

13.

BÖLÜM

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GİRİŞ

Kanserler, hücrelerin normal doku gelişiminin haricinde meydana gelen proliferasyonlarla sonuçlanan mutasyonlar kazandığı hastalıklardır. Bu somatik mutasyonlar, çevrelesel etkileşimlerle hücre dönüşümüne sebep olurlar. Bu çevre; genetik, yaşam tarzi, diyet, fizyoloji,immün sistemin durumu ve tümör hücrelerinin mikro ortamı gibi çok sayıda faktör tarafından belirlenir ancak bunlarla sınırlı değildir. Genler ve çevre arasındaki bu etkileşimin bağlantı noktasında da **hücresel metabolizma** bulunur.

Metabolizma, enzimlerin aracılık ettiği birbirine bağlı bir dizi kimyasal reaksiyonu içerir. Diyet ile makro besinler olarak alınan karbonhidratlar, yağlar ve proteinler, hücresel düzeyde glukoz, lipitler ve amino asitler olarak alınır ve daha sonra metabolik ağı oluşturan bir dizi birbirine bağlı kimyasal reaksiyonla dönüştürülürler. Bu besinler elektron açısından zengin indirgenmiş formlarındada saklanır ve katabolizma olarak bilinen süreç boyunca oksitlenir. Bu oksidatif süreçlerde elektronların serbest bırakılması ile, yaşamın devamını sürdürmek için gereken enerji üretilir. Metabolizma ayrıca katabolizmanın ara maddelerinin yapısal bileşenlere ya da protein, nükleik asit, hücre membranı yapımında kullanılan malzemelere dönüştürülmesine de izin verir. Buna **anabolizma** veya **anabolik metabolizma** denir. Metabolizma, proteinler ve nükleik asitlerle doğrudan reaksiyona giren ve onların aktivitesini değiştiren moleküller erek hücresel mekanizmanın diğer unsurlarıyla doğrudan iletişim kurabilir.

Kontrolsüz proliferasyonu sürdürmeli için, bütün bu metabolik fonksiyonlara adapte olabilme yeteneği malignitenin her bir aşamasının yaygın özelliğidir. Gerçekten de, farklı metabolik gereksinimleranoikiz, invazyon ve metastaz gibi kanserle ilişkili her süreçte kendini gösterir. Bu metabolik programlama, hem farklı metabolik süreçleri yönlendiren onkojenik sinyallerden hem de doku vaskülaritesi ve serum besin içeriği gibi çevresel faktörlerden ileri gelir. Metabolizma, diyet ve egzersiz gibi yaşam tarzi tarafından da manipüle edilebilir. Bu nedenle kanserin önlenmesi ve tedavisinde metabolizmadan yararlanmak çok önemli bir yoldur.

Bu bölümde, metabolik yeniden programmanın temelleri ve kanser hücrelerinde adaptasyon tartışılmaktadır. Hem bu değişmiş metabolizmaya yol açan ilkeler hem de sonuca ortaya çıkan özel gereksinimler ana hatlarıyla belirtilmektedir.

WARBURG ETKİSİ

Yeterli oksijen bulunan koşullarda, terminal olarak farklılaşmış hücreler genellikle glikozu üç enzimatik yol aracılığıyla metabolize eder; **glikoliz**, **trikarboksilik asit (TCA) döngüsü** ve **elektron taşıma zinciri (ETC)** [1,2].

Tam oksidasyonda; bir glikoz molekülünden sonuca 36 enerji depolayan molekül adenozin 5'-trifosfat (ATP) ve indirgenmiş nikotinamid adenin dinükleotid (NADH) [1,2] gibi enerji açısından zengin bileşikler elde edilir. Bu süreçler bölgelere ayrılmıştır: sitozolde glikoliz, mitokondride TCA döngüsü ve ETC.

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miktarda glikoza ihtiyaç duyar ve birçok kanser hücresiyle aynı derecede Warburg etkisine maruz kalır[12]. Yetersiz glikoz, T hücresi inaktivasyonuna veya ölümüne yol açabilir[129,130]. Gerçekten de, tümör mikro ortamında bulunan immuno-supresyonun çoğu, T hücrelerinin yeterli besinleri elde edememesine atfedilir[131]. Ek olarak, tümör hücreleri, Efektör T hücrelerinde hücre ölümüne neden olan triptofan katabolizmasını aktive etmek için indolamin 2'3'-dioksijenazı (IDO) upregüle eder[132]. Besin stresine ek olarak, oksijen miktarı da T hücre fonksiyonunu etkilemede önemli bir rol oynar. Hipoksinin neden olduğu HIF aktivasyonu, immün aktiviteyi baskılanan PD-1 ve CTLA4'ü transkripsiyonel olarak upregüle eder[133,134]. HIF ayrıca, immun aktiviteyi azaltan T regulatuar hücrelerin (Tregs) ekspresyonunu destekler[135]. Bu nedenle, T hücreleri tümör baskılıyıcı veya tümörü teşvik edici olabilir[136].

T hücrelerinin ex vivo ve in vivo uyarılması- na odaklanan immünoterapi tümörlerin yönetimi için umut verici bir yoldur.

SONUÇ

Kanser metabolizmasına olan ilgi artışı bu hücrelerdeki metabolik yolaklarının hücresel otonom değişikliklerine yönelik olsa da, şimdilerde bu tümör hücrelerini etkileyen mikroçevreleri, dokusal orjinleri, germ hücresi genetikleri, karaciğer fizyolojisi, mikrobiyom, diyet ve egzersiz dahil birçok unsuru içeren çevreleriyle olan etkileşimlerinin önemi çok daha iyi anlaşılmaktadır. Bu faktörlerdeki çeşitliliğin tümörlerdeki metabolik yolları nasıl etkileyebileceği çalışmaların ana konusu olacak gibi görünmektedir.

KAYNAKLAR

- Berg JM, Tymoczko JL, Stryer L, Stryer L. *Biochemistry*. 6th ed. New York: WH Freeman; 2007.
- Lehninger AL, Nelson DL, Cox MM. *Lehninger Principles of Biochemistry*. 6th ed. New York: W.H. Freeman; 2013.
- Rolland F, Winderickx J, Thevelein JM. Glucose-sensing and -signalling mechanisms in yeast. *FEMS Yeast Res*. 2002;2(2):183–201.
- Sherman IW. *Malaria: Parasite Biology, Pathogenesis, and Protection*. Washington, DC: ASM Press; 1998.
- Pfeiffer T, Schuster S, Bonhoeffer S. Cooperation and competition in the evolution of ATP-producing pathways. *Science*. 2001;292(5516):504–507.

- Shestov AA, Liu X, Ser Z, et al. Quantitative determinants of aerobic glycolysis identify flux through the enzyme GAPDH as a limiting step. *Elife*. 2014;3.
- Warburg O, Posener K, Negelein E. On the metabolism of carcinoma cells. *Biochem Z*. 1924;152:309–344.
- Warburg O, Wind F, Negelein E. The metabolism of tumors in the body. *J Gen Physiol*. 1927;8(6):519–530.
- Fantin VR, St-Pierre J, Leder P. Attenuation of LDH-A expression uncovers a link between glycolysis, mitochondrial physiology, and tumor maintenance. *Cancer Cell*. 2006;9(6):425–434.
- Shim H, Chun YS, Lewis BC, Dang CV. A unique glucose-dependent apoptotic pathway induced by c-Myc. *Proc Natl Acad Sci USA*. 1998;95(4):1511–1516.
- Le A, Stine ZE, Nguyen C, et al. Tumorigenicity of hypoxic respiring cancer cells revealed by a hypoxia-cell cycle dual reporter. *Proc Natl Acad Sci USA*. 2014;111(34):12486–12491.
- Gerriets VA, Kishton RJ, Nichols AG, et al. Metabolic programming and PDHK1 control CD4+ T cell subsets and inflammation. *J Clin Invest*. 2015;125(1):194–207.
- Vincent AS, Phan TT, Mukhopadhyay A, Lim HY, Halliwell B, Wong KP. Human skin keloid fibroblasts display bioenergetics of cancer cells. *J Invest Dermatol*. 2008;128(3):702–709.
- Ben-Haim S, Ell P. 18F-FDG PET and PET/CT in the evaluation of cancer treatment response. *J Nucl Med*. 2009;50(1):88–99.
- Liberti MV, Locasale JW. The Warburg effect: how does it benefit cancer cells? *Trends Biochem Sci*. 2016;41(3):211–218.
- Warburg O. On the origin of cancer cells. *Science*. 1956;123(3191):309–314.
- Slavov N, Budnik BA, Schwab D, Airolidi EM, van Oudenaarden A. Constant growth rate can be supported by decreasing energy flux and increasing aerobic glycolysis. *Cell Rep*. 2014;7(3):705–714.
- Locasale JW, Cantley LC. Metabolic flux and the regulation of mammalian cell growth. *Cell Metab*. 2011;14(4):443–451.
- Locasale JW, Grassian AR, Melman T, et al. Phosphoglycerate dehydrogenase diverts glycolytic flux and contributes to oncogenesis. *Nat Genet*. 2011;43(9):869–874.
- Lunt SY, Vander Heiden MG. Aerobic glycolysis: meeting the metabolic requirements of cell proliferation. *Annu Rev Cell Dev Biol*. 2011;27:441–464.
- Gatenby RA, Gawlinski ET. A reaction-diffusion model of cancer invasion. *Cancer Res*. 1996;56(24):5745–5753.
- Estrella V, Chen T, Lloyd M, et al. Acidity generated by the tumor microenvironment drives local invasion. *Cancer Res*. 2013;73(5):1524–1535.
- Boroughs LK, DeBerardinis RJ. Metabolic pathways promoting cancer cell survival and growth. *Nat Cell Biol*. 2015;17(4):351–359.
- Wise DR, DeBerardinis RJ, Mancuso A, et al. Myc regulates a transcriptional program that stimulates mitochondrial glutaminolysis and leads to glutamine addiction. *Proc Natl Acad Sci USA*. 2008;105(48):18782–18787.
- Gao P, Tchernyshyov I, Chang TC, et al. c-Myc suppression of miR-23a/b enhances mitochondrial glutaminase expression and glutamine metabolism. *Nature*. 2009;458(7239):762–765.

26. Son J, Lyssiotis CA, Ying H, et al. Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature*. 2013;496(7443):101–105.
27. Commissio C, Davidson SM, Soydaner-Azeloglu RG, et al. Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature*. 2013;497(7451):633–637.
28. Degenhardt K, Mathew R, Beaudoin B, et al. Autophagy promotes tumor cell survival and restricts necrosis, inflammation, and tumorigenesis. *Cancer Cell*. 2006;10(1):51–64.
29. Yang S, Wang X, Contino G, et al. Pancreatic cancers require autophagy for tumor growth. *Genes Dev*. 2011;25(7):717–729.
30. Altman BJ, Stine ZE, Dang CV. From Krebs to clinic: glutamine metabolism to cancer therapy. *Nat Rev Cancer*. 2016;16(10):619–634.
31. Bergstrom J, Furst P, Noree LO, Vinnars E. Intracellular free amino acid concentration in human muscle tissue. *J Appl Physiol*. 1974;36(6):693–697.
32. Welbourne TC. Ammonia production and glutamine incorporation into glutathione in the functioning rat kidney. *Can J Biochem*. 1979;57(3):233–237.
33. Jiang L, Shestov AA, Swain P, et al. Reductive carboxylation supports redox homeostasis during anchorage-independent growth. *Nature*. 2016;532(7598):255–258.
34. DeBerardinis RJ, Mancuso A, Daikhin E, et al. Beyond aerobic glycolysis: transformed cells can engage in glutamine metabolism that exceeds the requirement for protein and nucleotide synthesis. *Proc Natl Acad Sci USA*. 2007;104(49):19345–19350.
35. DeBerardinis RJ, Lum JJ, Hatzivassiliou G, Thompson CB. The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. *Cell Metab*. 2008;7(1):11–20.
36. Davidson SM, Papagiannakopoulos T, Olenchock BA, et al. Environment impacts the metabolic dependencies of ras-driven non-small cell lung cancer. *Cell Metab*. 2016;23(3):517–528.
37. Gross MI, Demo SD, Dennison JB, et al. Antitumor activity of the glutaminase inhibitor CB-839 in triple-negative breast cancer. *Mol Cancer Ther*. 2014;13(4):890–901.
38. Xiang Y, Stine ZE, Xia J, et al. Targeted inhibition of tumor-specific glutaminase diminishes cell-autonomous tumorigenesis. *J Clin Invest*. 2015;125(6):2293–2306.
39. Kung HN, Marks JR, Chi JT. Glutamine synthetase is a genetic determinant of cell type-specific glutamine independence in breast epithelia. *PLoS Genet*. 2011;7(8):e1002229.
40. Herranz D, Ambesi-Impiombato A, Suderth J, et al. Metabolic reprogramming induces resistance to anti-NOTCH1 therapies in T cell acute lymphoblastic leukemia. *Nat Med*. 2015;21(10):1182–1189.
41. MacFarlane AJ, Liu X, Perry CA, et al. Cytoplasmic serine hydroxymethyltransferase regulates the metabolic partitioning of methylenetetrahydrofolate but is not essential in mice. *J Biol Chem*. 2008;283(38):25846–25853.
42. Kalhan SC, Uppal SO, Moorman JL, et al. Metabolic and genomic response to dietary isocaloric protein restriction in the rat. *J Biol Chem*. 2011;286(7):5266–5277.
43. Yang M, Vousden KH. Serine and one-carbon metabolism in cancer. *Nat Rev Cancer*. 2016;16(10):650–662.
44. Locasale JW. Serine, glycine and one-carbon units: cancer metabolism in full circle. *Nat Rev Cancer*. 2013;13(8):572–583.
45. Mullarky E, Lucki NC, Beheshti Zavareh R, et al. Identification of a small molecule inhibitor of 3-phosphoglyceraldehyde dehydrogenase to target serine biosynthesis in cancers. *Proc Natl Acad Sci USA*. 2016;113(7):1778–1783.
46. Pacold ME, Brimacombe KR, Chan SH, et al. A PHGDH inhibitor reveals coordination of serine synthesis and one-carbon unit fate. *Nat Chem Biol*. 2016;12(6):452–458.
47. Wang Q, Liberti MV, Liu P, et al. Rational design of selective allosteric inhibitors of PHGDH and serine synthesis with anti-tumor activity. *Cell Chem Biol*. 2017;24(1):55–65.
48. Menth SJ, Mehrmohamadi M, Huang L, et al. Histone methylation dynamics and gene regulation occur through the sensing of one-carbon metabolism. *Cell Metab*. 2015;22(5):861–873.
49. Davis SR, Stacpoole PW, Williamson J, et al. Tracer-derived total and folate-dependent homocysteine remethylation and synthesis rates in humans indicate that serine is the main one-carbon donor. *Am J Physiol Endocrinol Metab*. 2004;286(2):E272–E279.
50. Dillon BJ, Prieto VG, Curley SA, et al. Incidence and distribution of argininosuccinate synthetase deficiency in human cancers: a method for identifying cancers sensitive to arginine deprivation. *Cancer*. 2004;100(4):826–833.
51. Grohmann U, Bronte V. Control of immune response by amino acid metabolism. *Immunol Rev*. 2010;236:243–264.
52. Mellor AL, Munn DH. IDO expression by dendritic cells: tolerance and tryptophan catabolism. *Nat Rev Immunol*. 2004;4(10):762–774.
53. Mayers JR, Wu C, Clish CB, et al. Elevation of circulating branched-chain amino acids is an early event in human pancreatic adenocarcinoma development. *Nat Med*. 2014;20(10):1193–1198.
54. Laplante M, Sabatini DM. mTOR signaling in growth control and disease. *Cell*. 2012;149(2):274–293.
55. Wallace DC. Mitochondria and cancer. *Nat Rev Cancer*. 2012;12(10):685–698.
56. Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants, and the degenerative diseases of aging. *Proc Natl Acad Sci USA*. 1993;90(17):7915–7922.
57. Weinberg F, Hamanaka R, Wheaton WW, et al. Mitochondrial metabolism and ROS generation are essential for Kras-mediated tumorigenicity. *Proc Natl Acad Sci USA*. 2010;107(19):8788–8793.
58. Weinhouse S, Millington RH, Wenner CE. Metabolism of neoplastic tissue. I. The oxidation of carbohydrate and fatty acids in transplanted tumors. *Cancer Res*. 1951;11(11):845–850.
59. Wenner CE, Spirtes MA, Weinhouse S. Metabolism of neoplastic tissue. II. A survey of enzymes of the citric acid cycle in transplanted tumors. *Cancer Res*. 1952;12(1):44–49.

60. Fogal V, Richardson AD, Karmali PP, Scheffler IE, Smith JW, Ruoslahti E. Mitochondrial p32 protein is a critical regulator of tumor metabolism via maintenance of oxidative phosphorylation. *Mol Cell Biol*. 2010;30(6):1303–1318.
61. Weinberg SE, Chandel NS. Targeting mitochondria metabolism for cancer therapy. *Nat Chem Biol*. 2015;11(1):9–15.
62. Vazquez F, Lim JH, Chim H, et al. PGC1alpha expression defines a subset of human melanoma tumors with increased mitochondrial capacity and resistance to oxidative stress. *Cancer Cell*. 2013;23(3):287–301.
63. Haq R, Shoag J, Andreu-Perez P, et al. Oncogenic BRAF regulates oxidative metabolism via PGC1alpha and MITF. *Cancer Cell*. 2013;23(3):302–315.
64. Ookhtens M, Kannan R, Lyon I, Baker N. Liver and adipose tissue contributions to newly formed fatty acids in an ascites tumor. *Am J Physiol*. 1984;247(1 Pt 2):R146–R153.
65. Medes G, Thomas A, Weinhouse S. Metabolism of neoplastic tissue. IV. A study of lipid synthesis in neoplastic tissue slices in vitro. *Cancer Res*. 1953;13(1):27–29.
66. Wakil SJ, Stoops JK, Joshi VC. Fatty acid synthesis and its regulation. *Annu Rev Biochem*. 1983;52:537–579.
67. Currie E, Schulze A, Zechner R, Walther TC, Farese RV Jr. Cellular fatty acid metabolism and cancer. *Cell Metab*. 2013;18(2):153–161.
68. Bauer DE, Hatzivassiliou G, Zhao F, Andreadis C, Thompson CB. ATP citrate lyase is an important component of cell growth and transformation. *Oncogene*. 2005;24(41):6314–6322.
69. Hatzivassiliou G, Zhao F, Bauer DE, et al. ATP citrate lyase inhibition can suppress tumor cell growth. *Cancer Cell*. 2005;8(4):311–321.
70. Migita T, Narita T, Nomura K, et al. ATP citrate lyase: activation and therapeutic implications in non-small cell lung cancer. *Cancer Res*. 2008;68(20):8547–8554.
71. Shaw RJ, Kosmatka M, Bardeesy N, et al. The tumor suppressor LKB1 kinase directly activates AMP-activated kinase and regulates apoptosis in response to energy stress. *Proc Natl Acad Sci USA*. 2004;101(10):3329–3335.
72. Chajes V, Cambot M, Moreau K, Lenoir GM, Joulin V. Acetyl-CoA carboxylase alpha is essential to breast cancer cell survival. *Cancer Res*. 2006;66(10):5287–5294.
73. Beckers A, Organe S, Timmermans L, et al. Chemical inhibition of acetyl-CoA carboxylase induces growth arrest and cytotoxicity selectively in cancer cells. *Cancer Res*. 2007;67(17):8180–8187.
74. Jeon SM, Chandel NS, Hay N. AMPK regulates NADPH homeostasis to promote tumour cell survival during energy stress. *Nature*. 2012;485(7400):661–665.
75. Pizer ES, Thupari J, Han WF, et al. Malonyl-coenzyme-A is a potential mediator of cytotoxicity induced by fatty-acid synthase inhibition in human breast cancer cells and xenografts. *Cancer Res*. 2000;60(2):213–218.
76. Menendez JA, Lupu R. Fatty acid synthase and the lipogenic phenotype in cancer pathogenesis. *Nat Rev Cancer*. 2007;7(10):763–777.
77. Carracedo A, Cantley LC, Pandolfi PP. Cancer metabolism: fatty acid oxidation in the limelight. *Nat Rev Cancer*. 2013;13(4):227–232.
78. Zaugg K, Yao Y, Reilly PT, et al. Carnitine palmitoyltransferase 1C promotes cell survival and tumor growth under conditions of metabolic stress. *Genes Dev*. 2011;25(10):1041–1051.
79. Schafer ZT, Grassian AR, Song L, et al. Antioxidant and oncogene rescue of metabolic defects caused by loss of matrix attachment. *Nature*. 2009;461(7260):109–113.
80. Camarda R, Zhou AY, Kohnz RA, et al. Inhibition of fatty acid oxidation as a therapy for MYC-overexpressing triple-negative breast cancer. *Nat Med*. 2016;22(4):427–432.
81. Loftus TM, Jaworsky DE, Frehywot GL, et al. Reduced food intake and body weight in mice treated with fatty acid synthase inhibitors. *Science*. 2000;288(5475):2379–2381.
82. Rohrig F, Schulze A. The multifaceted roles of fatty acid synthesis in cancer. *Nat Rev Cancer*. 2016;16(11):732–749.
83. Graeser R, Bornmann C, Esser N, et al. Antimetastatic effects of liposomal gemcitabine and empty liposomes in an orthotopic mouse model of pancreatic cancer. *Pancreas*. 2009;38(3):330–337.
84. Morgan HD, Santos F, Green K, Dean W, Reik W. Epigenetic reprogramming in mammals. *Hum Mol Genet*. 2005;14(Spec1):R47–R58.
85. Dawson MA, Kouzarides T. Cancer epigenetics: from mechanism to therapy. *Cell*. 2012;150(1):12–27.
86. Sharma S, Kelly TK, Jones PA. Epigenetics in cancer. *Carcinogenesis*. 2010;31(1):27–36.
87. Suva ML, Riggi N, Bernstein BE. Epigenetic reprogramming in cancer. *Science*. 2013;339(6127):1567–1570.
88. Ptashne M. How eukaryotic transcriptional activators work. *Nature*. 1988;335(6192):683–689.
89. Ullrich A, Schlessinger J. Signal transduction by receptors with tyrosine kinase activity. *Cell*. 1990;61(2):203–212.
90. Yun J, Johnson JL, Hanigan CL, Locasale JW. Interactions between epigenetics and metabolism in cancers. *Front Oncol*. 2012;2:163.
91. Gao X, Reid MA, Kong M, Locasale JW. Metabolic interactions with cancer epigenetics. *Mol Aspects Med*. 2016.
92. Kinnaird A, Zhao S, Wellen KE, Michelakis ED. Metabolic control of epigenetics in cancer. *Nat Rev Cancer*. 2016;16(11):694–707.
93. Mentch SJ, Locasale JW. One-carbon metabolism and epigenetics: understanding the specificity. *Ann NY Acad Sci*. 2016;1363:91–98.
94. Shyh-Chang N, Locasale JW, Lyssiotis CA, et al. Influence of threonine metabolism on S-adenosylmethionine and histone methylation. *Science*. 2013;339(6116):222–226.
95. Nishikawa K, Iwamoto Y, Kobayashi Y, et al. DNA methyltransferase 3a regulates osteoclast differentiation by coupling to an S-adenosylmethionine-producing metabolic pathway. *Nat Med*. 2015;21(3):281–287.
96. Cai L, Sutter BM, Li B, Tu BP. Acetyl-CoA induces cell growth and proliferation by promoting the acetylation of histones at growth genes. *Mol Cell*. 2011;42(4):426–437.
97. Wellen KE, Hatzivassiliou G, Sachdeva UM, Bui TV, Cross JR, Thompson CB. ATP-citrate lyase links cel-

- lular metabolism to histone acetylation. *Science*. 2009;324(5930):1076–1080.
98. Chalkiadaki A, Guarante L. The multifaceted functions of sirtuins in cancer. *Nat Rev Cancer*. 2015;15(10):608–624.
 99. Mardis ER, Ding L, Dooling DJ, et al. Recurring mutations found by sequencing an acute myeloid leukemia genome. *N Engl J Med*. 2009;361(11):1058–1066.
 100. Yan H, Parsons DW, Jin G, et al. IDH1 and IDH2 mutations in gliomas. *N Engl J Med*. 2009;360(8):765–773.
 101. Zhao S, Lin Y, Xu W, et al. Glioma-derived mutations in IDH1 dominantly inhibit IDH1 catalytic activity and induce HIF-1alpha. *Science*. 2009;324(5924):261–265.
 102. Dang L, White DW, Gross S, et al. Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. *Nature*. 2009;462(7274):739–744.
 103. Xu W, Yang H, Liu Y, et al. Oncometabolite 2-hydroxyglutarate is a competitive inhibitor of alpha-ketoglutarate-dependent dioxygenases. *Cancer Cell*. 2011;19(1):17–30.
 104. Lu C, Ward PS, Kapoor GS, et al. IDH mutation impairs histone demethylation and results in a block to cell differentiation. *Nature*. 2012;483(7390):474–478.
 105. Wang F, Travins J, DeLaBarre B, et al. Targeted inhibition of mutant IDH2 in leukemia cells induces cellular differentiation. *Science*. 2013;340(6132):622–626.
 106. Rohle D, Popovici-Muller J, Palaskas N, et al. An inhibitor of mutant IDH1 delays growth and promotes differentiation of glioma cells. *Science*. 2013;340(6132):626–630.
 107. Carey BW, Finley LW, Cross JR, Allis CD, Thompson CB. Intracellular alpha-ketoglutarate maintains the pluripotency of embryonic stem cells. *Nature*. 2015;518(7539):413–416.
 108. Pan M, Reid MA, Lowman XH, et al. Regional glutamine deficiency in tumours promotes dedifferentiation through inhibition of histone demethylation. *Nat Cell Biol*. 2016;18(10):1090–1101.
 109. Xiao M, Yang H, Xu W, et al. Inhibition of alpha-KG-dependent histone and DNA demethylases by fumarate and succinate that are accumulated in mutations of FH and SDH tumor suppressors. *Genes Dev*. 2012;26(12):1326–1338.
 110. Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell*. 2012;21(3):309–322.
 111. Less JR, Skalak TC, Sevick EM, Jain RK. Microvascular architecture in a mammary carcinoma: branching patterns and vessel dimensions. *Cancer Res*. 1991;51(1):265–273.
 112. McKeown SR. Defining normoxia, physoxia and hypoxia in tumours-implications for treatment response. *Br J Radiol*. 2014;87(1035):20130676.
 113. Rumsey WL, Schlosser C, Nuutinen EM, Robiolio M, Wilson DF. Cellular energetics and the oxygen dependence of respiration in cardiac myocytes isolated from adult rat. *J Biol Chem*. 1990;265(26):15392–15402.
 114. Rankin EB, Giaccia AJ. The role of hypoxia-inducible factors in tumorigenesis. *Cell Death Differ*. 2008;15(4):678–685.
 115. Nakazawa MS, Keith B, Simon MC. Oxygen availability and metabolic adaptations. *Nat Rev Cancer*. 2016;16(10):663–673.
 116. Semenza GL. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer*. 2003;3(10):721–732.
 117. Zeng W, Wan R, Zheng Y, Singh SR, Wei Y. Hypoxia, stem cells and bone tumor. *Cancer Lett*. 2011;313(2):129–136.
 118. Lee KE, Simon MC. From stem cells to cancer stem cells: HIF takes the stage. *Curr Opin Cell Biol*. 2012;24(2):232–235.
 119. Kamphorst JJ, Nofal M, Commissio C, et al. Human pancreatic cancer tumors are nutrient poor and tumor cells actively scavenge extracellular protein. *Cancer Res*. 2015;75(3):544–553.
 120. Reid MA, Wang WI, Rosales KR, Welliver MX, Pan M, Kong M. The B55alpha subunit of PP2A drives a p53-dependent metabolic adaptation to glutamine deprivation. *Mol Cell*. 2013;50(2):200–211.
 121. Reid MA, Lowman XH, Pan M, et al. IKKbeta promotes metabolic adaptation to glutamine deprivation via phosphorylation and inhibition of PFKFB3. *Genes Dev*. 2016;30(16):1837–1851.
 122. Tran TQ, Lowman XH, Reid MA, et al. Tumor-associated mutant p53 promotes cancer cell survival upon glutamine deprivation through p21 induction. *Oncogene*. 2016.
 123. Maddocks OD, Berkers CR, Mason SM, et al. Serine starvation induces stress and p53-dependent metabolic remodelling in cancer cells. *Nature*. 2013;493(7433):542–546.
 124. Hensley CT, Faubert B, Yuan Q, et al. Metabolic heterogeneity in human lung tumors. *Cell*. 2016;164(4):681–694.
 125. Swietach P, Vaughan-Jones RD, Harris AL. Regulation of tumor pH and the role of carbonic anhydrase 9. *Cancer Metastasis Rev*. 2007;26(2):299–310.
 126. Martinez-Zagulian R, Seftor EA, Seftor RE, Chu YW, Gillies RJ, Hendrix MJ. Acidic pH enhances the invasive behavior of human melanoma cells. *Clin Exp Metastasis*. 1996;14(2):176–186.
 127. Rothberg JM, Bailey KM, Wojtkowiak JW, et al. Acid-mediated tumor proteolysis: contribution of cysteine cathepsins. *Neoplasia*. 2013;15(10):1125–1137.
 128. Colegio OR, Chu NQ, Szabo AL, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nature*. 2014;513(7519):559–563.
 129. Wahl DR, Byersdorfer CA, Ferrara JL, Opiplari AW Jr, Glick GD. Distinct metabolic programs in activated T cells: opportunities for selective immunomodulation. *Immunol Rev*. 2012;249(1):104–115.
 130. Ron-Harel N, Sharpe AH, Haigis MC. Mitochondrial metabolism in T cell activation and senescence: a mini-review. *Gerontology*. 2015;61(2):131–138.
 131. Mellor AL, Munn DH. Creating immune privilege: active local suppression that benefits friends, but protects foes. *Nat Rev Immunol*. 2008;8(1):74–80.
 132. Fallarino F, Grohmann U, Vacca C, et al. T cell apoptosis by tryptophan catabolism. *Cell Death Differ*. 2002;9(10):1069–1077.
 133. Fooksman DR, Vardhana S, Vasiliver-Shamis G, et al. Functional anatomy of T cell activation and synapse formation. *Annu Rev Immunol*. 2010;28:79–105.

134. Patsoukis N, Bardhan K, Chatterjee P, et al. PD-1 alters T-cell metabolic reprogramming by inhibiting glycolysis and promoting lipolysis and fatty acid oxidation. *Nat Commun.* 2015;6:6692.
135. Clambey ET, McNamee EN, Westrich JA, et al. Hypoxia-inducible factor-1 alpha-dependent induction of FoxP3 drives regulatory T-cell abundance and function during inflammatory hypoxia of the mucosa. *Proc Natl Acad Sci USA.* 2012;109(41):E2784–E2793.
136. Kouidhi S, Noman MZ, Kieda C, Elgaaied AB, Chouaib S. Intrinsic and tumor microenvironment-induced metabolism adaptations of t cells and impact on their differentiation and function. *Front Immunol.* 2016;7:11