

Bölüm 3

TÜMÖR KAÇIŞ MEKANİZMALARI VE MEDİYATÖRLERİ

Gökhan AYTEKİN¹
Selma Erol AYTEKİN²

GİRİŞ

İmmünoloji ve onkoloji alanları 19. Yüzyılın sonlarına doğru sıkı bir birlilikçi içine girmiştir. Özellikle Willian Colley'in bir sarkom hastasında sarkom bölgelerine ölü bakteri enjeksiyonu sonrasında sarkomda küçülme göstergesi, bu birlilikçi başlangıcı kabul edilmektedir (1). O zamandan bu yana tüm kanser türlerinde immün süryevans (gözetim), immün yeniden düzenleme (editing) ve tümör büyümesi ve gelişmesi arasındaki ilişkiye araştırmaya yönelik birçok çalışma yapılmış, bu alanda birçok yeni gelişme yaşanmıştır. Kanser hücreleri temel olarak normal hücresel işlemler ile kontrol edilemeyen ve kontrol dışı çoğalan self hücrelerdir. Bu abnormal hücreler çevrelerindeki değişikliklere daha iyi adapte olmalarını sağlayan genetik hatalara yatkındır. Darwin'in doğal seçelim teorisinde olduğu gibi, tümör hücreleri üzerindeki immün baskısı, immün tanınmaya karşı dirençli tümör varyant hücrelerin seçiminini sağlar.

İmmün tanıma, immün sistemin birbirini tamamlayan iki kolu tarafından sağlanır. Adaptif-hümoral immün sistem transforme hücreleri self, non-self ayrimı yaparak tespit eder. CD8+ T lenfositler sitotoksik T lenfosit (CTL), CD4+ T lenfositler Th1/Th2 alt grupları ile beraber yardımcı T lenfositler olarak adlandırılır. Adaptif immün sistemin sitotoksik T hücreleri tüm çekirdekli hücrelerden eksprese edilen MHC I (Major Histo Compatability class I) molekülü ile eksprese edilen抗原leri tanırlar. Eğer bir tümör hücresi MHC I ile viral ya da anormal protein eksprese ederse抗原 spesifik CTL hücrelerce eliminé edilir. MHC II ise

¹ Uzm. Dr., Necmettin Erbakan Üniversitesi Meram Tip Fakültesi, İmmünoloji ve Alerji BD
ayteking@gmail.com

² Uzm. Dr., Necmettin Erbakan Üniversitesi Meram Tip Fakültesi Çocuk İmmünoloji ve Alerji BD
Dr.selmaerol@gmail.com

KAYNAKLAR

1. Coley WB. The treatment of malignant tumors by repeated inoculations of erysipelas. With a report of ten original cases. 1893. *Clin Orthop Relat Res.* 1991;262(3):3-11.
2. Schwartz RH. A cell culture model for T lymphocyte clonal anergy. *Science.* 1990;248(4961):1349-56.
3. Gras Navarro A, Bjorklund AT, Chekenya M. Therapeutic potential and challenges of natural killer cells in treatment of solid tumors. *Front Immunol.* 2015;6:202.
4. Savage PA, Leventhal DS, Malchow S. Shaping the repertoire of tumor-infiltrating effector and regulatory T cells. *Immunol Rev.* 2014;259(1):245-58.
5. Marvel D, Gabrilovich DI. Myeloid-derived suppressor cells in the tumor microenvironment: expect the unexpected. *J Clin Invest.* 2015;125(9):3356-64.
6. Bailey SR, Nelson MH, Himes RA, Li Z, Mehrotra S, Paulos CM. Th17 cells in cancer: the ultimate identity crisis. *Front Immunol.* 2014;5:276.
7. Laoui D, Van Overmeire E, De Baetselier P, Van Ginderachter JA, Raes G. Functional Relationship between Tumor-Associated Macrophages and Macrophage Colony-Stimulating Factor as Contributors to Cancer Progression. *Front Immunol.* 2014;5:489.
8. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science.* 2011;331(6024):1565-70.
9. Tran E, Turcotte S, Gros A, Robbins PF, Lu YC, Dudley ME, et al. Cancer immunotherapy based on mutation-specific CD4+ T cells in a patient with epithelial cancer. *Science.* 2014;344(6184):641-5.
10. Matsushita H, Vesely MD, Koboldt DC, Rickert CG, Uppaluri R, Magrini VJ, et al. Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoediting. *Nature.* 2012;482(7385):400-4.
11. Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, et al. Immune evasion in cancer: Mechanistic basis and therapeutic strategies. *Semin Cancer Biol.* 2015;35 Suppl:S185-S98.
12. Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N. Molecular and genetic properties of tumors associated with local immune cytolytic activity. *Cell.* 2015;160(1-2):48-61.
13. Catalan E, Charni S, Jaime P, Aguiló JI, Enriquez JA, Naval J, et al. MHC-I modulation due to changes in tumor cell metabolism regulates tumor sensitivity to CTL and NK cells. *Oncimmunology.* 2015;4(1):e985924.
14. Zaretsky JM, Garcia-Diaz A, Shin DS, Escuin-Ordinatas H, Hugo W, Hu-Lieskovian S, et al. Mutations Associated with Acquired Resistance to PD-1 Blockade in Melanoma. *N Engl J Med.* 2016;375(9):819-29.
15. Johnsen AK, Templeton DJ, Sy M, Harding CV. Deficiency of transporter for antigen presentation (TAP) in tumor cells allows evasion of immune surveillance and increases tumorigenesis. *J Immunol.* 1999;163(8):4224-31.
16. Reichel J, Chadburn A, Rubinstein PG, Giulino-Roth L, Tam W, Liu Y, et al. Flow sorting and exome sequencing reveal the oncogenome of primary Hodgkin and Reed-Sternberg cells. *Blood.* 2015;125(7):1061-72.
17. Rouas-Freiss N, Moreau P, Menier C, Carosella ED. HLA-G in cancer: a way to turn off the immune system. *Semin Cancer Biol.* 2003;13(5):325-36.
18. Latchman Y, Wood CR, Chernova T, Chaudhary D, Borde M, Chernova I, et al. PD-L2 is a second ligand for PD-1 and inhibits T cell activation. *Nat Immunol.* 2001;2(3):261-8.
19. Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J Exp Med.* 2000;192(7):1027-34.
20. Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature.* 2014;515(7528):568-71.
21. Ott PA, Hodi FS, Robert C. CTLA-4 and PD-1/PD-L1 blockade: new immunotherapeutic modalities with durable clinical benefit in melanoma patients. *Clin Cancer Res.* 2013;19(19):5300-9.

22. Fiore E, Fusco C, Romero P, Stamenkovic I. Matrix metalloproteinase 9 (MMP-9/gelatinase B) proteolytically cleaves ICAM-1 and participates in tumor cell resistance to natural killer cell-mediated cytotoxicity. *Oncogene*. 2002;21(34):5213-23.
23. Joon Yun A, Bazar KA, Lee PY. Tumors may modulate host immunity partly through hypoxia-induced sympathetic bias. *Med Hypotheses*. 2004;63(2):352-6.
24. Li R, Ruttinger D, Li R, Si LS, Wang YL. Analysis of the immunological microenvironment at the tumor site in patients with non-small cell lung cancer. *Langenbecks Arch Surg*. 2003;388(6):406-12.
25. Amend SR, Pienta KJ. Ecology meets cancer biology: the cancer swamp promotes the lethal cancer phenotype. *Oncotarget*. 2015;6(12):9669-78.
26. Guo F, Wang Y, Liu J, Mok SC, Xue F, Zhang W. CXCL12/CXCR4: a symbiotic bridge linking cancer cells and their stromal neighbors in oncogenic communication networks. *Oncogene*. 2016;35(7):816-26.
27. Levy DE, Lee CK. What does Stat3 do? *J Clin Invest*. 2002;109(9):1143-8.
28. Coffer PJ, Koenderman L, de Groot RP. The role of STATs in myeloid differentiation and leukemia. *Oncogene*. 2000;19(21):2511-22.
29. Wang T, Niu G, Kortylewski M, Burdelya L, Shain K, Zhang S, et al. Regulation of the innate and adaptive immune responses by Stat-3 signaling in tumor cells. *Nat Med*. 2004;10(1):48-54.
30. Catlett-Falcone R, Landowski TH, Oshiro MM, Turkson J, Levitzki A, Savino R, et al. Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity*. 1999;10(1):105-15.
31. Kelly JM, Takeda K, Darcy PK, Yagita H, Smyth MJ. A role for IFN-gamma in primary and secondary immunity generated by NK cell-sensitive tumor-expressing CD80 in vivo. *J Immunol*. 2002;168(9):4472-9.
32. Qin Z, Schwartzkopff J, Pradera F, Kammertoens T, Seliger B, Pircher H, et al. A critical requirement of interferon gamma-mediated angiostasis for tumor rejection by CD8+ T cells. *Cancer Res*. 2003;63(14):4095-100.
33. Ruiz-Ruiz C, Ruiz de Almodovar C, Rodriguez A, Ortiz-Ferron G, Redondo JM, Lopez-Rivas A. The up-regulation of human caspase-8 by interferon-gamma in breast tumor cells requires the induction and action of the transcription factor interferon regulatory factor-1. *J Biol Chem*. 2004;279(19):19712-20.
34. Nagao M, Nakajima Y, Kanehiro H, Hisanaga M, Aomatsu Y, Ko S, et al. The impact of interferon gamma receptor expression on the mechanism of escape from host immune surveillance in hepatocellular carcinoma. *Hepatology*. 2000;32(3):491-500.
35. Ferrara N. VEGF: an update on biological and therapeutic aspects. *Curr Opin Biotechnol*. 2000;11(6):617-24.
36. Santin AD, Hermonat PL, Ravaggi A, Cannon MJ, Pecorelli S, Parham GP. Secretion of vascular endothelial growth factor in ovarian cancer. *Eur J Gynaecol Oncol*. 1999;20(3):177-81.
37. Wang D, Dubois RN. Cyclooxygenase-2: a potential target in breast cancer. *Semin Oncol*. 2004;31(1 Suppl 3):64-73.
38. Eisengart CA, Mestre JR, Naama HA, Mackrell PJ, Rivadeneira DE, Murphy EM, et al. Prostaglandins regulate melanoma-induced cytokine production in macrophages. *Cell Immunol*. 2000;204(2):143-9.
39. Hug H. Fas-mediated apoptosis in tumor formation and defense. *Biol Chem*. 1997;378(12):1405-12.
40. Walker PR, Saas P, Dietrich PY. Role of Fas ligand (CD95L) in immune escape: the tumor cell strikes back. *J Immunol*. 1997;158(10):4521-4.
41. Hahne M, Rimoldi D, Schroter M, Romero P, Schreier M, French LE, et al. Melanoma cell expression of Fas(Apo-1/CD95) ligand: implications for tumor immune escape. *Science*. 1996;274(5291):1363-6.
42. Balachandran VP, Cavagnar MJ, Zeng S, Bamboat ZM, Ocuin LM, Obaid H, et al. Imatinib potentiates antitumor T cell responses in gastrointestinal stromal tumor through the inhibition of Ido. *Nat Med*. 2011;17(9):1094-100.

43. Spranger S, Bao R, Gajewski TF. Melanoma-intrinsic beta-catenin signalling prevents anti-tumour immunity. *Nature*. 2015;523(7559):231-5.
44. Yi X, Yin XM, Dong Z. Inhibition of Bid-induced apoptosis by Bcl-2. tBid insertion, Bax translocation, and Bax/Bak oligomerization suppressed. *J Biol Chem*. 2003;278(19):16992-9.
45. Sauter B, Albert ML, Francisco L, Larsson M, Somersan S, Bhardwaj N. Consequences of cell death: exposure to necrotic tumor cells, but not primary tissue cells or apoptotic cells, induces the maturation of immunostimulatory dendritic cells. *J Exp Med*. 2000;191(3):423-34.
46. Kadowaki N, Ho S, Antonenko S, Malefyt RW, Kastelein RA, Bazan F, et al. Subsets of human dendritic cell precursors express different toll-like receptors and respond to different microbial antigens. *J Exp Med*. 2001;194(6):863-9.
47. Fujii S, Liu K, Smith C, Bonito AJ, Steinman RM. The linkage of innate to adaptive immunity via maturing dendritic cells in vivo requires CD40 ligation in addition to antigen presentation and CD80/86 costimulation. *J Exp Med*. 2004;199(12):1607-18.
48. Cella M, Scheidegger D, Palmer-Lehmann K, Lane P, Lanzavecchia A, Alber G. Ligation of CD40 on dendritic cells triggers production of high levels of interleukin-12 and enhances T cell stimulatory capacity: T-T help via APC activation. *J Exp Med*. 1996;184(2):747-52.
49. Ciaravino G, Bhat M, Manbeian CA, Teng NN. Differential expression of CD40 and CD95 in ovarian carcinoma. *Eur J Gynaecol Oncol*. 2004;25(1):27-32.
50. Jakobson E, Jonsson G, Bjorck P, Paulie S. Stimulation of CD40 in human bladder carcinoma cells inhibits anti-Fas/APO-1 (CD95)-induced apoptosis. *Int J Cancer*. 1998;77(6):849-53.
51. Loro LL, Ohlsson M, Vintermyr OK, Liavaag PG, Jonsson R, Johannessen AC. Maintained CD40 and loss of polarised CD40 ligand expression in oral squamous cell carcinoma. *Anticancer Res*. 2001;21(1A):113-7.
52. Hu K, Chen F. Identification of significant pathways in gastric cancer based on protein-protein interaction networks and cluster analysis. *Genet Mol Biol*. 2012;35(3):701-8.
53. Yamaguchi H, Tanaka F, Sadanaga N, Ohta M, Inoue H, Mori M. Stimulation of CD40 inhibits Fas- or chemotherapy-mediated apoptosis and increases cell motility in human gastric carcinoma cells. *Int J Oncol*. 2003;23(6):1697-702.
54. Pammer J, Plettenberg A, Weninger W, Diller B, Mildner M, Uthman A, et al. CD40 antigen is expressed by endothelial cells and tumor cells in Kaposi's sarcoma. *Am J Pathol*. 1996;148(5):1387-96.
55. Sabel MS, Yamada M, Kawaguchi Y, Chen FA, Takita H, Bankert RB. CD40 expression on human lung cancer correlates with metastatic spread. *Cancer Immunol Immunother*. 2000;49(2):101-8.
56. Flaxenburg JA, Melter M, Lapchak PH, Briscoe DM, Pal S. The CD40-induced signaling pathway in endothelial cells resulting in the overexpression of vascular endothelial growth factor involves Ras and phosphatidylinositol 3-kinase. *J Immunol*. 2004;172(12):7503-9.
57. Groh V, Wu J, Yee C, Spies T. Tumour-derived soluble MIC ligands impair expression of NKG2D and T-cell activation. *Nature*. 2002;419(6908):734-8.