mucke L. 2009. Alzheimer's disease. *Nature* 461:895-7
2. Small DH, Cappai R. 2006. Alois Alzheimer and Alzheimer's disease: a centennial perspective. *Journal of neurochemistry* 99:708-10
3. Drummond E, Wisniewski T. 2017. Alzheimer's disease: experimental models and reality. *Acta neuropathologica* 133:155-75
4. Nelson PT, Alafuzoff I, Bigio EH, Bouras C, Braak H, et al. 2012. Correlation of Alzheimer disease neuropathologic changes with cognitive status: a review of the literature. *Journal of Neuropathology & Experimental Neurology* 71:362-81
5. Li X, Bao X, Wang R. 2016. Experimental models of Alzheimer's disease for deciphering the pathogenesis and therapeutic screening. *International journal of molecular medicine* 37:271-83
6. Glenner G, Wong C. 1988. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Alzheimer Disease & Associated Disorders* 2:134
7. Schindowski K, Bretteville A, Leroy K, Bégard S, Brion J-P, et al. 2006. Alzheimer's disease-like tau neuropathology leads to memory deficits and loss of functional synapses in a novel mutated tau transgenic mouse without any motor deficits. *The American journal of pathology* 169:599-616
8. Gonzalo-Ruiz A, Gonzalez I, Sanz-Anquela J. 2003. Effects of β-amyloid protein on serotoninergic, noradrenergic, and cholinergic markers in neurons of the pontomesencephalic tegmentum in the rat. *Journal of chemical neuroanatomy* 26:153-69
9. Moghul S, Wilkinson D. 2001. Use of acetylcholinesterase inhibitors in Alzheimer's disease. *Expert Review of Neurotherapeutics* 1:61-9
10. Cacace R, Sleegers K, Van Broeckhoven C. 2016. Molecular genetics of early-onset Alzheimer's disease revisited. *Alzheimer's & dementia* 12:733-48
11. Bertram L, Tanzi RE. 2012. The genetics of Alzheimer's disease. *Progress in molecular biology and translational science* 107:79-100
12. Cuyvers E, Sleegers K. 2016. Genetic variations underlying Alzheimer's disease: evidence from genome-wide association studies and beyond. *The Lancet Neurology* 15:857-68

13. Shinohara M, Fujioka S, Murray ME, Wojtas A, Baker M, et al. 2014. Regional distribution of synaptic markers and APP correlate with distinct clinicopathological features in sporadic and familial Alzheimer's disease. *Brain* 137:1533-49
14. Borenstein AR, Copenhaver CI, Mortimer JA. 2006. Early-life risk factors for Alzheimer disease. *Alzheimer Disease & Associated Disorders* 20:63-72
15. Knopman DS, Jack Jr CR, Lundt ES, Weigand SD, Vemuri P, et al. 2016. Evolution of neurodegeneration-imaging biomarkers from clinically normal to dementia in the Alzheimer disease spectrum. *Neurobiology of aging* 46:32-42
16. Elder GA, Gama Sosa MA, De Gasperi R. 2010. Transgenic mouse models of Alzheimer's disease. *Mount Sinai Journal of Medicine: A Journal of Translational and Personalized Medicine: A Journal of Translational and Personalized Medicine* 77:69-81
17. Games D, Adams D, Alessandrini R, Barbour R, Borthelette P, et al. 1995. Alzheimer-type neuropathology in transgenic mice overexpressing V717F β-amyloid precursor protein. *Nature* 373:523-7
18. Hsiao K, Chapman P, Nilsen S, Eckman C, Harigaya Y, et al. 1996. Correlative memory deficits, Aβ elevation, and amyloid plaques in transgenic mice. *Science* 274:99-103
19. STURCHLER-PIERRAT C, Staufenbiel M. 2000. Pathogenic mechanisms of Alzheimer's disease analyzed in the APP23 transgenic mouse model. *Annals of the New York Academy of Sciences* 920:134-9
20. Radde R, Bolmont T, Kaeser SA, Coomaraswamy J, Lindau D, et al. 2006. Aβ42-driven cerebral amyloidosis in transgenic mice reveals early and robust pathology. *EMBO reports* 7:940-6
21. Holcomb L, Gordon MN, McGowan E, Yu X, Benkovic S, et al. 1998. Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes. *Nature medicine* 4:97-100
22. Oakley H, Cole SL, Logan S, Maus E, Shao P, et al. 2006. Intraneuronal β-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation. *Journal of Neuroscience* 26:10129-40
23. Lewis J, Dickson DW, Lin W-L, Chisholm L, Corral A, et al. 2001. Enhanced neurofibrillary degeneration in transgenic mice expressing mutant tau and APP. *Science* 293:1487-91
24. Götz J, Probst A, Spillantini MG, Schäfer T, Jakes R, et al. 1995. Somatodendritic localization and hyperphosphorylation of tau protein in transgenic mice expressing the longest human brain tau isoform. *The EMBO journal* 14:1304-13
25. Allen B, Ingram E, Takao M, Smith MJ, Jakes R, et al. 2002. Abundant tau filaments and nonapoptotic neurodegeneration in transgenic mice expressing human P301S tau protein. *Journal of Neuroscience* 22:9340-51
26. Bolmont T, Clavaguera F, Meyer-Luehmann M, Herzig MC, Radde R, et al. 2007. Induction of tau pathology by intracerebral infusion of amyloid-β-containing brain extract and by amyloid-β deposition in APP× Tau transgenic mice. *The American journal of pathology* 171:2012-20
27. Grueninger F, Bohrmann B, Czech C, Ballard TM, Frey JR, et al. 2010. Phosphorylation of Tau at S422 is enhanced by Aβ in TauPS2APP triple transgenic mice. *Neurobiology of Disease* 37:294-306
28. Fryer JD, Simmons K, Parsadanian M, Bales KR, Paul SM, et al. 2005. Human apolipoprotein E4 alters the amyloid-β 40: 42 ratio and promotes the formation of cerebral amyloid angiopathy in an amyloid precursor protein transgenic model. *Journal of Neuroscience* 25:2803-10
29. Oddo S, Caccamo A, Shepherd JD, Murphy MP, Golde TE, et al. 2003. Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Aβ and synaptic dysfunction. *Neuron* 39:409-21

30. Janelsins MC, Mastrangelo MA, Oddo S, LaFerla FM, Federoff HJ, Bowers WJ. 2005. Early correlation of microglial activation with enhanced tumor necrosis factor-alpha and monocyte chemoattractant protein-1 expression specifically within the entorhinal cortex of triple transgenic Alzheimer's disease mice. *Journal of neuroinflammation* 2:1-12
31. Saito T, Matsuba Y, Mihira N, Takano J, Nilsson P, et al. 2014. Single App knock-in mouse models of Alzheimer's disease. *Nature neuroscience* 17:661-3
32. Do Carmo S, Cuello AC. 2013. Modeling Alzheimer's disease in transgenic rats. *Molecular neurodegeneration* 8:1-11
33. Cohen RM, Rezai-Zadeh K, Weitz TM, Rentsendorj A, Gate D, et al. 2013. A transgenic Alzheimer rat with plaques, tau pathology, behavioral impairment, oligomeric aβ, and frank neuronal loss. *Journal of Neuroscience* 33:6245-56
34. Leon WC, Canneva F, Partridge V, Allard S, Ferretti MT, et al. 2010. A novel transgenic rat model with a full Alzheimer's-like amyloid pathology displays pre-plaque intracellular amyloid-β-associated cognitive impairment. *Journal of Alzheimer's Disease* 20:113-26
35. Braidy N, Poljak A, Jayasena T, Mansour H, Inestrosa NC, Sachdev PS. 2015. Accelerating Alzheimer's research through 'natural'animal models. *Current opinion in psychiatry* 28:155-64
36. Camus S, Ko WKD, Pioli E, Bezard E. 2015. Why bother using non-human primate models of cognitive disorders in translational research? *Neurobiology of learning and memory* 124:123-9
37. Gearing M, Tigges J, Mori H, Mirra S. 1997. β-Amyloid (Aβ) deposition in the brains of aged orangutans. *Neurobiology of aging* 18:139-46
38. Rosen RF, Farberg AS, Gearing M, Dooyema J, M. Long P, et al. 2008. Tauopathy with paired helical filaments in an aged chimpanzee. *Journal of Comparative Neurology* 509:259-70
39. Perez SE, Raghanti MA, Hof PR, Kramer L, Ikonomovic MD, et al. 2013. Alzheimer's disease pathology in the neocortex and hippocampus of the western lowland gorilla (Gorilla gorilla gorilla). *Journal of Comparative Neurology* 521:4318-38
40. Schütt T, Helboe L, Pedersen LØ, Waldemar G, Berendt M, Pedersen JT. 2016. Dogs with cognitive dysfunction as a spontaneous model for early Alzheimer's disease: a translational study of neuropathological and inflammatory markers. *Journal of Alzheimer's Disease* 52:433-49
41. Smolek T, Madari A, Farbakova J, Kandrac O, Jadhav S, et al. 2016. Tau hyperphosphorylation in synaptosomes and neuroinflammation are associated with canine cognitive impairment. *Journal of Comparative Neurology* 524:874-95
42. Davis PR, Head E. 2014. Prevention approaches in a preclinical canine model of Alzheimer's disease: benefits and challenges. *Frontiers in pharmacology* 5:47
43. Bao S, Zheng H, Chen C, Zhang Y, Bao L, et al. 2021. Nfe2l1 deficiency mitigates streptozotocin-induced pancreatic β-cell destruction and development of diabetes in male mice. *Food and Chemical Toxicology* 158:112633
44. Voronkov D, Stavrovskaya A, Stelmashook E, Genrikhs E, Isaev N. 2019. Neurodegenerative Changes in Rat Brain in Streptozotocin Model of Alzheimer's Disease. *Bulletin of Experimental Biology & Medicine* 166
45. Kamat PK. 2015. Streptozotocin induced Alzheimer's disease like changes and the underlying neural degeneration and regeneration mechanism. *Neural regeneration research* 10:1050
46. Gao C, Liu Y, Jiang Y, Ding J, Li L. 2014. Geniposide Ameliorates Learning Memory Deficits, Reduces Tau Phosphorylation and Decreases Apoptosis via GSK 3β Pathway in Streptozotocin-Induced A lzheimer Rat Model. *Brain pathology* 24:261-9
47. Kamat PK, Rai S, Swarnkar S, Shukla R, Nath C. 2014. Mechanism of synapse redox stress in okadaic acid (ICV) induced memory impairment: role of NMDA receptor. *Neurochemistry international* 76:32-41

48. Paxinos G, Watson C. 2006. *The rat brain in stereotaxic coordinates: hard cover edition*. Elsevier
49. Moreira-Silva D, Carrettiero DC, Oliveira AS, Rodrigues S, dos Santos-Lopes J, et al. 2018. Anandamide effects in a streptozotocin-induced Alzheimer's disease-like sporadic dementia in rats. *Frontiers in neuroscience* 12:653
50. Hacımüftüoğlu A, Okkay U, Sağsöz M, Taşpınar M. 2021. DENEYSEL ALZHEİMER MODELİNDE HİPOKAMPÜSTE GLUTAMAT GERİ ALINIM PARAMETRELERİNİN DEĞERLENDİRİLMESİ.
51. Woods SC, Seeley RJ, Rushing PA, D'Alessio D, Tso P. 2003. A controlled high-fat diet induces an obese syndrome in rats. *The Journal of nutrition* 133:1081-7
52. Demetrius LA, Driver J. 2013. Alzheimer's as a metabolic disease. *Biogerontology* 14:641-9
53. Herculano B, Tamura M, Ohba A, Shimatani M, Kutsuna N, Hisatsune T. 2013. β-alanyl-L-histidine rescues cognitive deficits caused by feeding a high fat diet in a transgenic mouse model of Alzheimer's disease. *Journal of Alzheimer's Disease* 33:983-97
54. Haley RW, Dietschy JM. 2000. Is there a connection between the concentration of cholesterol circulating in plasma and the rate of neuritic plaque formation in Alzheimer disease? *Archives of neurology* 57:1410-2
55. Wood WG, Eckert GP, Igbavboa U, Müller WE. 2003. Amyloid beta-protein interactions with membranes and cholesterol: causes or casualties of Alzheimer's disease. *Biochimica et Biophysica Acta (BBA)-Biomembranes* 1610:281-90
56. Wu Y-Y, Wang X, Tan L, Liu D, Liu X-H, et al. 2013. Lithium attenuates scopolamine-induced memory deficits with inhibition of GSK-3β and preservation of postsynaptic components. *Journal of Alzheimer's Disease* 37:515-27
57. Vandal M, White PJ, Tremblay C, St-Amour I, Chevrier G, et al. 2014. Insulin reverses the high-fat diet–induced increase in brain Aβ and improves memory in an animal model of Alzheimer disease. *Diabetes* 63:4291-301
58. Pinton S, Brüning CA, Oliveira CES, Prigol M, Nogueira CW. 2013. Therapeutic effect of organoselenium dietary supplementation in a sporadic dementia of Alzheimer's type model in rats. *The Journal of nutritional biochemistry* 24:311-7
59. Nakamura S, Murayama N, Noshita T, Annoura H, Ohno T. 2001. Progressive brain dysfunction following intracerebroventricular infusion of beta1–42-amyloid peptide. *Brain research* 912:128-36
60. Winslow J, Camacho F. 1995. Cholinergic modulation of a decrement in social investigation following repeated contacts between mice. *Psychopharmacology* 121:164-72
61. Sain H, Sharma B, Jaggi A, Singh N. 2011. Pharmacological investigations on potential of peroxisome proliferator-activated receptor-gamma agonists in hyperhomocysteinemia-induced vascular dementia in rats. *Neuroscience* 192:322-33
62. Kamat PK, Rai S, Nath C. 2013. Okadaic acid induced neurotoxicity: an emerging tool to study Alzheimer's disease pathology. *Neurotoxicology* 37:163-72
63. Zhang J, Li P, Wang Y, Liu J, Zhang Z, et al. 2013. Ameliorative effects of a combination of baicalin, jasminoidin and cholic acid on ibotenic acid-induced dementia model in rats. *PLoS One* 8:e56658
64. Bonda DJ, Lee H-g, Blair JA, Zhu X, Perry G, Smith MA. 2011. Role of metal dyshomeostasis in Alzheimer's disease. *Metallomics* 3:267-70
65. Squire LR, Zola-Morgan S. 1988. Memory: brain systems and behavior. *Trends in neurosciences* 11:170-5
66. Liu J, Zhang Z, Li J-T, Zhu Y-H, Zhou H-L, et al. 2009. Effects of NT-4 gene modified fibroblasts transplanted into AD rats. *Neuroscience letters* 466:1-5
67. Savage LM, Sweet AJ, Castillo R, Langlais PJ. 1997. The effects of lesions to thalamic lateral internal medullary lamina and posterior nuclei on learning, memory and habituation in the rat. *Behavioural brain research* 82:133-47

68. V Colom L, T Castaneda M, Hernandez S, Perry G, Jaime S, Touhami A. 2011. Intrahippocampal amyloid-β (1-40) injections injure medial septal neurons in rats. *Current Alzheimer Research* 8:832-40
69. Sadigh-Eteghad S, Sabermarouf B, Majdi A, Talebi M, Farhoudi M, Mahmoudi J. 2015. Amyloid-beta: a crucial factor in Alzheimer's disease. *Medical principles and practice* 24:1-10
70. McLarnon JG, Ryu JK. 2008. Relevance of abeta1-42 intrahippocampal injection as an animal model of inflamed Alzheimer's disease brain. *Curr Alzheimer Res* 5:475-80
71. Yankner BA, Duffy LK, Kirschner DA. 1990. Neurotrophic and neurotoxic effects of amyloid beta protein: reversal by tachykinin neuropeptides. *Science* 250:279-82
72. Avaliani N, Sørensen AT, Ledri M, Bengzon J, Koch P, et al. 2014. Optogenetics reveal delayed afferent synaptogenesis on grafted human-induced pluripotent stem cell-derived neural progenitors. *Stem Cells* 32:3088-98
73. Green K, Smith I, Laferla F. 2007. Role of calcium in the pathogenesis of Alzheimer's disease and transgenic models. *Calcium Signalling and Disease*:507-21
74. Akhter R, Sanphui P, Biswas SC. 2014. The essential role of p53-up-regulated modulator of apoptosis (Puma) and its regulation by FoxO3a transcription factor in β-amyloid-induced neuron death. *Journal of Biological Chemistry* 289:10812-22
75. Nepovimova E, Uliassi E, Korabecny J, Pena-Altamira LE, Samez S, et al. 2014. Multitarget drug design strategy: quinone–tacrine hybrids designed to block amyloid-β aggregation and to exert anticholinesterase and antioxidant effects. *Journal of medicinal chemistry* 57:8576-89
76. Gong C-X, Lidsky T, Wegiel J, Grundke-Iqbal I, Iqbal K. 2001. Metabolically active rat brain slices as a model to study the regulation of protein phosphorylation in mammalian brain. *Brain research protocols* 6:134-40
77. Jang J, Yoo J-E, Lee J-A, Lee DR, Kim JY, et al. 2012. Disease-specific induced pluripotent stem cells: a platform for human disease modeling and drug discovery. *Experimental & molecular medicine* 44:202-13
78. Takahashi K, Okita K, Nakagawa M, Yamanaka S. 2007. Induction of pluripotent stem cells from fibroblast cultures. *Nature protocols* 2:3081-9
79. Yagi T, Ito D, Okada Y, Akamatsu W, Nihei Y, et al. 2011. Modeling familial Alzheimer's disease with induced pluripotent stem cells. *Human molecular genetics* 20:4530-9
80. Israel MA, Yuan SH, Bardy C, Reyna SM, Mu Y, et al. 2012. Probing sporadic and familial Alzheimer's disease using induced pluripotent stem cells. *Nature* 482:216-20
81. Macias MP, Gonzales AM, Siniard AL, Walker AW, Corneveaux JJ, et al. 2014. A cellular model of amyloid precursor protein processing and amyloid-β peptide production. *Journal of neuroscience methods* 223:114-22
82. Meng P, Yoshida H, Tanji K, Matsumiya T, Xing F, et al. 2015. Carnosic acid attenuates apoptosis induced by amyloid-β 1–42 or 1–43 in SH-SY5Y human neuroblastoma cells. *Neuroscience Research* 94:1-9
83. Raja WK, Mungenast AE, Lin Y-T, Ko T, Abdurrob F, et al. 2016. Self-organizing 3D human neural tissue derived from induced pluripotent stem cells recapitulate Alzheimer's disease phenotypes. *PLoS One* 11:e0161969
84. Paquet D, Bhat R, Sydow A, Mandelkow E-M, Berg S, et al. 2009. A zebrafish model of tauopathy allows in vivo imaging of neuronal cell death and drug evaluation. *The Journal of clinical investigation* 119:1382-95

DENEYSEL EPİLEPSİ MODELLERİ

BÖLÜM 2

Güven AKÇAY[1]

Giriş

Epilepsi, beyindeki nöronların anormal ve aşırı elektriksel deşarjları sonucu ortaya çıkan ve tekrarlayan spontan kronik beyin hastalığı olarak tanımlanmaktadır. Epilepsinin başlıca nedenleri arasında inme, kafa travmaları, beyin damarlarındaki yapısal bozukluklar, beyindeki hemorajiler ve genetik faktörler yer almaktadır. Epilepsi, psikolojik, fiziksel sosyal ve ekonomik problemlere yol açmaktadır. Her yıl yaklaşık olarak 2.4 milyon kişide epilepsi hastalığı görülmektedir. Epilepsi dünyada % 1 prevelansa sahip olduğu öngörülen, yaygın ve ciddi nörolojik bir bozukluktur. Dünya Sağlık Örgütü'nün 2019 verilerine göre, dünyada yaklaşık 50 milyon epilepsili hasta olduğu rapor edilmiştir. Epilepsi çocukluk çağında ve yaşlılıkta en yüksek insidansa sahip iken erken erişkinlikte ise daha düşük düzeyde olduğu gözlenmiştir. Epilepsi nöbetleri beyindeki sinir hücreleri arasındaki inhibisyon ve eksitasyon dengesinin bozulması sonucu ortaya çıkmaktadır. Epileptik hastalarının yaklaşık olarak % 20-30'unun antiepileptik ilaçlara dirençli olduğu bilinmektedir. Epilepsi tedavisinde genellikle nöbet sıklığının kontrolü üzerine odaklanılmıştır. Epilepsinin oluşumuna sebep olan mekanizmaların ve yeni tedavi yöntemlerin anlaşılmasında deneysel modellerin kullanılması sıklıkla tercih edilmektedir. Bu yüzden çok sayıda deneysel epilepsi modelleri geliştirilmiştir. Bu deneysel modeller hayvanlarda elektrik uyarımı, kimyasal konvülzanlar, genetik modeller, yapısal lezyonlar, fiziksel uyaranlar (soğuk, basınç, hipertermi, elektriksel) ile oluşturulmaktadır.

[1] Dr. Öğr. Üyesi, Hitit Üniversitesi Tıp Fakültesi, Biyofizik AD., guvenakcayibu@gmail.com

yıcıların işlev bozukluğunun epilepsiye yol açtığı bildirilmiştir (61). Voltajla aktive olan potasyum kanalının KCNQ2/3 alt birimlerindeki mutasyon, bir tür neonatal epilepsi sendromu olan Brief Fear of Negative Evaluation Scale (BFNE)'ye neden olmaktadır (62, 63). BFNE'li yenidoğan vakaların çoğunda kendiliğinden geçen nöbetler yaşarlar; ancak yenidoğanların %10-15'i ileriki yaşamlarında epilepsi geliştirmeye devam ederler (64). KCNQ2 ve KCNQ3 homozigot mutant farelerde, spontan nöbetler görülmektedir (65, 66). Sodyum kanallarının SCN1A, SCN2A veya SCN1B alt birimindeki mutasyon, bir tür çocukluk çağı epilepsisi olan ateşli nöbetlerle birlikte genel epilepsiye sebep olmaktadır. Bebeklik döneminde şiddetli miyoklonik epilepsisi (SMEI veya Dravet sendromu) olan hastalarda da SCN1A'da mutasyon tanımlanmıştır (67).

Sonuç ve Öneriler

Epilepsi, her yaştan insanı etkileyen en yaygın nörolojik durumlardan biridir. Epilepsi, spontan tekrarlayan nöbetlerin ortaya çıkması ile karakterizedir. Halihazırda mevcut ilaçlar, epilepsili hastaların yaklaşık üçte birinde nöbetlerin kontrolünde etkisizdir. Ayrıca, bu ilaçlar yan etkilerle ilişkilidir ve bunların hiçbiri bir nöbet sonrasında epilepsi gelişimini önlemede etkili değildir. Epilepsi (epileptogenez) gelişim sürecine müdahale edebilecek etkili bir terapötik strateji geliştirilmesinde epilepsi öncesi ve sonrası beyinde meydana gelen değişiklikleri incelemek çok önemlidir. Deneysel epilepsi modelleri, epileptogenez mekanizmasının yanı sıra nöbet oluşumunun anlaşılmasına büyük ölçüde katkıda bulunur. İnsan epilepsisinin bazı özelliklerini temsil edebilecek birçok model vardır. Her modelin avantajları ve dezavantajları bulunur. Araştırmacıların çalışmalarına başlamadan önce kullanacağı terapötik ilaca uygun modeli seçerken iyi analiz etmesi gerekir. Bu bağlamda sizlere yardımcı olabilmesi adına deneysel epilepsi modelleri bu bölümde özetlenmiştir.

Kaynaklar

1. Organization, W.H., *Epilepsy in the who africa region, bridging the gap: the global campaign against epilepsy*. 2004: Geneva, Switzerland.
2. Thurman, D.J., et al., *Standards for epidemiologic studies and surveillance of epilepsy.* Epilepsia, 2011. 52 Suppl 7: p. 2-26.
3. Fiest, K.M., et al., *Prevalence and incidence of epilepsy: A systematic review and meta-analysis of international studies.* Neurology, 2017. 88(3): p. 296-303.
4. Coimbra, E.R., et al., *Cognitive performance of patients with mesial temporal lobe epilepsy is not associated with human prion protein gene variant allele at codons 129 and 171.* Epilepsy Behav, 2006. 8(3): p. 635-42.

5. Allone, C., et al., *Neuroimaging and cognitive functions in temporal lobe epilepsy: A review of the literature.* J Neurol Sci, 2017. 381: p. 7-15.
6. Engel, J., Jr., *A proposed diagnostic scheme for people with epileptic seizures and with epilepsy: report of the ILAE Task Force on Classification and Terminology.* Epilepsia, 2001. 42(6): p. 796-803.
7. *Proposal for revised clinical and electroencephalographic classification of epileptic seizures. From the Commission on Classification and Terminology of the International League Against Epilepsy.* Epilepsia, 1981. 22(4): p. 489-501.
8. Jokeit, H. and A. Ebner, *Long term effects of refractory temporal lobe epilepsy on cognitive abilities: a cross sectional study.* J Neurol Neurosurg Psychiatry, 1999. 67(1): p. 44-50.
9. Hermann, B.P., et al., *Cognitive prognosis in chronic temporal lobe epilepsy.* Ann Neurol, 2006. 60(1): p. 80-7.
10. Keary, T.A., et al., *Multivariate neuropsychological prediction of seizure lateralization in temporal epilepsy surgical cases.* Epilepsia, 2007. 48(8): p. 1438-46.
11. Scharfman, H.E., *The neurobiology of epilepsy.* Curr Neurol Neurosci Rep, 2007. 7(4): p. 348-54.
12. Engel J, et al., *Causes of hurnan epilepsy*, in *Seizures and Epilepsy*. 1989: Philiadelphia,. p. 112-134.
13. Ettinger, A.B., *Structural causes of epilepsy. Tumors, cysts, stroke, and vascular malformations.* Neurol Clin, 1994. 12(1): p. 41-56.
14. Morimoto K, Fahnestock M, and R. RJ., *Kindling and status eplepticus models of epilepsy: Rewiring the brain*. 2004: Neurobiol. p. 1-60.
15. Shin, C. and J.O. McNamara, *Mechanism of epilepsy.* Annu Rev Med, 1994. 45: p. 379-89.
16. Hardman JG, Limbird LE, and G. GA., *The Pharmacological Basis Of Therapetics*. 10th ed. ed. 2001.
17. Hauser SL and B. MF., *Mechanisms of Neurologic Diseases*, in *Harrison's Principles of Internal Medicine*., Fauci AS, et al., Editors. 2008, Mc Graw Hill: United States of America.
18. Engel J and P. TA., *Epilepsy: A complehensive textbook*. 1997, Philadelphia: Lippincott-Raven.
19. Ueda, Y. and N. Tsuru, *Simultaneous monitoring of the seizure-related changes in extracellular glutamate and gamma-aminobutyric acid concentration in bilateral hippocampi following development of amygdaloid kindling.* Epilepsy Res, 1995. 20(3): p. 213-9.
20. Kaura, S., et al., *Effect of amygdaloid kindling on the content and release of amino acids from the amygdaloid complex: in vivo and in vitro studies.* J Neurochem, 1995. 65(3): p. 1240-9.
21. Vasiliev, D.S., et al., *(Morphofunctional changes in field CA1 of the rat hippocampus after pentylenetetrazole and lithium-pilocarpine induced seizures).* Zh Evol Biokhim Fiziol, 2014. 50(6): p. 463-9.
22. Pavlova, T.V., et al., *Pentylenetetrazole kindling induces activation of caspase-3 in the rat brain.* Neurosci Behav Physiol, 2004. 34(1): p. 45-7.
23. Patsoukis, N., et al., *Thiol redox state (TRS) and oxidative stress in the mouse hippocampus after pentylenetetrazol-induced epileptic seizure.* Neurosci Lett, 2004. 357(2): p. 83-6.
24. Zhu, L.J., et al., *Spatiotemporal changes of the N-methyl-D-aspartate receptor subunit levels in rats with pentylenetetrazole-induced seizures.* Neurosci Lett, 2004. 356(1): p. 53-6.
25. Ngugi, A.K., et al., *Estimation of the burden of active and life-time epilepsy: a meta-analytic approach.* Epilepsia, 2010. 51(5): p. 883-90.
26. Shorvon, S.D., *The etiologic classification of epilepsy.* Epilepsia, 2011. 52(6): p. 1052-7.
27. Van Den Herrewegen, Y., et al., *The Barnes Maze Task Reveals Specific Impairment of Spatial Learning Strategy in the Intrahippocampal Kainic Acid Model for Temporal Lobe Epilepsy.* Neurochem Res, 2019. 44(3): p. 600-608.

28. Akcay G., *Deneysel Serebral İskemi Modeline Bağlı Öğrenme ve Hafıza Değişikliklerine Transkraniyal Doğru Akım Stimülasyonunun Etkilerinin Araştırılması*, in *Biyofizik*. 2020, Akdeniz Üniversitesi. p. 102.
29. Isaacson, R.L., *Unsolved mysteries: the hippocampus.* Behav Cogn Neurosci Rev, 2002. 1(2): p. 87-107.
30. Schmidt, B., D.F. Marrone, and E.J. Markus, *Disambiguating the similar: the dentate gyrus and pattern separation.* Behav Brain Res, 2012. 226(1): p. 56-65.
31. Woollett, K. and E.A. Maguire, *Exploring anterograde associative memory in London taxi drivers.* Neuroreport, 2012. 23(15): p. 885-8.
32. Kitamura, T. and K. Inokuchi, *Role of adult neurogenesis in hippocampal-cortical memory consolidation.* Mol Brain, 2014. 7: p. 13.
33. Tanaka, K.Z., et al., *Cortical representations are reinstated by the hippocampus during memory retrieval.* Neuron, 2014. 84(2): p. 347-54.
34. C., M., *Deneysel epilepsi modelleri.* OMÜ Tıp Dergisi, 1997. 14(3): p. 147-186.
35. Bora İ, Y.S., Gürses C., *Epilepsi*. 2 ed. Vol. 1. 2018, İstanbul: Nobel Tıp Kitabevleri.
36. Dhir, A., *Pentylenetetrazol (PTZ) kindling model of epilepsy.* Curr Protoc Neurosci, 2012. Chapter 9: p. Unit9.37.
37. Aricioglu, F., I.A. Paul, and S. Regunathan, *Agmatine reduces only peripheral-related behavioral signs, not the central signs, of morphine withdrawal in nNOS deficient transgenic mice.* Neurosci Lett, 2004. 354(2): p. 153-7.
38. Bengzon, J., et al., *Suppression of epileptogenesis by modification of N-methyl-D-aspartate receptor subunit composition.* Eur J Neurosci, 1999. 11(3): p. 916-22.
39. Beghi, E., G. Giussani, and J.W. Sander, *The natural history and prognosis of epilepsy.* Epileptic Disord, 2015. 17(3): p. 243-53.
40. Beghi, E., et al., *Recommendation for a definition of acute symptomatic seizure.* Epilepsia, 2010. 51(4): p. 671-5.
41. Engel J, S.P., *What should be modeled*, in *Models of Seizures and Epilepsy*, S.P. Pitkanen A, Moshe SL, Elseiver, Amsterdam, Editor. 2006, Elseiver: Amsterdam. p. 1-14.
42. Engel J and S. PA., *What should be modeled*, in *Models of Seizures and Epilepsy*, Pitkanen A, Schwartzkroin PA, and M. SL., Editors. 2006, Elseiver: Amsterdam. p. 1-14.
43. Liao, Y., et al., *MicroRNA-328a regulates water maze performance in PTZ-kindled rats.* Brain Res Bull, 2016. 125: p. 205-10.
44. Racine, R.J., *Modification of seizure activity by electrical stimulation. II. Motor seizure.* Electroencephalogr Clin Neurophysiol, 1972. 32(3): p. 281-94.
45. Raol, Y.H. and A.R. Brooks-Kayal, *Experimental models of seizures and epilepsies.* Prog Mol Biol Transl Sci, 2012. 105: p. 57-82.
46. Hamilton, S.E., et al., *Disruption of the m1 receptor gene ablates muscarinic receptor-dependent M current regulation and seizure activity in mice.* Proc Natl Acad Sci U S A, 1997. 94(24): p. 13311-6.
47. Ahmed Juvale, I.I. and A.T. Che Has, *The evolution of the pilocarpine animal model of status epilepticus.* Heliyon, 2020. 6(7): p. e04557.
48. Ben-Ari, Y., *Limbic seizure and brain damage produced by kainic acid: mechanisms and relevance to human temporal lobe epilepsy.* Neuroscience, 1985. 14(2): p. 375-403.
49. Hellier, J.L., et al., *Recurrent spontaneous motor seizures after repeated low-dose systemic treatment with kainate: assessment of a rat model of temporal lobe epilepsy.* Epilepsy Res, 1998. 31(1): p. 73-84.
50. Williams, P.A., et al., *Development of spontaneous recurrent seizures after kainate-induced status epilepticus.* J Neurosci, 2009. 29(7): p. 2103-12.
51. Tauck, D.L. and J.V. Nadler, *Evidence of functional mossy fiber sprouting in hippocampal formation of kainic acid-treated rats.* J Neurosci, 1985. 5(4): p. 1016-22.

52. Sutula, T., J. Cavazos, and G. Golarai, *Alteration of long-lasting structural and functional effects of kainic acid in the hippocampus by brief treatment with phenobarbital.* J Neurosci, 1992. 12(11): p. 4173-87.
53. Akdogan, I.Y., GN., *Experimental Epilepsy Models and Morphologic Alterations of Experimental Epilepsy Models in Brain and Hippocampus*, in *Underlying Mechanisms of Epilepsy*. 2011. p. 269-282.
54. Löscher, W., S. Cramer, and U. Ebert, *Differences in kindling development in seven outbred and inbred rat strains.* Exp Neurol, 1998. 154(2): p. 551-9.
55. Kubová, H. and S.L. Moshé, *Experimental models of epilepsy in young animals.* J Child Neurol, 1994. 9 Suppl 1: p. S3-11.
56. Goddard, G.V., D.C. McIntyre, and C.K. Leech, *A permanent change in brain function resulting from daily electrical stimulation.* Exp Neurol, 1969. 25(3): p. 295-330.
57. Mares P, K.H., *Electrical stimulation-induced models of seizure*, in *Models of seizures and epilepsy*, S.P. Pitkanen A, Moshe SL, Editor. 2006, Elsevier. p. 153–9.
58. Lothman, E.W., et al., *Self-sustaining limbic status epilepticus induced by 'continuous' hippocampal stimulation: electrographic and behavioral characteristics.* Epilepsy Res, 1989. 3(2): p. 107-19.
59. Nissinen, J., et al., *A new model of chronic temporal lobe epilepsy induced by electrical stimulation of the amygdala in rat.* Epilepsy Res, 2000. 38(2-3): p. 177-205.
60. Mazarati, A., et al., *Epileptogenesis after self-sustaining status epilepticus.* Epilepsia, 2002. 43 Suppl 5: p. 74-80.
61. Reid, C.A., S.F. Berkovic, and S. Petrou, *Mechanisms of human inherited epilepsies.* Prog Neurobiol, 2009. 87(1): p. 41-57.
62. Rogawski, M.A., *KCNQ2/KCNQ3 K+ channels and the molecular pathogenesis of epilepsy: implications for therapy.* Trends Neurosci, 2000. 23(9): p. 393-8.
63. Singh, N.A., et al., *A novel potassium channel gene, KCNQ2, is mutated in an inherited epilepsy of newborns.* Nat Genet, 1998. 18(1): p. 25-9.
64. Ronen, G.M., et al., *Seizure characteristics in chromosome 20 benign familial neonatal convulsions.* Neurology, 1993. 43(7): p. 1355-60.
65. Singh, N.A., et al., *Mouse models of human KCNQ2 and KCNQ3 mutations for benign familial neonatal convulsions show seizures and neuronal plasticity without synaptic reorganization.* J Physiol, 2008. 586(14): p. 3405-23.
66. Peters, H.C., et al., *Conditional transgenic suppression of M channels in mouse brain reveals functions in neuronal excitability, resonance and behavior.* Nat Neurosci, 2005. 8(1): p. 51-60.
67. Escayg, A. and A.L. Goldin, *Sodium channel SCN1A and epilepsy: mutations and mechanisms.* Epilepsia, 2010. 51(9): p. 1650-8.

DENEYSEL MİGREN MODELLERİ

BÖLÜM 3

Güven AKÇAY[1]

Giriş

Migren, genellikle tekrarlayan baş ağrısı atakları, nörolojik ve sistemik semptomlarla karakterize beyin hastalığıdır. Dünyada yetişkin ve genç nüfusta ilk sırada yer alan engellilik sebebidir. Migren insanların işlevselliğini yaşam kalitesini olumsuz etkiler. Fotofobi, fonofobi, kutanöz allodini, bulantı ve kusma migrenin en karakteristik semptomlarıdır. Migren baş ağrısı genellikle tek taraflı (% 60) ve zonklayıcıdır (% 50). Bu ağrılar fiziksel aktivite (% 90) veya baş hareketleri ile daha da kötüleşir. Migrenin primer semptomları, ağrı başlangıcından saatler veya günler önce başlar. En çok karşılaşılan uyarıcı semptomlar yorgunluk, bozulmuş konsantrasyon ve boyun sertliğini içerir. Stres, anksiyete, depresyon, fotofobi, esneme, artan idrara çıkma, bulantı, ishal ve yiyecek istekleri ise ağrı başlangıcından önce ortaya çıkar. Migren, etiyolojisi tam bilinmeyen, karmaşık, çok faktörlü, tipik olarak epizodik nörovasküler hastalık olarak bilinir. Migrenin patofizyolojisi hala tam olarak anlaşılmış değildir. Patofizyolojisinin anlaşılması geliştirilecek tedaviler adına yol gösterici olacaktır. Günümüzde, terapötik ilaç ve tedavilere olan ihtiyaçtan dolayı migren ilişkili ağrının nörobiyolojisinin nedenlerini anlamak için deneysel hayvan çalışmaları önemli rol oynar. Deneysel modeller migrenin altında yatan iyonik, nörokimyasal ve hücresel mekanizmaların anlaşılmasını mümkün kılar.

[1] Dr. Öğr. Üyesi, Hitit Üniversitesi Tıp Fakültesi, Biyofizik AD., guvenakcayibu@gmail.com

kilde anlaşılmasına yol açmıştır. KYD modelini deney hayvanlarında; mekanik (iğne batırılması) , kimyasal (potasyum, glutamat, asetilkolin, yoğun foto-oksidasyon) ve elektriksel (tetanik yada galvanik) uyaranlar ile oluşturulabildiği gösterilmiştir (53, 54). KYD modelinin en basit modeli, hücre dışı K^+ artışı ile KYD'nin başlatılmasıdır.

3.5.1. KCl ile KYD Metodu

Anestezisi altına alınan hayvanlar stereotaksik çerçeveye yerleştirilerek, kafatasına 0.5 mm yarı çaplı bir delik açılıp (Bregmaya göre, 2 mm anterior ve 1 mm lateral) bu noktaya kalıcı bir kılavuz kanül yerleştirilerek 1M KCl ile bulaştırılmış bir insülin enjektörü ile pin-prick tatbik edilir (55). Akut KYD grubu genellikle birer saat ara ile 2 defa pin-prick uygulanarak 24. saatte deneyler sonlandırılır. Kronik KYD grubu ise her gün birer saat ara ile 2 defa 1M KCl uygulanarak, deney süresince tekrarlanarak uygulama yapılmalıdır. Uygulama gün sayısı deney protokolünüze göre değişiklik gösterebilir.

Sonuç ve Öneriler

Migren, yaşam kalitesini bozabilecek, değişen duyusal algı ile ilişkili, güçten düşürücü bir baş ağrısı hastalığıdır. Migren hastalığının patogenizi halen tam olarak anlaşılamamış ve sınırlı sayıda terapötik tedavi yöntemleri mevcuttur. Yeni terapötik ajanları geliştirilmesinde kimyasal, fiziksel veya genetik modifikasyonlu deneysel hayvan modelleri önemli rol oynar. Migren hastalığının altında yatan nedenleri anlamak amacıyla hayvan modelleri geliştirilmiş, ancak bunların hepsi dezavantajlara sahiptir. Hayvan modellerinin güvenilir olarak kabul edilmesi için insan migrenine benzer bir etiyoloji ve fenotip sergilemesi gerekir. Bu hastalığın değişken bir fenotiple karmaşık olduğu düşünülse de, şu anda tüm özelliklerini modelleyebilecek bir hayvan modeli bulunmamaktadır. Bu kitap bölümünde, migren hastalığı araştırmalarında sıklıkla tercih edilen kemirgen migren modelleri özetlendi.

Kaynaklar

1. WHO. *Headache disorders*. 8 April 2016; Available from: https://www.who.int/news-room/fact-sheets/detail/headache-disorders.
2. Arzani, M., et al., *Gut-brain Axis and migraine headache: a comprehensive review.* J Headache Pain, 2020. 21(1): p. 15.
3. Lipton, R.B., et al., *Prevalence and burden of migraine in the United States: data from the American Migraine Study II.* Headache, 2001. 41(7): p. 646-57.
4. Gormley, P., et al., *Common Variant Burden Contributes to the Familial Aggregation of Migraine in 1,589 Families.* Neuron, 2018. 98(4): p. 743-753.e4.

5. Harriott, A.M., et al., *Animal models of migraine and experimental techniques used to examine trigeminal sensory processing.* J Headache Pain, 2019. 20(1): p. 91.
6. Gazerani, P., *Migraine and Diet.* Nutrients, 2020. 12(6).
7. Agosti, R., *Migraine Burden of Disease: From the Patient's Experience to a Socio-Economic View.* Headache, 2018. 58 Suppl 1: p. 17-32.
8. Akerman, S., M. Romero-Reyes, and P.R. Holland, *Current and novel insights into the neurophysiology of migraine and its implications for therapeutics.* Pharmacol Ther, 2017. 172: p. 151-170.
9. Vuralli, D., et al., *Behavioral and cognitive animal models in headache research.* J Headache Pain, 2019. 20(1): p. 11.
10. Goadsby, P.J., R.B. Lipton, and M.D. Ferrari, *Migraine--current understanding and treatment.* N Engl J Med, 2002. 346(4): p. 257-70.
11. Hargreaves, R.J. and S.L. Shepheard, *Pathophysiology of migraine--new insights.* Can J Neurol Sci, 1999. 26 Suppl 3: p. S12-9.
12. Durham, P.L., *Calcitonin gene-related peptide (CGRP) and migraine.* Headache, 2006. 46 Suppl 1(Suppl 1): p. S3-8.
13. Durham, P.L., *CGRP-receptor antagonists--a fresh approach to migraine therapy?* N Engl J Med, 2004. 350(11): p. 1073-5.
14. Andreou, A.P. and L. Edvinsson, *Mechanisms of migraine as a chronic evolutive condition.* J Headache Pain, 2019. 20(1): p. 117.
15. Moriarty, M., et al., *Monoclonal Antibodies to CGRP or Its Receptor for Migraine Prevention.* The Journal for Nurse Practitioners, 2019. 15(10): p. 717-724.e1.
16. Silberstein, S.D., *Migraine pathophysiology and its clinical implications.* Cephalalgia, 2004. 24 Suppl 2: p. 2-7.
17. Arulmani, U., et al., *Experimental migraine models and their relevance in migraine therapy.* Cephalalgia, 2006. 26(6): p. 642-59.
18. Spierings, E.L., *Pathogenesis of the migraine attack.* Clin J Pain, 2003. 19(4): p. 255-62.
19. Schulte, L.H. and A. May, *The migraine generator revisited: continuous scanning of the migraine cycle over 30 days and three spontaneous attacks.* Brain, 2016. 139(Pt 7): p. 1987-93.
20. Welch, K.M., *Concepts of migraine headache pathogenesis: insights into mechanisms of chronicity and new drug targets.* Neurol Sci, 2003. 24 Suppl 2: p. S149-53.
21. Lauritzen, M., *Cortical spreading depression in migraine.* Cephalalgia, 2001. 21(7): p. 757-60.
22. Villalón, C.M., et al., *An introduction to migraine: from ancient treatment to functional pharmacology and antimigraine therapy.* Proc West Pharmacol Soc, 2002. 45: p. 199-210.
23. De Vries, P., C.M. Villalón, and P.R. Saxena, *Pharmacological aspects of experimental headache models in relation to acute antimigraine therapy.* Eur J Pharmacol, 1999. 375(1-3): p. 61-74.
24. Andreou, A.P., et al., *Animal models of headache: from bedside to bench and back to bedside.* Expert Rev Neurother, 2010. 10(3): p. 389-411.
25. Olesen, J., et al., *Origin of pain in migraine: evidence for peripheral sensitisation.* Lancet Neurol, 2009. 8(7): p. 679-90.
26. Bogduk, N., *Anatomy and physiology of headache.* Biomed Pharmacother, 1995. 49(10): p. 435-45.
27. Munro, G., I. Jansen-Olesen, and J. Olesen, *Animal models of pain and migraine in drug discovery.* Drug Discov Today, 2017. 22(7): p. 1103-1111.
28. Oshinsky, M.L. and S. Gomonchareonsiri, *Episodic dural stimulation in awake rats: a model for recurrent headache.* Headache, 2007. 47(7): p. 1026-36.
29. Melo-Carrillo, A. and A. Lopez-Avila, *A chronic animal model of migraine, induced by repeated meningeal nociception, characterized by a behavioral and pharmacological approach.* Cephalalgia, 2013. 33(13): p. 1096-105.
30. Boyer, N., et al., *General trigeminospinal central sensitization and impaired descending pain inhibitory controls contribute to migraine progression.* Pain, 2014. 155(7): p. 1196-1205.
31. Bates, E.A., et al., *Sumatriptan alleviates nitroglycerin-induced mechanical and thermal allodynia in mice.* Cephalalgia, 2010. 30(2): p. 170-8.
32. Markovics, A., et al., *Pituitary adenylate cyclase-activating polypeptide plays a key role in nitroglycerol-induced trigeminovascular activation in mice.* Neurobiol Dis, 2012. 45(1): p. 633-44.

33. De Felice, M., et al., *Triptan-induced enhancement of neuronal nitric oxide synthase in trigeminal ganglion dural afferents underlies increased responsiveness to potential migraine triggers.* Brain, 2010. 133(Pt 8): p. 2475-88.
34. Kopruszinski, C.M., et al., *Prevention of stress- or nitric oxide donor-induced medication overuse headache by a calcitonin gene-related peptide antibody in rodents.* Cephalalgia, 2017. 37(6): p. 560-570.
35. Romero-Reyes, M. and S. Akerman, *Update on animal models of migraine.* Curr Pain Headache Rep, 2014. 18(11): p. 462.
36. Chen, S.P., E.A. Tolner, and K. Eikermann-Haerter, *Animal models of monogenic migraine.* Cephalalgia, 2016. 36(7): p. 704-21.
37. van den Maagdenberg, A.M., et al., *A Cacna1a knockin migraine mouse model with increased susceptibility to cortical spreading depression.* Neuron, 2004. 41(5): p. 701-10.
38. van den Maagdenberg, A.M., et al., *High cortical spreading depression susceptibility and migraine-associated symptoms in Ca(v)2.1 S218L mice.* Ann Neurol, 2010. 67(1): p. 85-98.
39. Leo, L., et al., *Increased susceptibility to cortical spreading depression in the mouse model of familial hemiplegic migraine type 2.* PLoS Genet, 2011. 7(6): p. e1002129.
40. Tardiolo, G., P. Bramanti, and E. Mazzon, *Migraine: Experimental Models and Novel Therapeutic Approaches.* Int J Mol Sci, 2019. 20(12).
41. Mathew, R., et al., *Immunohistochemical characterization of calcitonin gene-related peptide in the trigeminal system of the familial hemiplegic migraine 1 knock-in mouse.* Cephalalgia, 2011. 31(13): p. 1368-80.
42. Park, J., et al., *Differential trigeminovascular nociceptive responses in the thalamus in the familial hemiplegic migraine 1 knock-in mouse: a Fos protein study.* Neurobiol Dis, 2014. 64: p. 1-7.
43. Langford, D.J., et al., *Coding of facial expressions of pain in the laboratory mouse.* Nat Methods, 2010. 7(6): p. 447-9.
44. Chanda, M.L., et al., *Behavioral evidence for photophobia and stress-related ipsilateral head pain in transgenic Cacna1a mutant mice.* Pain, 2013. 154(8): p. 1254-62.
45. Kulkarni, S., et al., *Adult enteric nervous system in health is maintained by a dynamic balance between neuronal apoptosis and neurogenesis.* Proc Natl Acad Sci U S A, 2017. 114(18): p. E3709-E3718.
46. Charles, A.C. and S.M. Baca, *Cortical spreading depression and migraine.* Nat Rev Neurol, 2013. 9(11): p. 637-44.
47. Bolay, H., et al., *Intrinsic brain activity triggers trigeminal meningeal afferents in a migraine model.* Nat Med, 2002. 8(2): p. 136-42.
48. Lauritzen, M., *Cerebral blood flow in migraine and cortical spreading depression.* Acta Neurol Scand Suppl, 1987. 113: p. 1-40.
49. Teive, H.A., et al., *Leao's cortical spreading depression: from experimental "artifact" to physiological principle.* Neurology, 2005. 65(9): p. 1455-9.
50. Somjen, G.G., *Mechanisms of spreading depression and hypoxic spreading depression-like depolarization.* Physiol Rev, 2001. 81(3): p. 1065-96.
51. Grafstein, B., *Mechanism of spreading cortical depression.* J Neurophysiol, 1956. 19(2): p. 154-71.
52. Van Harreveld, A. and J.S. Stamm, *Cerebral asphyxiation and spreading cortical depression.* Am J Physiol, 1953. 173(1): p. 171-5.
53. Lambert, G.A., et al., *Effect of cortical spreading depression on activity of trigeminovascular sensory neurons.* Cephalalgia, 1999. 19(7): p. 631-8.
54. Wahl, M., et al., *Involvement of calcitonin gene-related peptide (CGRP) and nitric oxide (NO) in the pial artery dilatation elicited by cortical spreading depression.* Brain Res, 1994. 637(1-2): p. 204-10.
55. Sukhotinsky, I., et al., *Chronic daily cortical spreading depressions suppress spreading depression susceptibility.* Cephalalgia, 2011. 31(16): p. 1601-8.

DENEYSEL MULTİPLE SKLEROZ MODELLERİ

BÖLÜM 4

Betül DANIŞMAN[1]

Giriş

Multiple skleroz, dünya çapında 2,5 milyondan fazla insanı etkilediği tahmin edilen, demiyelinizasyona yol açarak motor, görsel, duyusal ve otonom sistemlerde bozulmayla birlikte nöronal kaybının gerçekleştiği otoimmün bir hastalıktır. MS her hasta için farklı klinik bulgular ve ilerleyiş göstermesi nedeniyle öngörülemez bir durumdur. Sinir lifleri, koruyucu miyelin kılıf ile kaplıdır ve sağlıklı bir sinirde miyelin kılıf oluşan potansiyellerin daha hızlı iletilmesini sağlar. MS hastalığında ise bağışıklık sisteminin hücreleri beyin ve omuriliğindeki sinir liflerine saldırarak miyelin kılıfa zarar verir ve sinir hücrelerinin normal işlevi bozulur. MS'in hayvan modelleri, hastalığın patolojik mekanizmalarını ve tedavilerinin nasıl hedeflenebileceklerini tahmin etmede önemli rol oynar. Merkezi sinir sisteminde inflamasyon, demiyelinizasyon, remiyelinizasyon ve nörodejenerasyonun farklı yönlerini incelemek için oldukça yararlı olduğu kanıtlanan birçok farklı model mevcuttur. Mevcut modeller, MS patolojisinin karmaşıklığını ve heterojenliğini tam olarak yansıtmasa da, MS hastalarının tedavisi için ilaç geliştirilmesinde kullanılmaktadır. En çok kullanılan deneysel hayvan modelleri otoimmün ensefalomiyelit (EAE) ve toksin ve/veya virüs kaynaklı demiyelinizasyondur. Deneysel modeller, EAE, MS sırasında meydana gelen inflamasyon, MSS penetrasyonu, demiyelinizasyon, aksonopati ve bağışıklık hücrelerinin aracılık ettiği nöronal kayıp dâhil olmak üzere çeşitli patolojik süreçleri aydınlatmada etkili olmuştur. Bu bölümde MS araştırmalarında farklı hayvan modellerinin kullanımına ilişkin bilgiler özetlenecektir.

[1] Arş. Gör. Dr., Atatürk Üniversitesi, Biyofizik AD., brrhd02@gmail.com

modelleri, demiyelinizasyon ve remiyelinizasyon hakkında oldukça değerli bilgiler sağlar, ancak sonuçları doğrudan uygulanabilir değildir. Bir modelin veya diğerinin seçilmesi, çalışmanın özel amaçlarına bağlıdır.

Kaynaklar

1. Trapp, B.D. and K.-A. Nave, *Multiple sclerosis: an immune or neurodegenerative disorder?* Annu. Rev. Neurosci., 2008. **31**: p. 247-269.
2. Bjelobaba, I., et al., *Animal models of multiple sclerosis: Focus on experimental autoimmune encephalomyelitis.* Journal of neuroscience research, 2018. **96**(6): p. 1021-1042.
3. Huang, W.J., W.W. Chen, and X. Zhang, *Multiple sclerosis: pathology, diagnosis and treatments.* Experimental and therapeutic medicine, 2017. **13**(6): p. 3163-3166.
4. Börü, Ü.T., et al., *Prevalence of multiple sclerosis door-to-door survey in Maltepe, Istanbul, Turkey.* Neuroepidemiology, 2006. **27**(1): p. 17-21.
5. Oh, S., et al., *B-cells and humoral immunity in multiple sclerosis. Implications for therapy.* Immunologic research, 2008. **40**(3): p. 224-234.
6. Bjartmar, C., J. Wujek, and B. Trapp, *Axonal loss in the pathology of MS: consequences for understanding the progressive phase of the disease.* Journal of the neurological sciences, 2003. **206**(2): p. 165-171.
7. Kornek, B., et al., *Multiple sclerosis and chronic autoimmune encephalomyelitis: a comparative quantitative study of axonal injury in active, inactive, and remyelinated lesions.* The American journal of pathology, 2000. **157**(1): p. 267-276.
8. de Sa, J.C.C., et al., *Symptomatic therapy in multiple sclerosis: a review for a multimodal approach in clinical practice.* Therapeutic advances in neurological disorders, 2011. **4**(3): p. 139-168.
9. Slavin, A., et al., *Pathogenic mechanisms and experimental models of multiple sclerosis.* Autoimmunity, 2010. **43**(7): p. 504-513.
10. McAlpine, D. and A. Compston, *McAlpine's multiple sclerosis*. 2005: Elsevier Health Sciences.
11. Lassmann, H., *Multiple sclerosis pathology.* Cold Spring Harbor perspectives in medicine, 2018. **8**(3): p. a028936.
12. Ferguson, B., et al., *Axonal damage in acute multiple sclerosis lesions.* Brain: a journal of neurology, 1997. **120**(3): p. 393-399.
13. Hauser, S.L., et al., *B-cell depletion with rituximab in relapsing–remitting multiple sclerosis.* New England Journal of Medicine, 2008. **358**(7): p. 676-688.
14. Krishnan, C., et al., *Reduction of disease activity and disability with high-dose cyclophosphamide in patients with aggressive multiple sclerosis.* Archives of neurology, 2008. **65**(8): p. 1044-1051.
15. Baecher-Allan, C., B.J. Kaskow, and H.L. Weiner, *Multiple sclerosis: mechanisms and immunotherapy.* Neuron, 2018. **97**(4): p. 742-768.
16. Charcot, J.M., *Lectures on the diseases of the nervous system: delivered at La Salpetriere*. Vol. 1. 1877: New Sydenham Society.
17. Dawson, J.W., *The histology of disseminated sclerosis.* Edinburgh Medical Journal, 1916. **17**(5): p. 311.
18. Prineas, J., et al., *Multiple sclerosis: Remyelination of nascent lesions: Remyelination of nascent lesions.* Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society, 1993. **33**(2): p. 137-151.
19. Kidd, D., et al., *Cortical lesions in multiple sclerosis.* Brain, 1999. **122**(1): p. 17-26.
20. Kutzelnigg, A., et al., *Widespread demyelination in the cerebellar cortex in multiple sclerosis.* Brain pathology, 2007. **17**(1): p. 38-44.

21. Cifelli, A., et al., *Thalamic neurodegeneration in multiple sclerosis.* Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society, 2002. **52**(5): p. 650-653.
22. Fog, T., *Topographic distribution of plaques in the spinal cord in multiple sclerosis.* Archives of Neurology & Psychiatry, 1950. **63**(3): p. 382-414.
23. Bø, L., et al., *Subpial demyelination in the cerebral cortex of multiple sclerosis patients.* Journal of Neuropathology & Experimental Neurology, 2003. **62**(7): p. 723-732.
24. Frischer, J.M., et al., *The relation between inflammation and neurodegeneration in multiple sclerosis brains.* Brain, 2009. **132**(5): p. 1175-1189.
25. Blauth, K., et al., *Antibodies produced by clonally expanded plasma cells in multiple sclerosis cerebrospinal fluid cause demyelination of spinal cord explants.* Acta neuropathologica, 2015. **130**(6): p. 765-781.
26. J van der Star, B., et al., *In vitro and in vivo models of multiple sclerosis.* CNS & Neurological Disorders-Drug Targets (Formerly Current Drug Targets-CNS & Neurological Disorders), 2012. **11**(5): p. 570-588.
27. Majed, H.H., et al., *A novel role for Sema3A in neuroprotection from injury mediated by activated microglia.* Journal of Neuroscience, 2006. **26**(6): p. 1730-1738.
28. Kipp, M., et al., *Experimental in vivo and in vitro models of multiple sclerosis: EAE and beyond.* Multiple sclerosis and related disorders, 2012. **1**(1): p. 15-28.
29. Madill, M., et al., *In vitro and ex vivo models of multiple sclerosis.* Drug discovery today, 2016. **21**(9): p. 1504-1511.
30. de Mendoza, T.H., et al., *Organotypic cerebellar cultures: apoptotic challenges and detection.* JoVE (Journal of Visualized Experiments), 2011(51): p. e2564.
31. Shen, K. and T.J. Yuen, *Ex Vivo Myelination and Remyelination in Cerebellar Slice Cultures as a Quantitative Model for Developmental and Disease-Relevant Manipulations.* JoVE (Journal of Visualized Experiments), 2020(160): p. e61044.
32. Baker, D., et al., *Induction of chronic relapsing experimental allergic encephalomyelitis in Biozzi mice.* Journal of neuroimmunology, 1990. **28**(3): p. 261-270.
33. Croxford, A.L., F.C. Kurschus, and A. Waisman, *Mouse models for multiple sclerosis: historical facts and future implications.* Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease, 2011. **1812**(2): p. 177-183.
34. Linker, R.A. and D.-H. Lee, *Models of autoimmune demyelination in the central nervous system: on the way to translational medicine.* Experimental & translational stroke medicine, 2009. **1**(1): p. 1-10.
35. Rabchevsky, A.G., J.-D. Degos, and P.A. Dreyfus, *Peripheral injections of Freund's adjuvant in mice provoke leakage of serum proteins through the blood–brain barrier without inducing reactive gliosis.* Brain research, 1999. **832**(1-2): p. 84-96.
36. Stromnes, I.M. and J.M. Goverman, *Passive induction of experimental allergic encephalomyelitis.* Nature protocols, 2006. **1**(4): p. 1952-1960.
37. Waldner, H., et al., *Fulminant spontaneous autoimmunity of the central nervous system in mice transgenic for the myelin proteolipid protein-specific T cell receptor.* Proceedings of the National Academy of Sciences, 2000. **97**(7): p. 3412-3417.
38. Engelhardt, B., *Molecular mechanisms involved in T cell migration across the blood–brain barrier.* Journal of neural transmission, 2006. **113**(4): p. 477-485.
39. McQualter, J.L. and C.C. Bernard, *Multiple sclerosis: a battle between destruction and repair.* Journal of neurochemistry, 2007. **100**(2): p. 295-306.
40. Racke, M.K., *Experimental autoimmune encephalomyelitis (EAE).* Current protocols in neuroscience, 2001. **14**(1): p. 9.7. 1-9.7. 11.
41. AVRAHAM, B. and A.B. NUN, *VACCINATION AGAINST AUTOIMMUNE ENCEPHALOMYELITIS WITH T-LYMPHOCYTE LINE CELLS REACTIVE AGAINST MYELIN BASIC PROTEIN.* 1981.

42. Wekerle, H., et al., *Animal models.* Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society, 1994. **36**(S1): p. S47-S53.
43. Gold, R., C. Linington, and H. Lassmann, *Understanding pathogenesis and therapy of multiple sclerosis via animal models: 70 years of merits and culprits in experimental autoimmune encephalomyelitis research.* Brain, 2006. **129**(8): p. 1953-1971.
44. Procaccini, C., et al., *Animal models of multiple sclerosis.* European journal of pharmacology, 2015. **759**: p. 182-191.
45. Bjelobaba, I., D. Savic, and I. Lavrnja, *Multiple sclerosis and neuroinflammation: the overview of current and prospective therapies.* Current pharmaceutical design, 2017. **23**(5): p. 693-730.
46. Oleszak, E.L., et al., *Theiler's virus infection: a model for multiple sclerosis.* Clinical microbiology reviews, 2004. **17**(1): p. 174-207.
47. Sato, F., et al., *Theiler's virus infection: Pathophysiology of demyelination and neurodegeneration.* Pathophysiology, 2011. **18**(1): p. 31-41.
48. McCarthy, D.P., M.H. Richards, and S.D. Miller, *Mouse models of multiple sclerosis: experimental autoimmune encephalomyelitis and Theiler's virus-induced demyelinating disease*, in *Autoimmunity*. 2012, Springer. p. 381-401.
49. Lassmann, H. and M. Bradl, *Multiple sclerosis: experimental models and reality.* Acta neuropathologica, 2017. **133**(2): p. 223-244.
50. Libbey, J.E. and R.S. Fujinami, *Viral demyelinating disease in experimental animals.* Multiple sclerosis: immunology, pathology and pathophysiology. Demos, New York, NY, 2003: p. 125-133.
51. Theiler, M., *Spontaneous encephalomyelitis of mice, a new virus disease.* The Journal of experimental medicine, 1937. **65**(5): p. 705-719.
52. Tsunoda, I. and R.S. Fujinami, *Neuropathogenesis of Theiler's murine encephalomyelitis virus infection, an animal model for multiple sclerosis.* Journal of neuroimmune pharmacology, 2010. **5**(3): p. 355-369.
53. Pachner, A.R., *Experimental models of multiple sclerosis.* Current opinion in neurology, 2011. **24**(3): p. 291-299.
54. Miller, S.D., et al., *Persistent infection with Theiler's virus leads to CNS autoimmunity via epitope spreading.* Nature medicine, 1997. **3**(10): p. 1133-1136.
55. McGavern, D.B., P.D. Murray, and M. Rodriguez, *Quantitation of spinal cord demyelination, remyelination, atrophy, and axonal loss in a model of progressive neurologic injury.* Journal of neuroscience research, 1999. **58**(4): p. 492-504.
56. Jin, Y.H., et al., *The level of viral infection of antigen-presenting cells correlates with the level of development of Theiler's murine encephalomyelitis virus-induced demyelinating disease.* Journal of virology, 2015. **89**(3): p. 1867-1878.
57. Woodruff, R.H. and R.J. Franklin, *Demyelination and remyelination of the caudal cerebellar peduncle of adult rats following stereotaxic injections of lysolecithin, ethidium bromide, and complement/anti-galactocerebroside: A comparative study.* Glia, 1999. **25**(3): p. 216-228.
58. Kuypers, N.J., et al., *Functional consequences of ethidium bromide demyelination of the mouse ventral spinal cord.* Experimental neurology, 2013. **247**: p. 615-622.
59. Merrill, J.E., *In vitro and in vivo pharmacological models to assess demyelination and remyelination.* Neuropsychopharmacology, 2009. **34**(1): p. 55-73.
60. Blakemore, W. and R. Franklin, *Remyelination in experimental models of toxin-induced demyelination.* Advances in multiple Sclerosis and Experimental Demyelinating Diseases, 2008: p. 193-212.
61. Guazzo, E.P., *A technique for producing demyelination of the rat optic nerves.* Journal of Clinical Neuroscience, 2005. **12**(1): p. 54-58.
62. Goudarzvand, M., et al., *Focal injection of ethidium bromide as a simple model to study cognitive deficit and its improvement.* Basic and clinical neuroscience, 2016. **7**(1): p. 63.

63. Ibanez, C., et al., *Systemic progesterone administration results in a partial reversal of the age-associated decline in CNS remyelination following toxin-induced demyelination in male rats.* Neuropathology and applied neurobiology, 2004. **30**(1): p. 80-89.
64. Cao, Q., et al., *Transplantation of ciliary neurotrophic factor-expressing adult oligodendrocyte precursor cells promotes remyelination and functional recovery after spinalcord injury.* Journal of Neuroscience, 2010. **30**(8): p. 2989-3001.
65. Trotter, J. and M.E. Smith, *The role of phospholipases from inflammatory macrophages in demyelination.* Neurochemical research, 1986. **11**(3): p. 349-361.
66. Weltzien, H.U., *Cytolytic and membrane-perturbing properties of lysophosphatidylcholine.* Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes, 1979. **559**(2-3): p. 259-287.
67. Keough, M.B., S.K. Jensen, and V.W. Yong, *Experimental demyelination and remyelination of murine spinal cord by focal injection of lysolecithin.* JoVE (Journal of Visualized Experiments), 2015(97): p. e52679.
68. Shields, S.A., et al., *Remyelination occurs as extensively but more slowly in old rats compared to young rats following gliotoxin-induced CNS demyelination.* Glia, 1999. **28**(1): p. 77-83.
69. Lachapelle, F., et al., *Failure of remyelination in the nonhuman primate optic nerve.* Brain pathology, 2005. **15**(3): p. 198-207.
70. Smith, K. and S. Hall, *Central demyelination induced in vivo by the calcium ionophore ionomycin.* Brain, 1994. **117**(6): p. 1351-1356.
71. Horita, N., T. Ishii, and Y. Izumiyama, *Ultrastructure of 6-aminonicotinamide (6-AN)-induced lesions in the central nervous system of rats. II. Alterations of the nervous susceptibility with aging.* Acta Neuropathol, 1980. **49**(1): p. 19-27.
72. Blakemore, W., *Remyelination by Schwann cells of axons demyelinated by intraspinal injection of 6-aminonicotinamide in the rat.* Journal of neurocytology, 1975. **4**(6): p. 745-757.
73. Harrison, B., W. McDonald, and J. Ochoa, *Central demyelination produced by diphtheria toxin: an electron microscopic study.* Journal of the neurological sciences, 1972. **17**(3): p. 281-291.
74. Baba, M., et al., *Demyelination following diphtheria toxin in the presence of axonal atrophy.* Journal of the neurological sciences, 1984. **64**(2): p. 199-211.
75. Brockschnieder, D., et al., *Cell depletion due to diphtheria toxin fragment A after Cre-mediated recombination.* Molecular and cellular biology, 2004. **24**(17): p. 7636-7642.
76. Goldberg, J., et al., *Anatomical distribution of cuprizone-induced lesions in C57BL6 mice.* Journal of Molecular Neuroscience, 2015. **57**(2): p. 166-175.
77. Carlton, W.W., *Studies on the induction of hydrocephalus and spongy degeneration by cuprizone feeding and attempts to antidote the toxicity.* Life sciences, 1967. **6**(1): p. 11-19.
78. Blakemore, W., *Observations on oligodendrocyte degeneration, the resolution of status spongiosus and remyelination in cuprizone intoxication in mice.* Journal of neurocytology, 1972. **1**(4): p. 413-426.
79. Matsushima, G.K. and P. Morell, *The neurotoxicant, cuprizone, as a model to study demyelination and remyelination in the central nervous system.* Brain pathology, 2001. **11**(1): p. 107-116.
80. Miljković, D. and I. Spasojević, *Multiple sclerosis: molecular mechanisms and therapeutic opportunities.* Antioxidants & redox signaling, 2013. **19**(18): p. 2286-2334.
81. Buschmann, J.P., et al., *Inflammatory response and chemokine expression in the white matter corpus callosum and gray matter cortex region during cuprizone-induced demyelination.* Journal of Molecular Neuroscience, 2012. **48**(1): p. 66-76.
82. Doan, V., et al., *Abbreviated exposure to cuprizone is sufficient to induce demyelination and oligodendrocyte loss.* Journal of neuroscience research, 2013. **91**(3): p. 363-373.
83. Gudi, V., et al., *Glial response during cuprizone-induced de-and remyelination in the CNS: lessons learned.* Frontiers in cellular neuroscience, 2014. **8**: p. 73.

84. Baxi, E.G., et al., *Lineage tracing reveals dynamic changes in oligodendrocyte precursor cells following cuprizone-induced demyelination.* Glia, 2017. **65**(12): p. 2087-2098.
85. Skripuletz, T., et al., *De-and remyelination in the CNS white and grey matter induced by cuprizone: the old, the new, and the unexpected.* Histology and histopathology, Vol. 26, nº12 (2011), 2011.
86. Praet, J., et al., *Cellular and molecular neuropathology of the cuprizone mouse model: clinical relevance for multiple sclerosis.* Neuroscience & Biobehavioral Reviews, 2014. **47**: p. 485-505.
87. Franco-Pons, N., et al., *Behavioral deficits in the cuprizone-induced murine model of demyelination/remyelination.* Toxicology letters, 2007. **169**(3): p. 205-213.
88. Hibbits, N., et al., *Cuprizone demyelination of the corpus callosum in mice correlates with altered social interaction and impaired bilateral sensorimotor coordination.* ASN neuro, 2009. **1**(3): p. AN20090032.
89. Love, S., *Cuprizone neurotoxicity in the rat: morphologic observations.* Journal of the neurological sciences, 1988. **84**(2-3): p. 223-237.
90. Silvestroff, L., et al., *Cuprizone-induced demyelination in the rat cerebral cortex and thyroid hormone effects on cortical remyelination.* Experimental neurology, 2012. **235**(1): p. 357-367.
91. Taylor, L.C., W. Gilmore, and G.K. Matsushima, *SJL mice exposed to cuprizone intoxication reveal strain and gender pattern differences in demyelination.* Brain Pathology, 2009. **19**(3): p. 467-479.
92. Taylor, L.C., et al., *Cuprizone induces similar demyelination in male and female C57BL/6 mice and results in disruption of the estrous cycle.* Journal of neuroscience research, 2010. **88**(2): p. 391-402.
93. Kipp, M., et al., *The cuprizone animal model: new insights into an old story.* Acta neuropathologica, 2009. **118**(6): p. 723-736.
94. Shen, S., et al., *Age-dependent epigenetic control of differentiation inhibitors is critical for remyelination efficiency.* Nature neuroscience, 2008. **11**(9): p. 1024-1034.
95. Rüther, B.J., et al., *Combination of cuprizone and experimental autoimmune encephalomyelitis to study inflammatory brain lesion formation and progression.* Glia, 2017. **65**(12): p. 1900-1913.
96. O'Loughlin, E.K., et al., *C57BL6 mouse substrains demonstrate differences in susceptibility to the demyelinating effects of Cuprizone toxin.* bioRxiv, 2020.
97. Preston, M.A. and W.B. Macklin, *Zebrafish as a model to investigate CNS myelination.* Glia, 2015. **63**(2): p. 177-193.
98. Ackerman, S.D. and K.R. Monk, *The scales and tales of myelination: using zebrafish and mouse to study myelinating glia.* Brain research, 2016. **1641**: p. 79-91.
99. Gong, Z., B. Ju, and H. Wan, *Green fluorescent protein (GFP) transgenic fish and their applications.* Genetica, 2001. **111**(1): p. 213-225.
100. Buckley, C.E., P. Goldsmith, and R.J. Franklin, *Zebrafish myelination: a transparent model for remyelination?* Disease models & mechanisms, 2008. **1**(4-5): p. 221-228.
101. Kulkarni, P., et al., *Novel zebrafish EAE model: A quick in vivo screen for multiple sclerosis.* Multiple sclerosis and related disorders, 2017. **11**: p. 32-39.
102. Enriquez-Algeciras, M., et al., *Evaluation of a transgenic mouse model of multiple sclerosis with noninvasive methods.* Investigative ophthalmology & visual science, 2011. **52**(5): p. 2405-2411.

DENEYSEL NÖROPATİK AĞRI HAYVAN MODELLERİ

BÖLÜM 5

Büşra CESUR[1]

Giriş

Nöropatik ağrı (NA), primer lezyon ya da bir hastalık sonucunda gelişebilen teşhisi ve tedavisi konusunda hala aşılması gereken zorlukları olan, devamlı nosiseptif uyarının görüldüğü karmaşık bir ağrı çeşididir (1). NA klinikte sıklıkla görülen bir hastalıktır (2). NA'nın, santral ve periferik mekanizmalardan kaynaklandığı bilinmektedir. Bunlar travma, sinir hasarı, enfeksiyonlar, kemoterapötikler, toksinler, vitamin eksiklikleri gibi periferik ya da inme, tümör, multiple skleroz gibi santral rahatsızlıklar olabilmektedir. NA, sıklıkla kronik olup ciddi epidemiyolojik bir sorun olarak karşımıza çıkmaktadır. NA'da hiperaljezi, allodini ve hiperpati gibi bulgular görülmektedir. NA duygu durumu bozukluklarına yol açar ve hastanın yaşam kalitesini düşürür (3). Antikonvülzanlar, antidepresanlar, opioidler ve non opioidler NA tedavisinde kullanılsa da tam olarak etkinliği kanıtlanmış bir ilacın olmayışı, bu ağrının henüz aydınlatılmamış farklı mekanizmalardan meydana geldiğini düşündürmektedir. NA'nın, mekanizmasının yeterince aydınlatılamaması ve etkin bir tedavisinin olmayışı gibi sorunların aşılması için daha az yan etkili ve etkinliği yüksek ilaçların geliştirilmesine ihtiyaç duyulmaktadır. Bu nedenle, güncel bir problem haline gelen NA'nın anlaşılabilmesi yeni tedavi stratejilerinin geliştirilebilmesi için deneysel hayvan modellerine ihtiyaç duyulmaktadır (4). NA'nın patofizyolojisindeki farklılıklar nedeniyle pek çok NA modeli geliştirilmiştir. Bu modelleri periferik sinir hasarı, santral sinir sistemi hasarı, ilaçlara bağlı modeller, hastalık kaynaklı modeller ve geriye kalanlarını da diğer nöropati modelleri olarak sınıflandırmak mümkündür. Bu kitapta literatürde sıklıkla kullanılan deneysel nöropatik hayvan modelleri anlatılmıştır.

[1] Arş. Gör., Akdeniz Üniversitesi Tıp Fakültesi Tıbbi Farmakoloji AD., busracesur07@hotmail.com

Kaynaklar

1. Kankowski, S., et al., *Neuropathicpain: Spotlightinganatomy, experimentalmodels, mechanisms, andtherapeuticaspects.* European Journal of Neuroscience, 2021. 54(2): p. 4475-4496.
2. Scholz, J., et al., *The IASP classification of chronicpainfor ICD-11: chronicneuropathicpain.* Pain, 2019. 160(1): p. 53-59.
3. Finnerup, N.B., et al., *Neuropathicpain: frommechanismstotreatment.* Physiologicalreviews, 2021. 101(1): p. 259-301
4. Merskey, H. and N. Bogduk, *Descriptions of chronicpainsyndromesanddefinition of painterm.* IASP Press (Seattle), 1994.
5. Raja S.N., et al., *There vised International Association for the Study of Paindefinition of pain: concepts, challenges, andcompromises.* Pain, 2020; 161(9): p. 1976-1982.
6. Treede, R.D., et al., *Neuropathicpain: redefinitionand a gradingsystemforclinicalandresearchpurposes.* Neuroloy, 2008. *70*(18): p. 1630-1635.
7. Charak, S., et al., *Assessment and Management of Pain in Palliative Care.* Suggestions for Addressing Clinical and Non-Clinical Issues in Palliative Care, 2021.167.
8. Şentürk, İ.A. *Ağrı Değerlendirilmesi: Tipleri Ve Mekanizmaları.* Medical Research Reports, 2018,1(3), 78-81.
9. Bekircan-Kurt, C.E.,et al.,*NeuropathicPainFrequency in NeurologyOutpatients: A MulticenterStudy.* ArchNeuropsychiatry, 2021. *58*(4): p. 257-260.
10. Rosenberger, D.C., et al., *Challenges of neuropathic pain: focus on diabetic neuropathy.* Journal of Neural Transmission, 2020. 127(4): p. 589-624.
11. Luana, C., et al., *Neuropathic pain.* Nat. Rev. Dis. Primers, 2017. *3*: 17.
12. Kösehasanoğulları, M., ve N. Yılmaz, *Fibromiyalji sendromu ve nöropatik ağrı*. Ege Tıp Bilimleri Dergisi, 2018. *1*(1): p. 26-31.
13. Cavalli, E., et al., *The neuropathic pain: An overview of the current treatment and future therapeutic approaches.* International Journal of Immunopathology and Pharmacology, 2019. 33: 2058738419838383.
14. Bouali-Benazzouz, R., M. Landry, A. Benazzouz, and P. Fossat, *Neuropathic pain modeling: Focus on synaptic and ion channel mechanisms.* Progress in Neurobiology, 2021. *201*: 102030.
15. Yam, M. F., Loh, Y. C., Tan, C. S., Khadijah Adam, S., Abdul Manan, N., & Basir, R. *General pathways of pain sensation and the major neurotransmitters involved in pain regulation.* International journal of molecular sciences, 2018. 19(8), 2164.
16. Iyengar S, Ossipov M.H, Johnson K.W. *The role of calcitonin gene-related peptide in peripheral and central pain mechanisms including migraine.* Pain 2017; 158(4): 543-559
17. Anthony P. Ford ve Bradley J. Undem. *The therapeutic promise of ATP antagonism at P2X3 receptors in respiratory and urological disorders.* Front. Cell. Neurosci. 2013; 7: 267.
18. Yücel, A., & A. Çimen, *Nöropatik ağrı: Mekanizmalar, tanı ve tedavi.* Ağrı, 2005. 17: 1.
19. Kami, K., F. Tajima, and E. Senba, Exercise-induced hypoalgesia: potential mechanisms in animal models of neuropathic pain. *Anatomical science international*, 2017. *92*(1): p. 79-90.
20. Alvites, R., et al. *Peripheral nerve injury and axonotmesis: State of the art and recent advances.* Cogent Medicine, 2018. 5(1):p.1466404.
21. Wall, P.D., et al. *Autotomyfollowingperipheralnervelesions: experimentalanaesthesiadolorosa*. Pain, 1979. 7(2): p.103-11.
22. Amir, R., and M. Devor, *Ongoingactivity in neuromaafferentsbearingretrogradesprouts.* Brain Res, 1993. 630(1-2): p. 283-8.
23. HoKim, S., and J. MoChung, *An experimental model forperipheralneuropathyproducedbysegmentalspinalnerveligation in therat.*Pain, 1992. 50(3): p. 355–363
24. Ye, G.-L., et al., *Ligation of mouse L4 and L5 spinalnervespro-ducesrobustallodyniawithoutmajor motor functiondeficit.*Behavioural Brain Research, 2015. 276: p. 99–110.
25. Gopalsamy, B., et al., *Experimentalcharacterization of thechronicconstrictioninjury-inducedneuropathicpain model in mice.*NeurochemicalResearch, 2019. 44: p. 2123–2138.

26. Bennett, G.J., and Y.K. Xie, *A peripheralmononeuropathy in ratthatproducesdisorders of painsensationlikethoseseen in man.*Pain, 1988. 33(1): p. 87-107.
27. Authier, N., et al., *An animal model of nociceptiveperipheralneuropathyfollowing re-peated-cisplatininjections.*ExperimentalNeurology, 2003. 182(1): p. 12–20.
28. Ta, L. E., et al., *Micewithcis-platin andoxaliplatin-inducedpainfulneuropathydevelopdistinctearlyresponsestothermalstimuli.*MolecularPain, 2009. 5(1): p. 1-11.
29. Takasaki, I., et al., *Allodyniaandhyperalgesiainducedbyherpessimplexvirustype- 1 infec-tion in mice.*Pain, 2000. 86: p. 95–101.
30. Ahlgren S. C., and J.D. Levine, *Mechanicalhyperalgesia in streptozotocin-diabeticrats.* Neuroscience, 1993. 52(4): p. 1049–1055.
31. Murakami, T., et al., *Development of sensoryneuropathy in streptozotocin-induceddiabeticmice.* Brain andBehavior, 2013. 3, p. 35–41
32. Wall, P.D., et al., *Autotomyfollowingperipheralnervelesions: experimental an aesthesiadolorosa.* Pain, 1979. 7(2): p.103-11.
33. Sacerdote, P., et al., *Transient early expression of TNF-α sciatic nerve and dorsal root ganglia in a mouse model of painful peripheral neuropathy.* Neurosci Lett, 2008. 436(2): p. 210-3.
34. Muthuraman, A., A.S. Jaggi, N. Singh, and D. Singh, *Ameliorative effects of amiloride and pralidoxime in chronic constriction injury and vincristine-induced painful neuropathy in rats.* Eur J Pharmacol, 2008. 587(1-3): p. 104-11.
35. Jaggi, A.S., V. Jain, and N. Singh, *Animal models of neuropathic pain.* FundamClinPharmacol, 2011. 25(1): p. 1-28.
36. Seltzer, Z., R. Dubner, and Y. Shir, *A novelbehavioral model of neuropathicpaindisordersproduced in ratsbypartialsciaticnerveinjury.* Pain, 1990. 43: p. 205-218.
37. Wang, L.X. and Z.J. Wang, *Animalandcellularmodels of chronicpain.*AdvDrugDelivRev, 2003. 55: p. 949-65.
38. Kim, S.H., J.M. Chung, *An Experimental Model ForPeripheralNeuropathyProducedBySegmentalSpinalNerveLigation İn TheRat.* Pain,1992. 50: p. 355- 363.
39. Jaggi, A.S., V. Jain, N. Singh,*Animalmodels of neuropathicpain.* Fundam ClinPharmacol, 2011. 25(1): p. 1-28.
40. Kinnman, E., J.D. Levine, *Sensoryandsympatheticcontributionsto nevre injury-inducedsensoryabnormalities in therat.* Neuroscience, 1995. 64(3): p. 751-67.
41. La Buda, C.J., and P.J. Little, *Pharmacologicalevaluation of theselectivespinal nevre ligation model of neuropathicpain in therat.* J NeurosciMethods, 2005. 144(2): p. 175-81.
42. Yezierski, R.P, and S.H. Park, *Themechanosensitivity of spinalsensoryneuronsfollowingintraspinalinjections of quisqualicacid in therat.*NeurosciLett, 1993. 157: p. 115-119.
43. Kumar, A. H.Kaur, and A. Singh, *Neuropathicpainmodelscausedbydamagetocentralorperipheralnervoussystem.* PharmacolRep, 2018. 70(2): p. 206-16.
44. Allen, A.R., *Surgery of experimentallesion of spinalcordequivalenttocrushinjury of fracture-dislocation of spinalcolumn.* J AmMedAssoc, 1911. p. 57.
45. Siddall, P.J., C.L.Xu, and M.J. Cousins, *Allodyniafollowingtraumaticspinalcordinjury in therat.* Neuroreport, 1995. 6(9): p. 1241-4.
46. Drew, G.M., P.J. Siddall, and A.W. Duggan, *Responses of spinalneuronstocutaneousanddorsalrootstimuli in ratswithmechanicalallodyniaaftercontusivespinalcordinjury.* Brain Res, 2001. 893(1-2): p. 59-69.
47. Watson, B.D., et al., *Photochemicallyinducedspinalcordinjury in therat.* Brain Res, 1986. 367(1-2): p. 296-300.
48. Gaviria, M., et al., *A mouse model of acuteischemicspinalcordinjury.* J Neurotrauma, 2002. 19(2): p. 205-21.
49. Prado, R.,et al., *Photochemicallyinducedgradedspinalcordinfarction. Behavioral, electrophysiological, andmorphologicalcorrelates.* J Neurosurg, 1987. 67(5): p. 745-53.
50. Xu, X.J., et al., *Wiesenfeld-Hallin Z. Chronicpain-relatedsyndrome in ratsafterischemicspinalcordlesion: a possibleanimal model forpain in patientswithspinalcordinjury.* Pain, 1992. 48(2): p. 279-90.

51. Hao, J.X., et al., *Photochemicallyinducedtransientspinalischemiainducesbehavioralhypersensitivitytomechanicalandcoldstimuli, but not tonoxious-heatstimuli, in therat.* ExpNeurol, 1992. 118(2): p. 187-94.
52. Christensen, M.D., et al., *Mechanical and thermal allodynia in chronic central pain following spinal cord injury.* Pain, 1996. 68(1): p. 97-107.
53. Kim, J., et al., *Cold and mechanical allodynia in both hind paws and tail following thoracic spinal cord hemisection in rats: Time courses and their correlates.* Neurosci Lett, 2003. 343(3): p. 200-4.
54. Klit, H., N.B. Finnerup, and T.S. Jensen, *Central post-strokepain: clinicalcharacteristics, pathophysiology, andmanagement.*LancetNeurol, 2009. (8): p. 857–868.
55. Lu, H.F., et al., *A newcentral post-strokepainrat model: autologousbloodinjectedthalamichemorrhageinvolvedincreasedexpression of P2X4 receptor.*Neurosci. Lett, 2018. 687: p. 124–130.
56. Gritsch, S., et al., *Functionalcharacterization of a mouse model forcentral post-strokepain.* MolecularPain, 2016. 12: 1744806916629049.
57. Dickinson, B. D., et al., *Maldynia: Pathophysiologyandmanagement of neuropathicandmaladaptivepain—A report of the AMA Council on ScienceandPublicHealth.* PainMedicine, 2010. *11*(11): p. 1635-1653.
58. Authier, N., et al., *Assessment of allodynia and hyperalgesia after cisplatin administration to rats.*Neurosci Lett, 2000. 291: p. 73-76.
59. Pennypacker, S.D., et al., *Methodsandprotocolsforchemotherapy-inducedperipheralneuropathy (CIPN) mousemodelsusingpaclitaxel.* Methods in Cell Biology, AcademicPressInc., 2022.
60. Aley, K.O., D.B. Reichling, and J.D. Levine, *Vincristinehyperalgesia in therat: a model of painfulvincristineneuropathy in humans.* Neuroscience, 1996. 73: p. 259–265.
61. Authier, N., et al., *Description of a short-termtaxol-inducednociceptiveneuropathy in rats.* Brain Res, 2000. 887: p. 239–249.
62. Ledeboer, A., et al., *Intrathecal interleukin-10 gene therapyattenuatespaclitaxelinducedmechanicalallodyniaandproinflammatorycytokineexpression in dorsalrootganglia in rats.* Brain BehavImmun, 2007. 21(5): p. 686-98.
63. Flatters, S.J., and G.J. Bennett, *Studies of peripheralsensorynerves in paclitaxelinducedpainfulperipheralneuropathy: Evidenceformitochondrialdysfunction.* Pain, 2006. 122(3): p. 245-57.
64. Polomano, R.C., et al., *A painfulperipheralneuropathy in theratproducedbythechemotherapeuticdrug, paclitaxel.*Pain, 2001. 94(3): p. 293-304.
65. Ledeboer, A., et al., *Intrathecal interleukin-10 gene therapyattenuatespaclitaxel-inducedmechanicalallodyniaandproinflammatorycytokineexpression in dorsalrootganglia in rats.* Brain BehavImmun, 2007. 21(5): p. 686-98.
66. Argyriou, A.A., et al., *Chemotherapy-inducedperipheralneuropathy in adults: A comprehensiveupdate of the literatüre.* CancerManagRes, 2014. 6: p. 135-47.
67. Cece, R., et al., *An ultrastructuralstudy of neuronalchanges in dorsalrootganglia (DRG) of ratsafterchroniccisplatinadministrations.*HistolHistopathol, 1995. 10(4): p. 837-45
68. Vera, G., et al., *WIN 55,212-2 preventsmechanicalallodynia but not alterations in feedingbehaviourinducedbychroniccisplatin in therat.* Life Sci, 2007. 81(6): p.468-79.
69. Grolleau, F., et al., *A possibleexplanationfor a neurotoxiceffect of theanticanceragentoxaliplatin on neuronalvoltage-gatedsodiumchannels.* Journal of Neurophysiology, 2001. 85(5): p. 2293-2297.
70. Joseph, E.K., et al., *Sexualdimorphismfor protein kinase C epsilon signaling in a rat model of vincristine-inducedpainfulperipheralneuropathy.*Neuroscience, 2003. 119(3): p. 831-8.
71. Tanner, K.D., J.D. Levine, and K.S. Topp, *Microtubuledisorientationandaxonalswelling in unmyelinatedsensoryaxonsduringvincristine-inducedpainfulneuropathy in rat.* J Comp-Neurol, 1998. 395(4): p. 481-92.
72. Sweitzer, S.M., J.L. Pahl, and J.A. DeLeo, *Propentofyllineattenuatesvincristine-inducedperipheralneuropathy in therat.* NeurosciLett, 2006. 400(3): p. 258-61.
73. Tappe-Theodor, A., and R. Kuner, Studyingongoingandspontaneouspain in rodents–challengesandopportunities. Eur. J. Neurosci. 2014. 39: p. 1881–1890.

74. Bhangoo, S.K., et al., *CXCR4 chemokinereceptorsignalingmediatespainhypersensitivity in associationwithantiretroviraltoxicneuropathy.* Brain Behav. Immun, 2007. 21: p. 581–591.
75. Hicks, C.W., and E. Selvin, *Epidemiology of peripheralneuropathyandLowerextremitydisease in diabetes.* Curr. Diab. Rep, 2019. 19: p. 86.
76. Biessels, G. J., et al., *Phenotypinganimalmodels of diabeticneuropathy: a consensusstatement of thediabeticneuropathystudygroup of the EASD (Neurodiab).* Journal of thePeripheralNervousSystem, 2014. 19(2): p. 77-87.
77. Olukman, M., et al., *Treatmentwith NADPH oxidaseinhibitorapocyninalleviatesdiabeticneuropathicpain in rats.*NeuralRegenRes, 2018. 13(9): p. 1657-64.
78. Aley, K.O.,and J.D. Levine, *Rapidonsetpaininducedbyintravenousstreptozotocin in therat.* J Pain, 2001. 2(3): p. 146-50.
79. Zhu, Y.F., et al., *Rat model of cancer-induced bone pain: changes in nonnociceptivesensoryneurons in vivo.* Painreports, 2017. 2(4): p. e603.
80. Vecht, C.J., *Cancerpain: a neurologicalperspective.*Curr. Opin. Neurol, 2000. 13: p. 649–653.
81. Shimoyama, M., et al., *A mouse model of neuropathiccancerpain.* Pain, 2002. 99: p. 167–174.
82. Elramah, S., et al., *Spinal miRNA-124 regulatessynaptopodinandnociception in an animal model of bone cancerpain. Scientificreports*, 2017. *7*(1): p. 1-13.
83. Bandera, A., et al., HIV-associatedneurocognitiveimpairment in the modern ART era: areweclosetodiscoveringreliablebiomarkers in thesetting of virologicalsuppression?. *Frontiers in agingneuroscience*, 2019. 187.
84. Burdo, T.H., and A.D. Miller, *Animalmodels of HIV peripheralneuropathy.*FutureVirol, 2014. 9: p. 465–474.
85. Herzberg, U.,and J. Sagen,*Peripheralnerveexposureto HIV viralenvelope protein gp120 inducesneuropathicpainandspinalgliosis.* J Neuroimmunol, 2001. 116(1): p. 29-39.
86. Cao, L., et al., *Murineimmunodeficiencyvirus-inducedperipheralneuropathyandtheassociatedcytokineresponses.* J. Immunol, 2012. 189: p. 3724–3733
87. Valek, L., G. Auburger, and I. Tegeder, *Sensoryneuropathyandnociception in rodentmodels of Parkinson'sdisease.* DiseaseModels&Mechanisms, 2019. *12*(6): p. dmm039396.
88. Faivre, F., et al., *Thehiddenside of Parkinson'sdisease: Studyingpain, anxietyanddepression in animalmodels. Neuroscience&BiobehavioralReviews*, 2019. *96*: p. 335-352.
89. Charles, K. A., et al., *Alteration of nociceptiveintegration in thespinalcord of a rat model of Parkinson'sdisease.* MovementDisorders, 2018. *33*(6): p. 1010-1015.
90. Dina, O.A., et al., *Key role forthe epsilon isoform of protein kinase C in painfulalcoholicneuropathy in therat.* J Neurosci, 2000. 20(22): p. 8614-9.
91. Sereda, M.W., *Therapeuticadministration of progesterone antagonist in a model of Charcot-Marie-Toothdisease (CMT-1A).* NatMed, 2003. 9: p. 1533–1537.
92. MeyerzuHorste, G, et al., *Antiprogesteronetherapyuncouplesaxonallossfromdemyelination in a transgenicrat model of CMT1A neuropathy.*AnnNeurol, 2007. 61: p. 61–72.
93. Kankowski, S., C. Grothe, and K. Haastert-Talini, *Neuropathicpain: Spotlightinganatomy, experimentalmodels, mechanisms, andtherapeuticaspects.* EuropeanJournal of Neuroscience, 2021. *54*(2): p. 4475-4496.
94. Sewell, R.D., *Neuropathic pain models and outcome measures: a dual translational challenge.* Annals of Translational Medicine, 2018. *6*(Suppl 1).
95. Deuis, J. R., et al., Methods *used to evaluate pain behaviors in rodents.* Frontiers in molecular neuroscience, 2017: 284.
96. Tjolsen, A., et al., *An improved method for tail-flick testing with adjustment for tail-skin temperature.* J Neurosci Methods, 1989. 26: p. 259-265.
97. Dzoyem, J. P., et al., *Anti-inflammatory and anti-nociceptive activities of African medicinal spices and vegetables. In Medicinal spices and vegetables from Africa.* Academic Press, 2017: p. 239-270.
98. Bannon, A.W., and A.B., Malmberg,. *Models of nociception: hot-plate, tail-flick, and formalin tests in rodents.* Current Protocols in Neuroscience, 2007. 41(1): p. 1- 16.

DENEYSEL PARKİNSON HASTALIĞI HAYVAN MODELLERİ

BÖLÜM 6

Dilara NEMUTLU SAMUR[1]

Giriş

Parkinson hastalığı (PH); dopaminerjik sistemde meydana gelen nörodejenerasyon sonucu istirahat tremoru, rijidite ve bradikinezi gibi klasik üçlü motor belirti ile karakterize bir hareket hastalığı olarak tanımlanmaktadır (1). Ancak, günümüzde serotonerjik, kolinerjik ve noradrenerjik sistemler gibi diğer nörotransmitter sistemlerin de bu nörodejeneratif süreçten etkilendiği ve motor olmayan belirtilerin de hastalık tablosuna eşlik ettiği bilinmektedir (2). PH'nin prevalansının genellikle 100.000 kişide 100 ile 200 arasında değiştiği kabul edilmektedir ve yıllık insidansın 15/100.000 olduğu düşünülmektedir (3). Durmuş ve ark. tarafından yapılan Türkiye'de PH prevalansını belirlemeye yönelik ilk büyük popülasyon tabanlı çalışmada, PH prevalansı 202/100.000 bulunmuştur (4). PH'nin karakteristik özellikleri, SNpc'de nöromelanin pigmenti içeren nöronlarda meydana gelen nöron kaybı ve yaygın hücre içi protein (α-sinüklein) birikimi ile meydana gelen yuvarlak, hiyalin nöronal sitoplazmik inklüzyonların (Lewy cisimcikleri) varlığıdır (2). Değişen başlangıç yaşı, semptomlar ve ilerleme hızına sahip heterojen bir hastalık olan PH'de, hastalığın farklı yönlerini incelemek için çeşitli hayvan modellerinin kullanılması gerekir (5). PH'yi deney hayvanlarında modellemek için üç ana yaklaşım kullanılır: (i) nörotoksinler, (ii) genetik manipülasyonlar ve (iii) transkripsiyon faktörlerini hedef alan stratejiler (5). Bu kitap bölümünde PH'nin motor ve motor olmayan semptomlarını taklit etmede başarılı modeller olarak kullanılan nörotoksin modelleri ele alınmıştır.

[1] Dr. Öğr. Üyesi, Alanya Alaaddin Keykubat Üniversitesi Tıp Fakültesi, Tıbbi Farmakoloji AD., dilaranemutlu@gmail.com

Sonuç ve Öneriler

Nörotoksin modelleri hem uygulamaları hem de önceki çalışmalarla karşılaştırmaları kolay olduğu için oldukça değerlidir. 6-OHDA ve MPTP, deney hayvanlarında dopaminerjik nigrostriatal yolakta seçici ve etkili dejenerasyonunu indüklemek için en sık kullanılan modellerdir. Bunun yanı sıra rotenon gibi pestisit modelleri de hem nigral hem de ekstranigral patolojiyi kolaylıkla taklit edebildiği için değerlidir. Bir başka pestisit ve herbisit modeli olan parakuat/maneb etki mekanizması tam olarak aydınlatılamamış ve daha az sıklıkta kullanılan modellerdendir. Nörotoksin modellerinin her gibi çeşitli avantajlara sahip olmakla birlikte bu nörotoksin modellerinin hiçbiri PH'nin tüm patolojik özelliklerini taklit edemez. Bu nedenle test edilmek istenen hipoteze uygun avantajları olan nörotoksin PH modeli oluşturmak için seçilmelidir. Nörotoksin modelleri ile yapılan çalışmalar nöroprotektif tedavilerin geliştirilmesine veya PH patogenezinin aydınlatılmasına katkıda bulunabilir.

Kaynaklar

1. Schapira, A.H.V., K.R. Chaudhuri, and P. Jenner, *Non-motor features of Parkinson disease.* Nat Rev Neurosci, 2017. 18(7): p. 435-450.
2. Poewe, W., et al., *Parkinson disease.* Nature Reviews Disease Primers, 2017. 3(1): p. 17013.
3. Tysnes, O.B. and A. Storstein, *Epidemiology of Parkinson's disease.* J Neural Transm (Vienna), 2017. 124(8): p. 901-905.
4. Durmus, H., M.A. Gokalp, and H.A. Hanagasi, *Prevalence of Parkinson's disease in Baskale, Turkey: a population based study.* Neurol Sci, 2015. 36(3): p. 411-3.
5. Konnova, E.A. and M. Swanberg, *Animal Models of Parkinson's Disease*, in *Parkinson's Disease: Pathogenesis and Clinical Aspects*, T.B. Stoker and J.C. Greenland, Editors. 2018, Codon Publications: Brisbane (AU).
6. Kin, K., et al., *Animal Models for Parkinson's Disease Research: Trends in the 2000s.* Int J Mol Sci, 2019. 20(21).
7. Kalita, J. and U.K. Misra, *Management of advanced Parkinson disease: Still a riddle.* Neurol India, 2018. 66(6): p. 1853-1854.
8. Raudino, F., *The Parkinson disease before James Parkinson.* Neurol Sci, 2012. 33(4): p. 945-8.
9. Jankovic, J., *Parkinson's disease: clinical features and diagnosis.* J Neurol Neurosurg Psychiatry, 2008. 79(4): p. 368-76.
10. Del Rey, N.L.-G., et al., *Advances in Parkinson's Disease: 200 Years Later.* Frontiers in Neuroanatomy, 2018. 12(113).
11. Halliday, G.M., et al., *The neurobiological basis of cognitive impairment in Parkinson's disease.* Movement disorders : official journal of the Movement Disorder Society, 2014. 29(5): p. 634-650.
12. Sanjari Moghaddam, H., et al., *Neurotransmission systems in Parkinson's disease.* Rev Neurosci, 2017. 28(5): p. 509-536.
13. Massano, J. and K.P. Bhatia, *Clinical approach to Parkinson's disease: features, diagnosis, and principles of management.* Cold Spring Harb Perspect Med, 2012. 2(6): p. a008870.

14. Biundo, R., L. Weis, and A. Antonini, *Cognitive decline in Parkinson's disease: the complex picture.* npj Parkinson's Disease, 2016. 2(1): p. 16018.
15. Gubellini, P. and P. Kachidian, *Animal models of Parkinson's disease: An updated overview.* Rev Neurol (Paris), 2015. 171(11): p. 750-61.
16. Aarsland, D., et al., *Cognitive decline in Parkinson disease.* Nat Rev Neurol, 2017. 13(4): p. 217-231.
17. Benskey, M.J., R.G. Perez, and F.P. Manfredsson, *The contribution of alpha synuclein to neuronal survival and function - Implications for Parkinson's disease.* J Neurochem, 2016. 137(3): p. 331-59.
18. Bellucci, A., et al., *alpha-Synuclein synaptic pathology and its implications in the development of novel therapeutic approaches to cure Parkinson's disease.* Brain Res, 2012. 1432: p. 95-113.
19. Buhlman, L.M., *Parkin loss-of-function pathology: Premature neuronal senescence induced by high levels of reactive oxygen species?* Mech Ageing Dev, 2016.
20. Lopes da Fonseca, T., A. Villar-Pique, and T.F. Outeiro, *The Interplay between Alpha-Synuclein Clearance and Spreading.* Biomolecules, 2015. 5(2): p. 435-71.
21. Ciechanover, A. and Y.T. Kwon, *Degradation of misfolded proteins in neurodegenerative diseases: therapeutic targets and strategies.* Exp Mol Med, 2015. 47: p. e147.
22. Perier, C. and M. Vila, *Mitochondrial biology and Parkinson's disease.* Cold Spring Harb Perspect Med, 2012. 2(2): p. a009332.
23. Franco-Iborra, S., M. Vila, and C. Perier, *The Parkinson Disease Mitochondrial Hypothesis: Where Are We at?* Neuroscientist, 2016. 22(3): p. 266-77.
24. Luo, Y., et al., *Mitochondria: A Therapeutic Target for Parkinson's Disease?* Int J Mol Sci, 2015. 16(9): p. 20704-30.
25. Williams, A., *MPTP parkinsonism.* Br Med J (Clin Res Ed), 1984. 289(6456): p. 1401-2.
26. Greenamyre, J.T., R. Betarbet, and T.B. Sherer, *The rotenone model of Parkinson's disease: genes, environment and mitochondria.* Parkinsonism Relat Disord, 2003. 9 Suppl 2: p. S59-64.
27. Ghavami, S., et al., *Autophagy and apoptosis dysfunction in neurodegenerative disorders.* Prog Neurobiol, 2014. 112: p. 24-49.
28. Gonzalez-Burgos, E., C. Fernandez-Moriano, and M.P. Gomez-Serranillos, *Potential neuroprotective activity of Ginseng in Parkinson's disease: a review.* J Neuroimmune Pharmacol, 2015. 10(1): p. 14-29.
29. Wang, Q., Y. Liu, and J. Zhou, *Neuroinflammation in Parkinson's disease and its potential as therapeutic target.* Transl Neurodegener, 2015. 4: p. 19.
30. Rodriguez-Nogales, C., et al., *Brain aging and Parkinson's disease: New therapeutic approaches using drug delivery systems.* Maturitas, 2016. 84: p. 25-31.
31. Foltynie, T., C. Brayne, and R.A. Barker, *The heterogeneity of idiopathic Parkinson's disease.* Journal of Neurology, 2002. 249(2): p. 138-145.
32. Jagmag, S.A., et al., *Evaluation of Models of Parkinson's Disease.* Front Neurosci, 2015. 9: p. 503.
33. Meredith, G.E., P.K. Sonsalla, and M.F. Chesselet, *Animal models of Parkinson's disease progression.* Acta Neuropathol, 2008. 115(4): p. 385-98.
34. Dawson, T.M., H.S. Ko, and V.L. Dawson, *Genetic animal models of Parkinson's disease.* Neuron, 2010. 66(5): p. 646-61.
35. Potashkin, J.A., S.R. Blume, and N.K. Runkle, *Limitations of animal models of Parkinson's disease.* Parkinsons Dis, 2010. 2011: p. 658083.
36. Barker, R.A. and A. Björklund, *Animal Models of Parkinson's Disease: Are They Useful or Not?* J Parkinsons Dis, 2020. 10(4): p. 1335-1342.
37. Tieu, K., *A guide to neurotoxic animal models of Parkinson's disease.* Cold Spring Harb Perspect Med, 2011. 1(1): p. a009316.
38. Jackson-Lewis, V., J. Blesa, and S. Przedborski, *Animal models of Parkinson's disease.* Parkinsonism & Related Disorders, 2012. 18: p. S183-S185.

39. Moore, D.J. and T.M. Dawson, *Value of genetic models in understanding the cause and mechanisms of Parkinson's disease.* Curr Neurol Neurosci Rep, 2008. 8(4): p. 288-96.
40. Zeng, X.-S., W.-S. Geng, and J.-J. Jia, *Neurotoxin-Induced Animal Models of Parkinson Disease: Pathogenic Mechanism and Assessment.* ASN neuro, 2018. 10: p. 1759091418777438-1759091418777438.
41. Costello, S., et al., *Parkinson's disease and residential exposure to maneb and paraquat from agricultural applications in the central valley of California.* Am J Epidemiol, 2009. 169(8): p. 919-26.
42. Lane, E.L., et al., *Animal Models of Parkinson's Disease*, in *Sourcebook of Models for Biomedical Research*, P.M. Conn, Editor. 2008, Humana Press: Totowa, NJ. p. 313-322.
43. Blesa, J. and S. Przedborski, *Parkinson's disease: animal models and dopaminergic cell vulnerability.* Front Neuroanat, 2014. 8: p. 155.
44. Duty, S. and P. Jenner, *Animal models of Parkinson's disease: a source of novel treatments and clues to the cause of the disease.* British journal of pharmacology, 2011. 164(4): p. 1357-1391.
45. Rodríguez Díaz, M., et al., *Motor behavioural changes after intracerebroventricular injection of 6-hydroxydopamine in the rat: an animal model of Parkinson's disease.* Behav Brain Res, 2001. 122(1): p. 79-92.
46. Simola, N., M. Morelli, and A.R. Carta, *The 6-hydroxydopamine model of Parkinson's disease.* Neurotox Res, 2007. 11(3-4): p. 151-67.
47. Masini, D., et al., *A Guide to the Generation of a 6-Hydroxydopamine Mouse Model of Parkinson's Disease for the Study of Non-Motor Symptoms.* Biomedicines, 2021. 9(6).
48. Torres, E. and S. Dunnett, *6-OHDA Lesion Models of Parkinson's Disease in the Rat.* Neuromethods, 2011. 61: p. 267-279.
49. Guillaumin, A., B. Vlcek, and Å. Wallén-Mackenzie, *Improving Well-Being and Survival in the 6-OHDA Lesion Model of Parkinson´s Disease in Mice: Step-By-Step Protocol.* 2021, Research Square.
50. Delaville, C., et al., *Emerging dysfunctions consequent to combined monoaminergic depletions in Parkinsonism.* Neurobiol Dis, 2012. 45(2): p. 763-73.
51. Prasad, E.M. and S.-Y. Hung, *Behavioral Tests in Neurotoxin-Induced Animal Models of Parkinson's Disease.* Antioxidants (Basel, Switzerland), 2020. 9(10): p. 1007.
52. Davis, G.C., et al., *Chronic Parkinsonism secondary to intravenous injection of meperidine analogues.* Psychiatry Res, 1979. 1(3): p. 249-54.
53. Fuller, R.W., *MPTP: A Parkinsonism-causing Neurotoxic Substance*, in *Current Aspects of the Neurosciences: Volume 1*, N.N. Osborne, Editor. 1990, Macmillan Education UK: London. p. 183-201.
54. Langston, J.W., et al., *Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis.* Science, 1983. 219(4587): p. 979-80.
55. Sonsalla, P.K. and R.E. Heikkila, *The influence of dose and dosing interval on MPTP-induced dopaminergic neurotoxicity in mice.* Eur J Pharmacol, 1986. 129(3): p. 339-45.
56. Meredith, G.E. and D.J. Rademacher, *MPTP mouse models of Parkinson's disease: an update.* Journal of Parkinson's disease, 2011. 1(1): p. 19-33.
57. Hirsch, E.C., S. Vyas, and S. Hunot, *Neuroinflammation in Parkinson's disease.* Parkinsonism & Related Disorders, 2012. 18: p. S210-S212.
58. Rodríguez-Cruz, A., et al., *"Effect of valerenic acid on neuroinflammation in a MPTP-induced mouse model of Parkinson's disease".* IBRO Reports, 2020. 8: p. 28-35.
59. Sonsalla, P.K., G.D. Zeevalk, and D.C. German, *Chronic intraventricular administration of 1-methyl-4-phenylpyridinium as a progressive model of Parkinson's disease.* Parkinsonism & Related Disorders, 2008. 14: p. S116-S118.
60. Meredith, G.E. and U.J. Kang, *Behavioral models of Parkinson's disease in rodents: a new look at an old problem.* Mov Disord, 2006. 21(10): p. 1595-606.
61. Machado, V., et al., *Microglia-Mediated Neuroinflammation and Neurotrophic Factor-Induced Protection in the MPTP Mouse Model of Parkinson's Disease-Lessons from Transgenic Mice.* International journal of molecular sciences, 2016. 17(2): p. 151.

62. Petroske, E., et al., *Mouse model of Parkinsonism: a comparison between subacute MPTP and chronic MPTP/probenecid treatment.* Neuroscience, 2001. 106(3): p. 589-601.
63. Goldberg, N.R., et al., *Dopaminergic and behavioral correlates of progressive lesioning of the nigrostriatal pathway with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine.* Neuroscience, 2011. 180: p. 256-71.
64. Petzinger, G.M., et al., *Using the MPTP Mouse Model to Understand Neuroplasticity: A New Therapeutic Target for Parkinson's Disease?*, in *Animal Models of Movement Disorders: Volume I*, E.L. Lane and S.B. Dunnett, Editors. 2012, Humana Press: Totowa, NJ. p. 353-369.
65. Huang, D., et al., *Dynamic Changes in the Nigrostriatal Pathway in the MPTP Mouse Model of Parkinson's Disease.* Parkinsons Dis, 2017. 2017: p. 9349487.
66. Sun, M.F., et al., *Neuroprotective effects of fecal microbiota transplantation on MPTP-induced Parkinson's disease mice: Gut microbiota, glial reaction and TLR4/TNF-α signaling pathway.* Brain Behav Immun, 2018. 70: p. 48-60.
67. Fornai, F., et al., *Parkinson-like syndrome induced by continuous MPTP infusion: convergent roles of the ubiquitin-proteasome system and alpha-synuclein.* Proc Natl Acad Sci U S A, 2005. 102(9): p. 3413-8.
68. McCormack, A.L., et al., *Environmental risk factors and Parkinson's disease: selective degeneration of nigral dopaminergic neurons caused by the herbicide paraquat.* Neurobiol Dis, 2002. 10(2): p. 119-27.
69. Manning-Bog, A.B., et al., *The herbicide paraquat causes up-regulation and aggregation of alpha-synuclein in mice: paraquat and alpha-synuclein.* J Biol Chem, 2002. 277(3): p. 1641-4.
70. Uversky, V.N., *Neurotoxicant-induced animal models of Parkinson's disease: understanding the role of rotenone, maneb and paraquat in neurodegeneration.* Cell Tissue Res, 2004. 318(1): p. 225-41.
71. Bastías-Candia, S., et al., *Combined exposure to agriculture pesticides, paraquat and maneb, induces alterations in the N/OFQ-NOPr and PDYN/KOPr systems in rats: Relevance to sporadic Parkinson's disease.* Environ Toxicol, 2015. 30(6): p. 656-63.
72. Takahashi, R.N., R. Rogerio, and M. Zanin, *Maneb enhances MPTP neurotoxicity in mice.* Res Commun Chem Pathol Pharmacol, 1989. 66(1): p. 167-70.
73. Cicchetti, F., et al., *Systemic exposure to paraquat and maneb models early Parkinson's disease in young adult rats.* Neurobiol Dis, 2005. 20(2): p. 360-71.
74. Wang, K., et al., *Taurine improves neuron injuries and cognitive impairment in a mouse Parkinson's disease model through inhibition of microglial activation.* NeuroToxicology, 2021. 83: p. 129-136.
75. Thiruchelvam, M., et al., *Age-related irreversible progressive nigrostriatal dopaminergic neurotoxicity in the paraquat and maneb model of the Parkinson's disease phenotype.* Eur J Neurosci, 2003. 18(3): p. 589-600.
76. Saint-Pierre, M., et al., *Temporal effects of paraquat/maneb on microglial activation and dopamine neuronal loss in older rats.* J Neurochem, 2006. 98(3): p. 760-72.
77. Vegh, C., et al., *Combined Ubisol-Q(10) and Ashwagandha Root Extract Target Multiple Biochemical Mechanisms and Reduces Neurodegeneration in a Paraquat-Induced Rat Model of Parkinson's Disease.* Antioxidants (Basel, Switzerland), 2021. 10(4): p. 563.
78. Fathy, S.M., H.A. El-Dash, and N.I. Said, *Neuroprotective effects of pomegranate (Punica granatum L.) juice and seed extract in paraquat-induced mouse model of Parkinson's disease.* BMC complementary medicine and therapies, 2021. 21(1): p. 130-130.
79. Prasad, E.M. and S.Y. Hung, *Behavioral Tests in Neurotoxin-Induced Animal Models of Parkinson's Disease.* Antioxidants (Basel), 2020. 9(10).
80. Cristóvão, A.C., et al., *Characterization of a Parkinson's disease rat model using an upgraded paraquat exposure paradigm.* Eur J Neurosci, 2020. 52(4): p. 3242-3255.

81. Ivanov, M.V., et al., *Immunomorphological Changes in Neuronal and Non-Neuronal Structures in the Rat Intestine in a Toxin-Induced Model of Parkinsonism.* Bulletin of Experimental Biology and Medicine, 2021. 171(1): p. 94-99.
82. Anselmi, L., et al., *Ingestion of subthreshold doses of environmental toxins induces ascending Parkinsonism in the rat.* NPJ Parkinsons Dis, 2018. 4: p. 30.
83. Chen, Y.-B., et al., *A novel idea for establishing Parkinson's disease mouse model by intranasal administration of paraquat.* Neurological Research, 2021. 43(4): p. 267-277.
84. Vijayanathan, Y., et al., *Adult Endogenous Dopaminergic Neuroregeneration Against Parkinson's Disease: Ideal Animal Models?* Neurotox Res, 2021. 39(2): p. 504-532.
85. Wills, J., et al., *Paraquat, but Not Maneb, Induces Synucleinopathy and Tauopathy in Striata of Mice through Inhibition of Proteasomal and Autophagic Pathways.* PLOS ONE, 2012. 7(1): p. e30745.
86. Schmidt, W.J. and M. Alam, *Controversies on new animal models of Parkinson's disease pro and con: the rotenone model of Parkinson's disease (PD).* J Neural Transm Suppl, 2006(70): p. 273-6.
87. Betarbet, R., et al., *Chronic systemic pesticide exposure reproduces features of Parkinson's disease.* Nat Neurosci, 2000. 3(12): p. 1301-6.
88. Höglinger, G.U., et al., *Chronic systemic complex I inhibition induces a hypokinetic multisystem degeneration in rats.* J Neurochem, 2003. 84(3): p. 491-502.
89. Xiong, N., et al., *Stereotaxical infusion of rotenone: a reliable rodent model for Parkinson's disease.* PLoS One, 2009. 4(11): p. e7878.
90. Mansour, R.M., et al., *Montelukast attenuates rotenone-induced microglial activation/p38 MAPK expression in rats: Possible role of its antioxidant, anti-inflammatory and antiapoptotic effects.* Toxicol Appl Pharmacol, 2018. 358: p. 76-85.
91. Kandil, E.A., et al., *Hypoxia-inducible factor 1 alpha and nuclear-related receptor 1 as targets for neuroprotection by albendazole in a rat rotenone model of Parkinson's disease.* Clin Exp Pharmacol Physiol, 2019. 46(12): p. 1141-1150.
92. Zhang, X., et al., *Therapeutic effects of baicalein on rotenone-induced Parkinson's disease through protecting mitochondrial function and biogenesis.* Scientific Reports, 2017. 7(1): p. 9968.
93. Pan-Montojo, F., et al., *Progression of Parkinson's disease pathology is reproduced by intragastric administration of rotenone in mice.* PLoS One, 2010. 5(1): p. e8762.
94. Alam, M., A. Mayerhofer, and W.J. Schmidt, *The neurobehavioral changes induced by bilateral rotenone lesion in medial forebrain bundle of rats are reversed by L-DOPA.* Behav Brain Res, 2004. 151(1-2): p. 117-24.
95. Cannon, J.R., et al., *A highly reproducible rotenone model of Parkinson's disease.* Neurobiol Dis, 2009. 34(2): p. 279-90.
96. Toyoda, H., et al., *Intranasal Administration of Rotenone Reduces GABAergic Inhibition in the Mouse Insular Cortex Leading to Impairment of LTD and Conditioned Taste Aversion Memory.* Int J Mol Sci, 2020. 22(1).
97. Sasajima, H., et al., *Intranasal administration of rotenone in mice attenuated olfactory functions through the lesion of dopaminergic neurons in the olfactory bulb.* Neurotoxicology, 2015. 51: p. 106-15.
98. Sasajima, H., et al., *Intranasal Administration of Rotenone to Mice Induces Dopaminergic Neurite Degeneration of Dopaminergic Neurons in the Substantia Nigra.* Biol Pharm Bull, 2017. 40(1): p. 108-112.
99. Murakami, S., et al., *Long-Term Systemic Exposure to Rotenone Induces Central and Peripheral Pathology of Parkinson's Disease in Mice.* Neurochem Res, 2015. 40(6): p. 1165-78.
100. Chiu, C.C., et al., *Neuroprotective effects of aldehyde dehydrogenase 2 activation in rotenone-induced cellular and animal models of parkinsonism.* Exp Neurol, 2015. 263: p. 244-53.

101. Miyazaki, I. and M. Asanuma, *The Rotenone Models Reproducing Central and Peripheral Features of Parkinson's Disease.* NeuroSci, 2020. 1(1): p. 1-14.
102. von Wrangel, C., et al., *The rotenone-induced rat model of Parkinson's disease: behavioral and electrophysiological findings.* Behav Brain Res, 2015. 279: p. 52-61.
103. Johnson, M.E. and L. Bobrovskaya, *An update on the rotenone models of Parkinson's disease: Their ability to reproduce the features of clinical disease and model gene–environment interactions.* NeuroToxicology, 2015. 46: p. 101-116.

DENEYSEL SEREBRAL İSKEMİ MODELLERİ

BÖLÜM 7

Güven AKÇAY[1]

Giriş

Beynimiz, görme, koklama, yürüme, koşma, düşünme, hissetme, akıl yürütme, dikkat ve hafıza gibi tüm bilinçli ve bilinçaltı fizyolojik fonksiyonlardan sorumlu organımızdır. Beyin uyku, uyanıklık, bilişsel ve fiziksel aktiviteleri yerine getirebilmek için gerekli düzeyde sürekli olarak oksijene ve enerjiye ihtiyaç duymaktadır. Beyin, vücut ağırlığının % 2'i kadarı olmasına karşın kardiyak kalp debisinin yaklaşık % 20'sini kullanmaktadır. Beyin beslenmesinde serebral kan akımı hayati önem taşımaktadır. Beynin bir bölgesinde veya tümünde serebral kan akımının azalması sonucu serebral iskemi meydana gelmektedir. Serebral iskemi, beyni besleyen damarların tıkanması veya kanaması sonucu oluşmaktadır. Dünyada her yıl yaklaşık 17 milyon inme vakası görülürken, ülkemizde de yaklaşık olarak 132.000 inme vakası görülmektedir. Her geçen yıl inme vakaları artmakta ve gelecekte bu durumun sağlıkla ve ekonomiyle ilgili ciddi sorunlara yol açacağı öngörülmektedir. Bundan dolayı inmenin önlenmesi ve etkin tedavi yöntemlerinin uygulanması hayati önem taşımaktadır. Bu tedavi yöntemlerinin araştırılmasında yeni tedavi protokollerinin geliştirilmesi ve yeni ajanların keşfedilmesinde deneysel hayvan modelleri sıklıkla tercih edilmektedir. Klinikteki serebral iskemik vakalarının fizyopatolojisinin araştırılmasında sıklıkla sıçan ve fare gibi kemirgenler üzerinde yapılan geçici global serebral iskemi, geçici fokal serebral iskemi ve geçici ön beyin iskemi modelleri kullanılmaktadır. Bu kitapta; serebral iskeminin epidemiyolojisi, patofizyolojisi ve deneysel serebral iskemi hayvan modelleri arasında en çok kullanılan orta serebral arter oklüzyon yöntemi ile ilgili bilgiler sunulmuştur.

[1] Dr. Öğr. Üyesi, Hitit Üniversitesi Tıp Fakültesi, Biyofizik AD., guvenakcayibu@gmail.com

reperfüzyon aşaması oluşturmakta ve tedavi yöntemleri reperfüzyon hasarını önlemeye yönelik olmalıdır. Serebral iskeminin tedavi yöntemlerinin gelişmesinde özellikle deneysel çalışmalar büyük katkı sağlamaktadır. Yapılan deneysel patofizyolojik çalışmaların çoğu iskemi sonrası reperfüzyon hasarını önlemek, hastaların iyileşmesini hızlandıracak tedavi yöntemlerin geliştirilmesi üzerine yapılmaktadır İskemi reperfüzyonun tedavisi için yeni ajanların keşfedilmesi ve yeni tedavi protokollerinin geliştirilmesi için deneysel geçici serebral iskemi hayvan modelleri sıklıkla tercih edilmektedir. Bu nedenle deneysel geçici orta serebral oklüzyon modeli serebral iskemi araştırmalarında hayati önem arz etmekte ve yeni tedavi yöntemleri için daha fazla deneysel araştırmalar yapılmalıdır.

Kaynaklar

1. Akçay, G., *Cerebral Ischemia Model Created by Transient Middle Cerebral Artery Occlusion*. Turk Hij Den Biyol Derg, 2021. 78(2): p. 0-0.
2. Akcay, G. and A. Bahadır, *The Therapeutic Effects of Non-Invasive Transcranial Direct Current Stimulation (tDCS) in Experimental Models of Stroke*, in *Research Advancements In Health Scıences*, P.M. Chernopolskı, N.L. Shapekova, and B. Ak, Editors. 2021: St. Klıment Ohrıdskı Unıversıty Press. p. 28-49.
3. Bahadır, A. and G. Akcay, *The Recovery Effects of Non-Invasive Repetitive Transcranial Magnetic Stimulation (rTMS) in Experimental Models of Stroke*, in *Research Advancements In Health Scıences*, P.M. Chernopolskı, N.L. Shapekova, and B. Ak, Editors. 2021: St. Klıment Ohrıdskı Unıversıty Press. p. 2-27.
4. Akcay G., *Deneysel Serebral İskemi Modeline Bağlı Öğrenme ve Hafıza Değişikliklerine Transkraniyal Doğru Akım Stimülasyonunun Etkilerinin Araştırılması*, in *Biyofizik*. 2020, Akdeniz Üniversitesi. p. 102.
5. Kumral, E., *Santral Sinir Sisteminin Damarsal Hastalıkları*. 2011, Güneş Tıp Kitapevleri.
6. Kablan, Y., *İnme: Epidemiyoloji ve Risk Faktörleri*. 2018: Türkiye Klinikleri. p. 1-19.
7. Benjamin, E.J., et al., *Heart Disease and Stroke Statistics-2018 Update: A Report From the American Heart Association.* Circulation, 2018. 137(12): p. e67-e492.
8. Krishnamurthi, R.V., et al., *Stroke Prevalence, Mortality and Disability-Adjusted Life Years in Adults Aged 20-64 Years in 1990-2013: Data from the Global Burden of Disease 2013 Study.* Neuroepidemiology, 2015. 45(3): p. 190-202.
9. Akcay G., Aslan M., and Derin N. *Serebral İskemi Sonrası Motor Fonksiyonların Tedavisinde Transkraniyal Doğru Akım Stimülasyonu Etkileri*. in *18. Ulusal Sinirbilim Kongresi*. 6-9 Kasım 2020. Bilkent Üniversitesi, Ankara (Türkiye).
10. Kaya, D. and Y.G. Özdemir, *Serebral Kan Akımı ve Metabolizması*, in *Santral Sinir Sisteminin Damarsal Hastalıkları*, E. Kumral, Editor. 2011: Güneş Kitapevleri.
11. Astrup, J., *Energy-requiring cell functions in the ischemic brain. Their critical supply and possible inhibition in protective therapy.* J Neurosurg, 1982. 56(4): p. 482-97.
12. Siesjo, B.K. and F. Bengtsson, *Calcium fluxes, calcium antagonists, and calcium-related pathology in brain ischemia, hypoglycemia, and spreading depression: a unifying hypothesis.* J Cereb Blood Flow Metab, 1989. 9(2): p. 127-40.
13. Oğul, E., *Klinik Nöroloji*. Birinci Baskı ed. 2002, Nobrl&Güneş.
14. Sahan, M., et al., *İskemik İnme ve Akut Faz Reaktanları.* Arşiv Kaynak Tarama Dergisi, 2010. 19(2): p. 85-140.

15. Berg, J.M., J.L. Tymoczko, and L. Stryer, *The Glycolytic Pathway Is Tightly Controlled*. 5th edition ed. Biochemistry. 2002.
16. Sims, N.R. and H. Muyderman, *Mitochondria, oxidative metabolism and cell death in stroke.* Biochim Biophys Acta, 2010. 1802(1): p. 80-91.
17. Yager, J.Y., R.M. Brucklacher, and R.C. Vannucci, *Cerebral energy metabolism during hypoxia-ischemia and early recovery in immature rats.* Am J Physiol, 1992. 262(3 Pt 2): p. H672-7.
18. Zhang, Y., et al., *Remifentanil mimics cardioprotective effect of ischemic preconditioning via protein kinase C activation in open chest of rats.* Acta Pharmacol Sin, 2005. 26(5): p. 546-50.
19. Simard, J.M., K.V. Tarasov, and V. Gerzanich, *Non-selective cation channels, transient receptor potential channels and ischemic stroke.* Biochim Biophys Acta, 2007. 1772(8): p. 947-57.
20. Bogousslavsky, J., G. Van Melle, and F. Regli, *The Lausanne Stroke Registry: analysis of 1,000 consecutive patients with first stroke.* Stroke, 1988. 19(9): p. 1083-92.
21. Liang, D., T.M. Dawson, and V.L. Dawson, *What have genetically engineered mice taught us about ischemic injury?* Curr Mol Med, 2004. 4(2): p. 207-25.
22. Shalak, L. and J.M. Perlman, *Hypoxic-ischemic brain injury in the term infant-current concepts.* Early Hum Dev, 2004. 80(2): p. 125-41.
23. Xing, C., et al., *Pathophysiologic cascades in ischemic stroke.* Int J Stroke, 2012. 7(5): p. 378-85.
24. Chan, P.H., *Role of oxidants in ischemic brain damage.* Stroke, 1996. 27(6): p. 1124-9.
25. Simonian, N.A. and J.T. Coyle, *Oxidative stress in neurodegenerative diseases.* Annu Rev Pharmacol Toxicol, 1996. 36: p. 83-106.
26. Nicotera, P. and S.A. Lipton, *Excitotoxins in neuronal apoptosis and necrosis.* J Cereb Blood Flow Metab, 1999. 19(6): p. 583-91.
27. Yamashima, T., *Implication of cysteine proteases calpain, cathepsin and caspase in ischemic neuronal death of primates.* Prog Neurobiol, 2000. 62(3): p. 273-95.
28. Hillered, L., et al., *Dynamics of extracellular metabolites in the striatum after middle cerebral artery occlusion in the rat monitored by intracerebral microdialysis.* J Cereb Blood Flow Metab, 1989. 9(5): p. 607-16.
29. Davalos, A., A. Shuaib, and N.G. Wahlgren, *Neurotransmitters and pathophysiology of stroke: evidence for the release of glutamate and other transmitters/mediators in animals and humans.* J Stroke Cerebrovasc Dis, 2000. 9(6 Pt 2): p. 2-8.
30. Lee, J.M., et al., *Brain tissue responses to ischemia.* J Clin Invest, 2000. 106(6): p. 723-31.
31. Yamashima, T., et al., *Sustained calpain activation associated with lysosomal rupture executes necrosis of the postischemic CA1 neurons in primates.* Hippocampus, 2003. 13(7): p. 791-800.
32. Memezawa, H., M.L. Smith, and B.K. Siesjo, *Penumbral tissues salvaged by reperfusion following middle cerebral artery occlusion in rats.* Stroke, 1992. 23(4): p. 552-9.
33. Salford, L.G., F. Plum, and J.B. Brierley, *Graded hypoxia-oligemia in rat brain. II. Neuropathological alterations and their implications.* Arch Neurol, 1973. 29(4): p. 234-8.
34. Belov Kirdajova, D., et al., *Ischemia-Triggered Glutamate Excitotoxicity From the Perspective of Glial Cells.* Front Cell Neurosci, 2020. 14: p. 51.
35. Ghosh, A., et al., *Neuroprotective role of nanoencapsulated quercetin in combating ischemia-reperfusion induced neuronal damage in young and aged rats.* PLoS One, 2013. 8(4): p. e57735.
36. Bouma, G.J., et al., *Ultra-early evaluation of regional cerebral blood flow in severely head-injured patients using xenon-enhanced computerized tomography.* J Neurosurg, 1992. 77(3): p. 360-8.
37. Suarez, J.I., *Outcome in neurocritical care: advances in monitoring and treatment and effect of a specialized neurocritical care team.* Crit Care Med, 2006. 34(9 Suppl): p. S232-8.
38. Yuan, H.J., et al., *Noninvasive delayed limb ischemic preconditioning in rats increases antioxidant activities in cerebral tissue during severe ischemia-reperfusion injury.* J Surg Res, 2012. 174(1): p. 176-83.
39. Li, M.H., et al., *Calcium-permeable ion channels involved in glutamate receptor-independent ischemic brain injury.* Acta Pharmacol Sin, 2011. 32(6): p. 734-40.

40. Valko, M., H. Morris, and M.T. Cronin, *Metals, toxicity and oxidative stress.* Curr Med Chem, 2005. 12(10): p. 1161-208.
41. Ward, J.D., et al., *Failure of prophylactic barbiturate coma in the treatment of severe head injury.* J Neurosurg, 1985. 62(3): p. 383-8.
42. Xiong, L.-z., *Neuroanesthesia and neuroprotection: where are we now?* Chinese Medical Journal, 2006. 119(11): p. 883-886.
43. Sivenius, J., et al., *Continuous 15-year decrease in incidence and mortality of stroke in Finland: the FINSTROKE study.* Stroke, 2004. 35(2): p. 420-5.
44. Gasche, Y., et al., *Early appearance of activated matrix metalloproteinase-9 after focal cerebral ischemia in mice: a possible role in blood-brain barrier dysfunction.* J Cereb Blood Flow Metab, 1999. 19(9): p. 1020-8.
45. Granger, D.N., G. Rutili, and J.M. McCord, *Superoxide radicals in feline intestinal ischemia.* Gastroenterology, 1981. 81(1): p. 22-9.
46. Perlman, J.M., *Intervention strategies for neonatal hypoxic-ischemic cerebral injury.* Clin Ther, 2006. 28(9): p. 1353-65.
47. Behringer, W., et al., *Rapid induction of mild cerebral hypothermia by cold aortic flush achieves normal recovery in a dog outcome model with 20-minute exsanguination cardiac arrest.* Acad Emerg Med, 2000. 7(12): p. 1341-8.
48. Böttiger, B.W., et al., *The cerebral 'no-reflow' phenomenon after cardiac arrest in rats--influence of low-flow reperfusion.* Resuscitation, 1997. 34(1): p. 79-87.
49. Hossmann, K.A., *Cerebral ischemia: models, methods and outcomes.* Neuropharmacology, 2008. 55(3): p. 257-70.
50. Kofler, J., et al., *Histopathological and behavioral characterization of a novel model of cardiac arrest and cardiopulmonary resuscitation in mice.* J Neurosci Methods, 2004. 136(1): p. 33-44.
51. Makarenko, N.V., *(The higher nervous activity in dogs revived following prolonged terms of clinical death from drowning or bloodletting).* Zh Vyssh Nerv Deiat Im I P Pavlova, 1972. 22(1): p. 82-8.
52. Pulsinelli, W.A. and J.B. Brierley, *A new model of bilateral hemispheric ischemia in the unanesthetized rat.* Stroke, 1979. 10(3): p. 267-72.
53. Levine, S. and H. Payan, *Effects of ischemia and other procedures on the brain and retina of the gerbil (Meriones unguiculatus).* Exp Neurol, 1966. 16(3): p. 255-62.
54. Holloway, P.M. and F.N. Gavins, *Modeling Ischemic Stroke In Vitro: Status Quo and Future Perspectives.* Stroke, 2016. 47(2): p. 561-9.
55. Goldberg, M.P. and D.W. Choi, *Combined oxygen and glucose deprivation in cortical cell culture: calcium-dependent and calcium-independent mechanisms of neuronal injury.* J Neurosci, 1993. 13(8): p. 3510-24.
56. Matsumoto, K., et al., *Role of pyruvate in ischaemia-like conditions on cultured neurons.* Neurol Res, 1994. 16(6): p. 460-4.
57. Newell, D.W., et al., *Glutamate and non-glutamate receptor mediated toxicity caused by oxygen and glucose deprivation in organotypic hippocampal cultures.* J Neurosci, 1995. 15(11): p. 7702-11.
58. Wolf, P.A., et al., *Secular trends in stroke incidence and mortality. The Framingham Study.* Stroke, 1992. 23(11): p. 1551-5.
59. Kelly, S., J. McCulloch, and K. Horsburgh, *Minimal ischaemic neuronal damage and HSP70 expression in MF1 strain mice following bilateral common carotid artery occlusion.* Brain Res, 2001. 914(1-2): p. 185-95.
60. Herrmann, M., et al., *Effect of inherent epileptic seizures on brain injury after transient cerebral ischemia in Mongolian gerbils.* Exp Brain Res, 2004. 154(2): p. 176-82.
61. Paschen, W., et al., *Biochemical changes during graded brain ischemia in gerbils. Part 2. Regional evaluation of cerebral blood flow and brain metabolites.* J Neurol Sci, 1983. 58(1): p. 37-44.

62. Levine, S., *Anoxic-ischemic encephalopathy in rats.* Am J Pathol, 1960. 36(1): p. 1-17.
63. Overgaard, K., *Thrombolytic therapy in experimental embolic stroke.* Cerebrovasc Brain Metab Rev, 1994. 6(3): p. 257-86.
64. Tsirka, S.E., A.D. Rogove, and S. Strickland, *Neuronal cell death and tPA.* Nature, 1996. 384(6605): p. 123-4.
65. Uzdensky, A.B., *Photothrombotic Stroke as a Model of Ischemic Stroke.* Transl Stroke Res, 2018. 9(5): p. 437-451.
66. Labat-gest, V. and S. Tomasi, *Photothrombotic ischemia: a minimally invasive and reproducible photochemical cortical lesion model for mouse stroke studies.* J Vis Exp, 2013(76).
67. Klebe, D., et al., *Intracerebral Hemorrhage in Mice.* Methods Mol Biol, 2018. 1717: p. 83-91.
68. Manaenko, A., et al., *Comparison of different preclinical models of intracerebral hemorrhage.* Acta Neurochir Suppl, 2011. 111: p. 9-14.
69. Gupta, Y.K. and S. Briyal, *Animal models of cerebral ischemia for evaluation of drugs.* Indian J Physiol Pharmacol, 2004. 48(4): p. 379-94.
70. Chen, J.C., et al., *Down-regulation of the glial glutamate transporter GLT-1 in rat hippocampus and striatum and its modulation by a group III metabotropic glutamate receptor antagonist following transient global forebrain ischemia.* Neuropharmacology, 2005. 49(5): p. 703-14.
71. Liu, F. and L.D. McCullough, *Middle cerebral artery occlusion model in rodents: methods and potential pitfalls.* J Biomed Biotechnol, 2011. 2011: p. 464701.
72. Shahjouei, S., et al., *Middle Cerebral Artery Occlusion Model of Stroke in Rodents: A Step-by-Step Approach.* J Vasc Interv Neurol, 2016. 8(5): p. 1-8.
73. Longa, E.Z., et al., *Reversible middle cerebral artery occlusion without craniectomy in rats.* Stroke, 1989. 20(1): p. 84-91.
74. Lee, S., et al., *Comparison of surgical methods of transient middle cerebral artery occlusion between rats and mice.* J Vet Med Sci, 2014. 76(12): p. 1555-61.
75. Wattanathorn, J., et al., *Cerebroprotective Effect against Cerebral Ischemia of the Combined Extract of Oryza sativa and Anethum graveolens in Metabolic Syndrome Rats.* Oxid Med Cell Longev, 2019. 2019: p. 9658267.
76. Lin, T.N., et al., *Effect of brain edema on infarct volume in a focal cerebral ischemia model in rats.* Stroke, 1993. 24(1): p. 117-21.
77. Schimidt, H.L., et al., *Memory deficits and oxidative stress in cerebral ischemia-reperfusion: neuroprotective role of physical exercise and green tea supplementation.* Neurobiol Learn Mem, 2014. 114: p. 242-50.
78. Sharifi, Z.N., et al., *Neuroprotective Treatment With FK506 Reduces Hippocampal Damage and Prevents Learning and Memory Deficits After Transient Global Ischemia in Rat.* Archives of Neuroscience, 2013. 1: p. 35-40.
79. Braun, R., et al., *Transcranial direct current stimulation accelerates recovery of function, induces neurogenesis and recruits oligodendrocyte precursors in a rat model of stroke.* Exp Neurol, 2016. 279: p. 127-136.
80. Pascual, J.M., et al., *Glutamate, glutamine, and GABA as substrates for the neuronal and glial compartments after focal cerebral ischemia in rats.* Stroke, 1998. 29(5): p. 1048-56; discussion 1056-7.
81. Pulsinelli, W.A., *The therapeutic window in ischemic brain injury.* Curr Opin Neurol, 1995. 8(1): p. 3-5.
82. Pulsinelli, W.A., et al., *Ischemic brain injury and the therapeutic window.* Ann N Y Acad Sci, 1997. 835: p. 187-93.
83. Krzyzanowska, W., et al., *Glutamate transporters in brain ischemia: to modulate or not?* Acta Pharmacol Sin, 2014. 35(4): p. 444-62.
84. Hu, Y., et al., *Increased Susceptibility to Ischemic Brain Injury in Neuroplastin 65-Deficient Mice Likely via Glutamate Excitotoxicity.* Front Cell Neurosci, 2017. 11: p. 110.

DENEYSEL TRAVMATİK BEYİN HASARI MODELLERİ

BÖLÜM 8

Güven AKÇAY[1]

Giriş

Travmatik beyin hasarı, dünya çapında önde gelen mortalite ve morbidite nedenidir. Travmatik beyin hasarı sonrası kişilerde sıklıkla motor, bilişsel ve sensoriyel fonksiyon kaybı görülür. Travmatik beyin hasarı, ölüm, sakatlık ve ruhsal bozukluklar gibi önemli sağlık sorunlarına yol açar. Travmatik beyin hasarı, tüm dünyada giderek artan sağlık sorunu olmaya devam etmektedir. Her yıl yaklaşık 1,7 milyon insanın kafa travması geçirdiği ve bu bireylerden yaklaşık 50.000 kişinin hayatını kaybettiği tahmin edilmektedir. Travmatik beyin hasarı her yaşta ve popülasyonda görülmesine rağmen, vakanın en yüksek olduğu yaş popülasyonunu çocuklar ve yaşlılar oluşturur. Düşme, sportif faaliyetler ve motorlu araç kazaları travmatik beyin hasarının en büyük risk faktörü olarak karşımıza çıkar. Travmatik beyin hasarına yönelik teşhis ve tedavi yöntemlerinin geliştirilmesi için nöropatolojisinin altında yatan moleküler ve hücresel mekanizmaların bilinmesi gerekir. Bundan dolayı farklı modellerde tanımlanmış hafif, orta ve şiddetli deneysel travmatik beyin hasarı modelleri kullanılır. Travmatik beyin hasarının hayvan modellerini genel olarak fokal, difüz ve karışık hasar olarak sınıflandırılır. Travmatik beyin hasarı deneysel araştırmalarında sıvı perküsyon, kontrollü kortikal etki, ağırlık düşürme ve patlama dalgası en sık tercih edilen modellerdir. Bu bölümde, travmatik beyin hasarı için mevcut kemirgen modellerinin güçlü ve zayıf yönleri açıklanacaktır.

[1] Dr. Öğr. Üyesi, Hitit Üniversitesi Tıp Fakültesi, Biyofizik AD., guvenakcayibu@gmail.com

için bir puan verilir: 13–18, şiddeti hasar; 7–12, orta orta hasar; 1-6, hafif hasar. Deneysel çalışmalarda nörolojik şiddet skoru ve motor fonksiyon testleri gibi davranış değişiklikleri değerlendirilmesinin yanında kilo kaybı ve kafa içi basınç artışı gibi fizyolojik değişiklikler; enfarktüs hacmi ve nöronal kayıp gibi histolojik değişiklikler de kullanılır (8, 79, 80).

Sonuç ve Öneriler

Travmatik beyin hasarı, ölüm ve sakatlığın önde gelen nedenlerinden biridir. TBH beyin dokusunun mekanik olarak bozulmasına neden olan dış gücün ve hasarı şiddetlendiren gecikmiş patojenik olayların sonucudur. Bu patojenik yaralanma süreçleri yeterince anlaşılmamıştır ve bu nedenle şimdiye kadar etkili nöroprotektif tedavi mevcut değildir. Deneysel modeller, TBH'nin fizyolojik ve patofizyolojik mekanizmalarının araştırması, yeni terapötik ajanları test edilmesi, klinik denemelerin güvenli ve başarılı olmasını sağlamak için hayvan modelleri gereklidir. İnsan TBH ile ilişkili farklı yaralanma mekanizmalarını modellemek için çeşitli kemirgen TBH modelleri geliştirilmiştir. Travmatik beyin hasarının kemirgen modelleri arasında en sık kullanılanları sıvı perküsyon, kortikal kontüzyon etki, ağırlık düşürme ve patlama dalgası modelleridir. Travmatik beyin hasarında meydana gelebilecek olayların tamamı tek bir kemirgen modeliyle kapsanamayacağından, belirli bir modelin tasarımı ve seçimi sinirbilimciler için büyük bir zorluk teşkil etmektedir. Bu bölümde, travmatik beyin hasarı için mevcut kemirgen modellerinin güçlü ve zayıf yönleri açıklanmıştır.

Kaynaklar

1. Masel, B.E. and D.S. DeWitt, *Traumatic brain injury: a disease process, not an event.* J Neurotrauma, 2010. 27(8): p. 1529-40.
2. Michael G, et al., *A Review of Traumatic Brain Injury Animal Models: Are We Lacking Adequate Models Replicating Chronic Traumatic Encephalopathy?* Journal of Neurology and Neurobiology, 2015. 2(1).
3. Langlois, J.A., W. Rutland-Brown, and M.M. Wald, *The epidemiology and impact of traumatic brain injury: a brief overview.* J Head Trauma Rehabil, 2006. 21(5): p. 375-8.
4. Maas, A.I., N. Stocchetti, and R. Bullock, *Moderate and severe traumatic brain injury in adults.* Lancet Neurol, 2008. 7(8): p. 728-41.
5. Ma, X., et al., *Animal Models of Traumatic Brain Injury and Assessment of Injury Severity.* Mol Neurobiol, 2019. 56(8): p. 5332-5345.
6. Briones, T.L., *Chapter 3 animal models of traumatic brain injury: is there an optimal model that parallels human brain injury?* Annu Rev Nurs Res, 2015. 33: p. 31-73.
7. Davis, A.E., *Mechanisms of traumatic brain injury: biomechanical, structural and cellular considerations.* Crit Care Nurs Q, 2000. 23(3): p. 1-13.
8. Cernak, I., *Animal models of head trauma.* NeuroRx, 2005. 2(3): p. 410-22.

9. Gaetz, M., *The neurophysiology of brain injury.* Clin Neurophysiol, 2004. 115(1): p. 4-18.
10. Hay, J., et al., *Chronic Traumatic Encephalopathy: The Neuropathological Legacy of Traumatic Brain Injury.* Annu Rev Pathol, 2016. 11: p. 21-45.
11. Xiong, Y., A. Mahmood, and M. Chopp, *Emerging treatments for traumatic brain injury.* Expert Opin Emerg Drugs, 2009. 14(1): p. 67-84.
12. Doppenberg, E.M., S.C. Choi, and R. Bullock, *Clinical trials in traumatic brain injury: lessons for the future.* J Neurosurg Anesthesiol, 2004. 16(1): p. 87-94.
13. Maas, A.I., *Neuroprotective agents in traumatic brain injury.* Expert Opin Investig Drugs, 2001. 10(4): p. 753-67.
14. Faden, A.I., *Neuroprotection and traumatic brain injury: theoretical option or realistic proposition.* Curr Opin Neurol, 2002. 15(6): p. 707-12.
15. Schouten, J.W., *Neuroprotection in traumatic brain injury: a complex struggle against the biology of nature.* Curr Opin Crit Care, 2007. 13(2): p. 134-42.
16. Irvine, K.-A. and J.D. Clark, *Chronic Pain After Traumatic Brain Injury: Pathophysiology and Pain Mechanisms.* Pain Medicine, 2018. 19: p. 1315–1333.
17. Povlishock, J.T. and C.W. Christman, *The pathobiology of traumatically induced axonal injury in animals and humans: a review of current thoughts.* J Neurotrauma, 1995. 12(4): p. 555-64.
18. Xiong, Y., A. Mahmood, and M. Chopp, *Animal models of traumatic brain injury.* Nat Rev Neurosci, 2013. 14(2): p. 128-42.
19. Raghupathi, R., *Cell death mechanisms following traumatic brain injury.* Brain Pathol, 2004. 14(2): p. 215-22.
20. Bramlett, H.M. and W.D. Dietrich, *Progressive damage after brain and spinal cord injury: pathomechanisms and treatment strategies.* Prog Brain Res, 2007. 161: p. 125-41.
21. Raghupathi, R., D.I. Graham, and T.K. McIntosh, *Apoptosis after traumatic brain injury.* J Neurotrauma, 2000. 17(10): p. 927-38.
22. Yakovlev, A.G., et al., *Differential expression of apoptotic protease-activating factor-1 and caspase-3 genes and susceptibility to apoptosis during brain development and after traumatic brain injury.* J Neurosci, 2001. 21(19): p. 7439-46.
23. Margulies, S. and R. Hicks, *Combination therapies for traumatic brain injury: prospective considerations.* J Neurotrauma, 2009. 26(6): p. 925-39.
24. Marklund, N., et al., *Evaluation of pharmacological treatment strategies in traumatic brain injury.* Curr Pharm Des, 2006. 12(13): p. 1645-80.
25. McIntosh, T.K., et al., *Traumatic brain injury in the rat: characterization of a lateral fluid-percussion model.* Neuroscience, 1989. 28(1): p. 233-44.
26. Chen J, Xu XM, and X. ZC, *Animal Models of Acute Neurological Injuries.* 2009, Berlin, Germany.
27. Gennarelli, T.A., *Animate models of human head injury.* J Neurotrauma, 1994. 11(4): p. 357-68.
28. Spain, A., et al., *Mild fluid percussion injury in mice produces evolving selective axonal pathology and cognitive deficits relevant to human brain injury.* J Neurotrauma, 2010. 27(8): p. 1429-38.
29. Su, Y., et al., *Taurine improves functional and histological outcomes and reduces inflammation in traumatic brain injury.* Neuroscience, 2014. 266: p. 56-65.
30. Kabadi, S.V., et al., *CR8, a novel inhibitor of CDK, limits microglial activation, astrocytosis, neuronal loss, and neurologic dysfunction after experimental traumatic brain injury.* J Cereb Blood Flow Metab, 2014. 34(3): p. 502-13.
31. Huang, E.Y., et al., *Amantadine ameliorates dopamine-releasing deficits and behavioral deficits in rats after fluid percussion injury.* PLoS One, 2014. 9(1): p. e86354.
32. Hameed, M.Q., et al., *A rapid lateral fluid percussion injury rodent model of traumatic brain injury and post-traumatic epilepsy.* Neuroreport, 2014. 25(7): p. 532-6.
33. Lifshitz, J., B.J. Kelley, and J.T. Povlishock, *Perisomatic thalamic axotomy after diffuse traumatic brain injury is associated with atrophy rather than cell death.* J Neuropathol Exp Neurol, 2007. 66(3): p. 218-29.

34. Cao, T., et al., *Morphological and genetic activation of microglia after diffuse traumatic brain injury in the rat.* Neuroscience, 2012. 225: p. 65-75.
35. Thompson, H.J., et al., *Lateral fluid percussion brain injury: a 15-year review and evaluation.* J Neurotrauma, 2005. 22(1): p. 42-75.
36. Hicks, R., et al., *Temporal and spatial characterization of neuronal injury following lateral fluid-percussion brain injury in the rat.* Acta Neuropathol, 1996. 91(3): p. 236-46.
37. Liu, Y.R., et al., *Progressive metabolic and structural cerebral perturbations after traumatic brain injury: an in vivo imaging study in the rat.* J Nucl Med, 2010. 51(11): p. 1788-95.
38. Hamm, R.J., *Neurobehavioral assessment of outcome following traumatic brain injury in rats: an evaluation of selected measures.* J Neurotrauma, 2001. 18(11): p. 1207-16.
39. Pierce, J.E., et al., *Enduring cognitive, neurobehavioral and histopathological changes persist for up to one year following severe experimental brain injury in rats.* Neuroscience, 1998. 87(2): p. 359-69.
40. Dixon, C.E., et al., *A controlled cortical impact model of traumatic brain injury in the rat.* J Neurosci Methods, 1991. 39(3): p. 253-62.
41. Chen, Y., et al., *A modified controlled cortical impact technique to model mild traumatic brain injury mechanics in mice.* Front Neurol, 2014. 5: p. 100.
42. Smith, S.L. and E.D. Hall, *Mild pre- and posttraumatic hypothermia attenuates blood-brain barrier damage following controlled cortical impact injury in the rat.* J Neurotrauma, 1996. 13(1): p. 1-9.
43. Osier, N.D. and C.E. Dixon, *The Controlled Cortical Impact Model: Applications, Considerations for Researchers, and Future Directions.* Front Neurol, 2016. 7: p. 134.
44. Washington, P.M., et al., *The effect of injury severity on behavior: a phenotypic study of cognitive and emotional deficits after mild, moderate, and severe controlled cortical impact injury in mice.* J Neurotrauma, 2012. 29(13): p. 2283-96.
45. Fox, G.B., et al., *Sustained sensory/motor and cognitive deficits with neuronal apoptosis following controlled cortical impact brain injury in the mouse.* J Neurotrauma, 1998. 15(8): p. 599-614.
46. Marklund, N. and L. Hillered, *Animal modelling of traumatic brain injury in preclinical drug development: where do we go from here?* Br J Pharmacol, 2011. 164(4): p. 1207-29.
47. Dixon, C.E., et al., *Amantadine improves water maze performance without affecting motor behavior following traumatic brain injury in rats.* Restor Neurol Neurosci, 1999. 14(4): p. 285-294.
48. Dixon, C.E., et al., *One-year study of spatial memory performance, brain morphology, and cholinergic markers after moderate controlled cortical impact in rats.* J Neurotrauma, 1999. 16(2): p. 109-22.
49. Kochanek, P.M., et al., *Cerebral blood flow at one year after controlled cortical impact in rats: assessment by magnetic resonance imaging.* J Neurotrauma, 2002. 19(9): p. 1029-37.
50. Mychasiuk, R., A. Farran, and M.J. Esser, *Assessment of an experimental rodent model of pediatric mild traumatic brain injury.* J Neurotrauma, 2014. 31(8): p. 749-57.
51. Morales, D.M., et al., *Experimental models of traumatic brain injury: do we really need to build a better mousetrap?* Neuroscience, 2005. 136(4): p. 971-89.
52. Viano, D.C., et al., *Evaluation of three animal models for concussion and serious brain injury.* Ann Biomed Eng, 2012. 40(1): p. 213-26.
53. Marmarou, A., et al., *A new model of diffuse brain injury in rats. Part I: Pathophysiology and biomechanics.* J Neurosurg, 1994. 80(2): p. 291-300.
54. Flierl, M.A., et al., *Mouse closed head injury model induced by a weight-drop device.* Nat Protoc, 2009. 4(9): p. 1328-37.
55. Adelson, P.D., et al., *A model of diffuse traumatic brain injury in the immature rat.* J Neurosurg, 1996. 85(5): p. 877-84.
56. Statler, K.D., et al., *Isoflurane exerts neuroprotective actions at or near the time of severe traumatic brain injury.* Brain Res, 2006. 1076(1): p. 216-24.

57. Feeney, D.M., et al., *Responses to cortical injury: I. Methodology and local effects of contusions in the rat.* Brain Res, 1981. 211(1): p. 67-77.
58. Albert-Weissenberger, C. and A.L. Sirén, *Experimental traumatic brain injury.* Exp Transl Stroke Med, 2010. 2(1): p. 16.
59. Nilsson, P., et al., *Continuous measurement of changes in regional cerebral blood flow following cortical compression contusion trauma in the rat.* J Neurotrauma, 1996. 13(4): p. 201-7.
60. Lindh, C., et al., *Differences in cell death between high and low energy brain injury in adult rats.* Acta Neurochir (Wien), 2008. 150(12): p. 1269-75;discussion 1275.
61. Shapira, Y., et al., *Experimental closed head injury in rats: mechanical, pathophysiologic, and neurologic properties.* Crit Care Med, 1988. 16(3): p. 258-65.
62. Chen, Y., et al., *An experimental model of closed head injury in mice: pathophysiology, histopathology, and cognitive deficits.* J Neurotrauma, 1996. 13(10): p. 557-68.
63. Zhu, F., et al., *Biomechanical responses of a pig head under blast loading: a computational simulation.* Int J Numer Method Biomed Eng, 2013. 29(3): p. 392-407.
64. Reneer, D.V., et al., *A multi-mode shock tube for investigation of blast-induced traumatic brain injury.* J Neurotrauma, 2011. 28(1): p. 95-104.
65. Koliatsos, V.E., et al., *A mouse model of blast injury to brain: initial pathological, neuropathological, and behavioral characterization.* J Neuropathol Exp Neurol, 2011. 70(5): p. 399-416.
66. Yi, J., et al., *Chronic traumatic encephalopathy.* Curr Sports Med Rep, 2013. 12(1): p. 28-32.
67. Geddes, J.F., et al., *Neuronal cytoskeletal changes are an early consequence of repetitive head injury.* Acta Neuropathol, 1999. 98(2): p. 171-8.
68. Weber, J.T., *Experimental models of repetitive brain injuries.* Prog Brain Res, 2007. 161: p. 253-61.
69. Uryu, K., et al., *Repetitive mild brain trauma accelerates Abeta deposition, lipid peroxidation, and cognitive impairment in a transgenic mouse model of Alzheimer amyloidosis.* J Neurosci, 2002. 22(2): p. 446-54.
70. DeFord, S.M., et al., *Repeated mild brain injuries result in cognitive impairment in B6C3F1 mice.* J Neurotrauma, 2002. 19(4): p. 427-38.
71. Huang, L., et al., *Tissue vulnerability is increased following repetitive mild traumatic brain injury in the rat.* Brain Res, 2013. 1499: p. 109-20.
72. Luo, J., et al., *Long-term cognitive impairments and pathological alterations in a mouse model of repetitive mild traumatic brain injury.* Front Neurol, 2014. 5: p. 12.
73. Rákos, G., et al., *Evans Blue fluorescence permits the rapid visualization of non-intact cells in the perilesional rim of cold-injured rat brain.* Acta Neurobiol Exp (Wars), 2007. 67(2): p. 149-54.
74. Raslan, F., et al., *Inhibition of bradykinin receptor B1 protects mice from focal brain injury by reducing blood-brain barrier leakage and inflammation.* J Cereb Blood Flow Metab, 2010. 30(8): p. 1477-86.
75. Brotfain, E., et al., *Neuroprotection by Estrogen and Progesterone in Traumatic Brain Injury and Spinal Cord Injury.* Curr Neuropharmacol, 2016. 14(6): p. 641-53.
76. Prins, M.L. and D.A. Hovda, *Developing experimental models to address traumatic brain injury in children.* J Neurotrauma, 2003. 20(2): p. 123-37.
77. Potts, M.B., et al., *Traumatic injury to the immature brain: inflammation, oxidative injury, and iron-mediated damage as potential therapeutic targets.* NeuroRx, 2006. 3(2): p. 143-53.
78. Rabinowitz, A.R. and H.S. Levin, *Cognitive sequelae of traumatic brain injury.* Psychiatr Clin North Am, 2014. 37(1): p. 1-11.
79. Osier, N.D., et al., *Chronic Histopathological and Behavioral Outcomes of Experimental Traumatic Brain Injury in Adult Male Animals.* J Neurotrauma, 2015. 32(23): p. 1861-82.
80. Johnson, V.E., et al., *Animal models of traumatic brain injury.* Handb Clin Neurol, 2015. 127: p. 115-28.

IN VİVO VOLTAMETRİ İLE NÖROTRANSMİTTER ÖLÇÜMLERİ

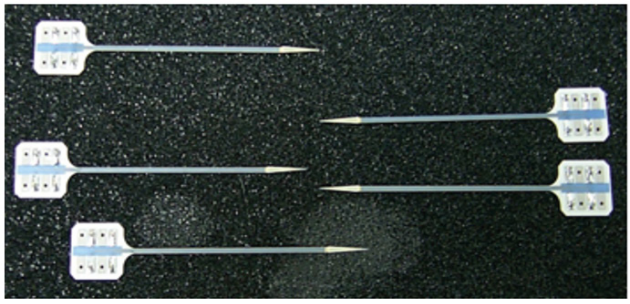
BÖLÜM 9

Ahmet HACIMÜFTÜOĞLU[1]
Fatma YEŞİLYURT[2]

Giriş

Merkezi Sinir Sistemi (MSS)'nde bulunan sinirler arasında iletiyi ve iletişimi sağlayan nörotransmitterlerin düzey değişimlerinin Parkinson, epilepsi, şizofreni, ilaç bağımlılığı ve birçok beyin hastalığında rollerinin olduğu birçok çalışmayla gösterilmiştir. Son 20 yıldır çok yaygın şekilde kullanılan mikrodiyaliz metodlar, nörotransmitterlerin dakika başı ölçümlerini vermiştir. Fakat nörotransmitterlerin hızlı dinamikleri, saniye temelinde ölçüm yapan bir tekniğe ihtiyaç doğurmuştur. In vivo voltametri sayesinde nörotransmitterlerin yaşayan canlıda miktar ölçümleri yüksek özgünlükte ve seçicilikte artık ortaya konabilmektedir. Bu teknikte elektrik akımını algılama ve iletme özelliğine sahip beyin elektrotları kullanılmaktadır. İstenen nörotransmitterlerin ölçülebilmesi için seçici bariyerlerle ve özel enzimlerle elektrotlar kaplanmaktadır. Bu teknikte uygulanan voltaj, ortamdaki nörotransmitterin enzimle reaksiyona girerek akım oluşturabilen maddelere dönüşümünü sağlamaktadır. Sonuçta sistemde algılanan elektrik akım miktarı ortamdaki nörotransmitter miktarıyla birebir ilişkili bulunmaktadır. Ayrıca beyine sokulan elektrot uçlarının hasar vermemesi yine mikrodiyaliz problarının ölçümlerine göre in vivo voltametri tekniğine önemli bir üstünlük sağlamaktadır. Şu an mevcut sistemler içinde anestezi altında veya serbest hareket edebilen hayvanlarda o bölgede en net nörotransmitter ölçümünün in vivo voltammetri tekniği ile yapılabileceği iddia edilebilir.

[1] Prof. Dr., Atatürk Üniversitesi Tıp Fakültesi, Tıbbi Farmakoloji AD, hacimuftuoglu@gmail.com
[2] Arş Gör., Atatürk Üniversitesi Tıp Fakültesi, Tıbbi Farmakoloji AD, fatmayesilyurttt@gmail.com



Şekil 14. Etik izinleri alınmış, deney hayvanlarında yapılan deneysel bir çalışma

7.2.İMPLANTASYONUN HİSTOPATOLOJİSİ

Kronik olarak implante edilen L-glutamat elektrodların çevre beyin dokusunda ya çok az ya da hiç beyin hasarına yol açmadığı gösterilmiştir. Sıçanlara 2, 4 veya 8 hafta süresince kronik olarak mikroelektrotlar implante edilmiş ve bu sürelerin sonunda sıçanların beyinleri çıkarılmış, kesitlere ayrılmış ve hem astrositler hem de mikroglialar boyanmıştır. Boyama sonucunda her üç zaman periyodu içinde implant çevresinde kontrol ile kıyaslanınca astrositlerin sayısında ve mikroglia aktivasyonunda kontrole göre istatistiksel bir fark görülmemiştir.

Kaynaklar

1. Binnetoğlu, D., Aricioglu, F., Ozcan, H. Antidepressant-Like Effects of Ceftriaxone in Chronic Unpredictable Mild Stress Model in Rats: The Importance of Reuptake Time of Glutamate. *Clinical and Experimental Health Sciences*, 2019, 9(4), 294-299.
2. Taspinar, N., Hacimuftuoglu, A., Butuner, S. Differential effects of inhibitors of PTZ- induced kindling on glutamate transporters and enzyme expression. *Clinical and Experimental Pharmacology and Physiology*, 2021, 48(12), 1662-1673.

3. Faingold, C. L. Emergent properties of CNS neuronal networks as targets for pharmacology: application to anticonvulsant drug action. *Progress in Neurobiology*, 2004, 72(1), 55-85.
4. D'Antoni, S., Berretta, A., Bonaccorso, C. M. Metabotropic glutamate receptors in glial cells. *Neurochemical Research*, 2008, 33(12), 2436-2443.
5. Engel Jr, J., Thompson, P. M., Stern, J. M. Connectomics and epilepsy. *Current Opinion in Neurology*, 2013, 26(2), 186.
6. Masliah, E., Hansen, L., Alford, M. Deficient glutamate tranport is associated with neurodegeneration in Alzheimer's disease. Annals of Neurology: *Official Journal of the American Neurological Association and the Child Neurology Society*, 1996, 40(5), 759-766.
7. Hacimuftuoglu, A., Tatar, A., Cetin, D.(). Astrocyte/neuron ratio and its importance on glutamate toxicity: an in vitro voltammetric study. *Cytotechnology*, 2016, 68(4), 1425-1433.
8. Zhou, Y., & Danbolt, N. C. Glutamate as a neurotransmitter in the healthy brain. *Journal of Neural Transmission*, 2014, 121(8), 799-817.
9. Burmeister, J. J., Moxon, K., & Gerhardt, G. A. Ceramic-based multisite microelectrodes for electrochemical recordings. *Analytical Chemistry*, 2000, 72(1), 187-192.
10. Yeni, Y., Cakir, Z., Hacimuftuoglu, A. A Selective Histamine H4 Receptor Antagonist, JNJ7777120, Role on glutamate Transporter Activity in Chronic Depression. *Journal of Personalized Medicine*, 2022, 12(2), 246.
11. Ku, S., Palanisamy, S., & Chen, S. M. Highly selective dopamine electrochemical sensor based on electrochemically pretreated graphite and nafion composite modified screen printed carbon electrode. *Journal of Colloid and İnterface Science*, 2013, 411, 182-186.
12. Zhao, J., Yu, Y., Weng, B. Sensitive and selective dopamine determination in human serum with inkjet printed Nafion/MWCNT chips. *Electrochemistry communications*, 2013, 37, 32-35.
13. Noroozifar, M., Khorasani-Motlagh, M., Nadiki, H. Modified fluorine-doped tin oxide electrode with inorganic ruthenium red dye-multiwalled carbon nanotubes for simultaneous determination of a dopamine, uric acid, and tryptophan. *Sensors and Actuators B: Chemical*, 2014, 204, 333-341.
14. Wang, L., Liu, Q., & Ye, B. An electroanalytical method for glabridin investigation based on poly (diallyldimethylammonium chloride)-functionalized graphene-modified electrode. *Microchemical Journal*, 2020, 158, 105279.
15. Hascup, K. N., Rutherford, E. C., Quintero, J. E. Second-by-second measures of L-glutamate and other neurotransmitters using enzyme-based microelectrode arrays. *Electrochemical Methods for Neuroscience*. 2007
16. Bortz, D. M., Mikkelsen, J. D., & Bruno, J. P. Localized infusions of the partial alpha 7 nicotinic receptor agonist SSR180711 evoke rapid and transient increases in prefrontal glutamate release. *Neuroscience*, 2013, *255*, 55-67.
17. Adams, R. N. In vivo electrochemical measurements in the CNS. *Progress in Neurobiology*, 1990, 35(4), 297-311.