

ELEKTROFİZYOLOJİ

Sami YILMAZ¹

1. Giriş

Göz yüzeyi, periorbital cilt, ya da kafa derisi üzerine yerleştirilen elektrotlar kullanılarak, görsel uyarınların elektrik potansiyellerine çevrilerek kaydedilmesi işlemi, gözün ve görme yollarının elektrofizyolojik değerlendirilmesi olarak isimlendirilir. Görme sistemi içerisinde farklı konumlarda yer alan hücre türleri ile ilgili girişimsel olmayan, objektif bilgiler sağlar. Uluslararası Klinik Görsel Elektrofizyoloji Derneği (ISCEV) gözün ve görme yollarının elektrofizyolojik değerlendirilmesi için standartlar ve rehberler yayımlamaktadır (1-5).

Santral siniri sisteminin geri kalanı gibi, retina da embriyolojik olarak nöral tüpten köken alır (6). Retina çevresel enerjiyi elektrik potansiyellerine çeviren özelleşmiş bir hücre yapısına sahiptir. Retina iki kısımdan oluşmaktadır: 1) Işığın elektrik potansiyellerine çeviren fotoreseptör hücreler olarak adlandırılan rod (basil) ve kon (koni) hücrelerinden oluşan sensöriyel (duyusal) retina ve 2) bipolar, horizontal, amakrin ve ganglion hücrelerinden oluşan nöral retina (7).

Retina bu özelleşmiş hücrelerin çekirdeklerinden ve onların uzantılarından oluşmuş düzenli bir tabakalı yapıya sahiptir. Retina pigment epitel tabakasının hemen üzerinde yerleşen dış nükleer tabaka (DNL), rod ve kon hücrelerinin hücre gövdelerini içermekte, iç nükleer tabaka (İNL) horizontal, bipolar, amakrin ve glial hücrelerin (Müller hücreleri) hücre gövdelerinden oluşmaktadır. Ganglion ve amakrin hücre gövdelerinden oluşan ganglion hücre tabakası (GHT) vitreusa yakın yüzeyde yer almaktadır. Ganglion hücrelerin aksonları optik siniri oluşturur. Bu hücre gövdelerinden oluşan 3 tabaka arasında hücrelerin dentrit

¹ Op. Dr. Sami YILMAZ, Bursa Retina Göz Hastanesi samifmf1978@yahoo.com

Kaynaklar

1. McCulloch DL, Marmor MF, Brigell MG, et al. ISCEV standart for full-field clinical electroretinography (2015 update). *Doc Ophthalmol* 2015;130:1-12.
2. Bach M, Brigell MG, Hawlina M, et al. ISCEV standart for clinical pattern electroretinography (PERG): 2012 update. *Doc Ophthalmol* 2013;126:1-7.
3. Hood DC, Bach M, Brigell M, et al. ISCEV standart for clinical multifocal electroretinography (mfERG) (2011 edition). *Doc Ophthalmol* 2012;124:1-13.
4. Constable PA, Bach M, Frishman LJ, et al. International society for clinical electrophysiology of vision. ISCEV standart for clinical electro-oculography (2017 update). *Doc Ophthalmol* 2017;134:1-9.
5. Odom JV, Bach M, Brigell M, et al. ISCEV standart for clinical visual evoked potentials (2016 update). *Doc Ophthalmol* 2016;133(1):1-9.
6. O'Rahilly R, Müller F. Neurulation in the normal human embryo. *Ciba Found Symp* 1994;181:70-82, discussion 82-9.
7. Masland RH. The functional architecture of the retina. *Sci Am* 1986;255(6):102-11.
8. Wassle H. Parallel processing in the mammalian retina. *Nat Rev Neurosci* 2004;5(10):747-57.
9. Curcio CA, Sloan KR, Kalina RE, Hendrickson AE. Human photoreceptor topography. *J Comp Neurol* 1990;292(4):497-523.
10. Ahnelt PK, Kolb H. The mammalian photoreceptor mosaic-adaptive design. *Prog Retin Eye Res*. 2000;19(6):711-77.
11. DeMonasterio FM, Schein SJ, McCrane EP. Staining of blue-sensitive cones of the macaque retina by a fluorescent dye. *Science* 1981;213(4513):1278-81.
12. Chun MH, Grunert U, Martin PR, Wässle H. The synaptic complex of cones in the fovea and in the periphery of the macaque monkey retina. *Vision Res* 1996;36(21):3383-95.
13. Lennie P, Fairchild MD. Ganglion cell pathways for rod vision. *Vision Res* 1994;34(4):477-82.
14. Mills SL, Massey SC. AII amacrine cells limit scotopic acuity in central macaque retina: A confocal analysis of calretinin labeling. *J Comp Neurol* 1999;411(1):19-34.
15. Rao-Miroznik R, Harkins AB, Buchsbaum G, Sterling P. Mammalian rod terminal: Architecture of a binary synapse. *Neuron* 1995;14(3):561-9.
16. Verweij J, Kamermans M, Spekreijse H. Horizontal cells feed back to cones by shifting the cone calcium-current activation range. *Vision Res* 1996;36(24):3943-53.
17. Sterling P. How retinal circuits optimize the transfer of visual information. In: Chalupa LM, editor. *The visual neurosciences*. Boston, MA: Cambridge; 2004. p. 234-59.
18. Kamermans M, Kraaij DA, Spekreijse H. The cone/horizontal cell network: a possible site for color constancy. *Vis Neurosci* 1998;15(5):787-97.
19. Dacey DM, Lee BB, Stafford DK, et al. Horizontal cells of the primate retina: cone specificity without spectral opponency. *Science* 1996;271(5249):565-9.
20. Packer OS, Dacey DM. Receptive field structure of H1 horizontal cells in macaque

- monkey retina. *J Vis* 2002;2(4):272-92.
21. Boycott BB, Wassle H. Morphological classification of bipolar cells of the primate retina. *Eur J Neurosci* 1991;3(11):1069-88.
 22. Schubert T, Kerschensteiner D, Eggers ED, et al. Development of the presynaptic inhibition onto retinal bipolar cell axon terminals is subclass-specific. *J Neurophysiol* 2008;100(1):304-16.
 23. Dhingra A, Lyubarsky A, Jiang M, et al. The light response of ON bipolar neurons requires G_{alpha}i/o. *J Neurosci* 2000;20(24):9053-8.
 24. MacNeil AA, Masland RH. Extreme diversity among amacrine cells: implications for function. *Neuron* 1998;20(5):971-82.
 25. Vardi N, Smith RG. The AII amacrine network: coupling can increase correlated activity. *Vision Res* 1996;36(23):3743-57.
 26. Sanes JR, Masland RH. The types of retinal ganglion cells: current status and implications for neuronal classification. *Annu Rev Neurosci* 2015;38:221-46.
 27. Gjötterberg M. Electrodes for electroretinography: a comparison of four different types. *Arch Ophthalmol* 1986;104:569-70.
 28. Brown KT. The electroretinogram: its components and their origins. *Vision Res* 1968;8:633-77.
 29. Newman EA, Odette LL. Model of electroretinogram b-wave generation: a test of the K⁺ hypothesis. *J Neurophysiol* 1984;51:164-82.
 30. Wachtmeister L, Dowling JE. The oscillatory potentials of the mudpuppy retina. *Invest Ophthalmol Vis Sci* 1978;17:1176-88.
 31. Kondo M, Piao CH, Tanikawa A, et al. Amplitude decrease of photopic ERG b-wave at higher stimulus intensities in humans. *Jpn J Ophthalmol* 2000;44:20-8.
 32. Peachey NS, Alexander KR, Fishman GA, Derlacki DJ. Properties of the human cone system electroretinogram during light adaptation. *Appl Opt* 1989;28:1145-50.
 33. Berninger T, Schuurmans RP. Spatial tuning of the pattern ERG across temporal frequency. *Doc Ophthalmol* 1985;61:17-25.
 34. Marmor MF, Holder GE, Porciatti V, et al. Guidelines for basic pattern electroretinography: recommendations by the International Society for Clinical Electrophysiology of Vision. *Doc Ophthalmol* 1995-1996;91:291-8.
 35. Holder GE. PERG abnormalities in anterior visual pathway disease. *Electroencephalogr Clin Neurophysiol* 1985;61:365.
 36. Sutter EE, Tran D. The field topography of ERG components in man-I. The photopic luminance response. *Vision Res* 1992;32:433-46.
 37. Sutter E. The interpretation of multifocal binary kernels. *Doc Ophthalmol* 2000;100:49-75.
 38. Hood DC, Freishman LJ, Saszik S, et al. Retinal origins of the primate multifocal ERG: implications for the human response. *Invest Ophthalmol Vis Sci* 2002;43:1673-85.
 39. Du Bois Reymond EH. Chapter 3. Von dem ruhen Nervenstrom. Untersuchungen

- Über Thierische Electricität, vol. 2. Berlin: G Reimer; 1849. 251-88.
40. Arden GB, Barrada A, Kelsey JH. New clinical test of retinal function based upon the standing potential of the eye. *Br J Ophthalmol* 1962;46:449-67.
 41. Robson AG, Nilsson J, Li S, et al. ISCEV guide to visual electrodiagnostic procedures. *Doc Ophthalmol* 2018; 136:1-26.
 42. Weleber RG, Watzke RC, Shults WT, et al. Clinical and electrophysiologic characterization of paraneoplastic and autoimmune retinopathies associated with anti-enolase antibodies. *Am J Ophthalmol* 2005;139:780-94.
 43. Galloway NR, Barber C. The pattern evoked response in disorders of the optic nerve. *Doc Ophthalmol* 1986;63:31-6.
 44. Holder GE. The incidence of abnormal pattern electroretinography in optic nerve demyelination. *Electroenceph Clin Neurophysiol* 1991;78:18-26.
 45. Thompson PD, Mastaglia FL, Carroll WM. Anterior ischaemic optic neuropathy: a correlative clinical and visual evoked potential study of 18 patients. *J Neurol Neurosurg Psychiatry* 1986;49:128-35.
 46. Livingstone IR, Mastaglia FL, Howe JW, Aherne GE. Leber's optic neuropathy: clinical and visual evoked response studies in asymptomatic and symptomatic members of a 4 generation family. *Br J Ophthalmol* 1980;64(10):751-7.
 47. Flanagan JG, Harding GF. Multichannel visual evoked potentials in early compressive lesions of the chiasma. *Doc Ophthalmol* 1988;69:271-81.
 48. Holder GE, Votruba M, Carter AC, et al. Electrophysiological findings in dominant optic atrophy linking to the OPA1 locus on chromosome 3q 28-qter. *Doc Ophthalmol* 1998-1999;95:217-28.
 49. Troncoso J, Mancall EL, Schatz NY. Visual evoked response in pernicious anemia. *Arch Neurol* 1979;36:168-9.
 50. Bodis WI, Atkin A, Raab E, Wolkstein M. Visual associated cortex and vision in man: pattern evoked occipital potentials in a blind boy. *Science* 1977;198:629-31.
 51. Robson JG, Frishman LJ. Response linearity and kinetics of the cat retina: the bipolar cell component of the dark-adapted electroretinogram. *Vis Neurosci* 1995;12:837-50.
 52. Berson EL. Retinitis pigmentosa and allied disease: applications of electroretinographic testing. *Int Ophthalmol* 1981;4:7-22.
 53. Schubert G, Bornschein H. Analysis of the human retinogram. *Ophthalmologica* 1952;123(6):396-413.
 54. Riggs LA. Electroretinography in cases of night blindness. *Am J Ophthalmol* 1954;38(1:2):70-8.
 55. Carr RE, Ripps H, Siegel IM. Visual pigment kinetics and adaptation in fundus albipunctatus. In: Dodt E, Pearlman JT (eds) XIth I.S.C.E.R.G. Symposium. *Documenta Ophthalmologica Proceedings Series*, Springer, Dordrecht, vol 4. 1974:193-204.
 56. Niwa Y, Kondo M, Ueno S, et al. Cone and rod dysfunction in fundus albipunctatus with RDH5 mutation: an electrophysiological study. *Invest Ophthalmol Vis Sci* 2005;46(4):1480-5.

57. Carr RE, Gouras P. Oguchi's disease. *Arch Ophthalmol* 1965;73:646-56.
58. McBain VA, Egan CA, Pieris SJ, et al. Functional observations in vitamin A deficiency: diagnosis and time course of recovery. *Eye (Lond)* 2007;21(3):367-76.
59. Miyake Y, Yagasaki K, Horiguchi M, et al. Congenital stationary night blindness with negative electroretinogram: a new classification. *Arch Ophthalmol* 1986;104(7):1013-20.
60. Miyake Y. [Establishment of the concept of new clinical entities complete and incomplete form of congenital stationary night blindness]. *Nihon Ganka Gakkai Zasshi* 2002;106(12):737-55, discussion 56.
61. Alexander KR, Fishman GA, Peachey NS, et al. "On" response defect in paraneoplastic night blindness with cutaneous malignant melenoma. *Invest Ophthalmol Vis Sci* 1992;33(3):477-83.
62. Simunovic MP, Moore AT. The cone dystrophies. *Eye (Lond)* 1998;12(Pt 3b):553-65.
63. Robson AG, Michaelides M, Luong VA, et al. Functional correlates of fundus autofluorescence abnormalities in patients with RPGR or RIMS 1 mutations causing cone or cone rod dystrophy. *Br J Ophthalmol* 2008;92(1):95-102.
64. O'Connor PS, Tredici TJ, Ivan DJ, et al. Achromatopsia: clinical diagnosis and treatment. *J Clin Neuroophthalmol* 1982;2(4):219-26.
65. Gouras P, MacKay CJ. Electroretinographic responses of the short-wavelength-sensitive cones. *Invest Ophthalmol Vis Sci* 1990;31(7):1203-9.
66. Hirose T, Katsumi O, Pruett RC, et al. Retinal function in birdshot retinochoroidopathy. *Acta Ophthalmol (Copenh)* 1991;69(3):327-37.
67. Gass JD, Agarwal A, Scott IU. Acute zonal occult outer retinopathy: a long-term follow-up study. *Am J Ophthalmol* 2002;134(3):329-39.
68. Henkes HE. Electroretinography in circulatory disturbances of the retina. II: The electroretinogram in cases of occlusion of the central retinal artery or of its branches. *Arch Ophthalmol* 1954;51(1):42-53.
69. Ventura LM, Porciatti V. Pattern electroretinogram in glaucoma. *Curr Opinion in Ophthalmol* 2006;17:196-202.
70. Good PA, Searle AE, Campbell S, Crews SJ. Value of the ERG in congenital nystagmus. *Br J Ophthalmol* 1989;73(7):512-5.
71. Hayreh SS, Klugman MR, Podhajsky P, Kolder HE. Electroretinography in central retinal vein occlusion. Correlation of electroretinographic changes with pupillary abnormalities. *Graefes Arch Clin Exp Ophthalmol* 1989;227(6):549-61.
72. Brown GC, Magargal LE. The ocular ischemic syndrome. *Int Ophthalmol* 1988;11(4):239-51.
73. Tzekov R, Arden GB. The electroretinogram in diabetic retinopathy. *Surv Ophthalmol* 1999;44(1):53-60.
74. Galloway NR. Electrophysiological testing of eyes with opaque media. *Eye* 1988;2:615-24.
75. Fishman GA, Weinberg AB, McMahon TT. X-linked recessive retinitis pigmentosa. Clinical characteristics of carriers. *Arch Ophthalmol* 1986;104(9):1329-35.

76. Holder GE, Robson AG, Pavesio C, et al. Electrophysiological characterisation and monitoring in the management of birdshot chorioretinopathy. *Br J Ophthalmol* 2005;89(6):709-18.
77. Maturi RK, Yu M, Weleber RG. Multifocal electroretinographic evaluation of long-term hydroxychloroquine users. *Arch Ophthalmol* 2004;122(7):973-81.
78. Imaizumi M, Matsumoto CS, Yamada K, et al. Electroretinographic assessment of early changes in ocular siderosis. *Ophthalmologica* 2000;214(5):354-9.
79. Brodie SE. Tips and tricks for successful electroretinography in children. *Curr Opin Ophthalmol* 2014;25(5):366-73.
80. Sokol S. Abnormal evoked potential latencies in amblyopia. *Br J Ophthalmol* 1983;67:310-4.
81. Lambert SR, Kriss A, Taylor D. Delayed visual maturation: a longitudinal clinical and electrophysiological assessment. *Ophthalmology* 1989;96(4):524-9.
82. Ponjavic V, Andréasson S. Multifocal ERG and full-field ERG in patients on long-term vigabatrin medication. *Doc Ophthalmol* 2001;102(1):63-72.
83. Cooper LZ, Krugman S. Clinical manifestations of postnatal and congenital rubella. *Arch Ophthalmol* 1967;77(4):434-9.
84. Berger W, Kloeckener-Gruissem B, Neidhart J. The molecular basis of human retinal and vitreoretinal disease. *Prog Retin Eye Res* 2010;29:335-75.
85. HGMD. Human Gene Mutation Database, <<http://www.hgmd.cf.ac.uk/>>.
86. Nussbaum RL, McInnes RR, Williard HF. Thompson and Thompson Genetics in medicine. 8th ed. Philadelphia, PA: Saunders Elsevier; 2015.
87. Weleber RG, Carr RE, Murphey WH, et al. Phenotypic variation including retinitis pigmentosa, pattern dystrophy, and fundus flavimaculatus in a single family with a deletion of codon 153 or 154 of the peripherin/RDS gene. *Arch Ophthalmol* 1993;111:1531-42.
88. Lyon MF. The Lyon and the LINE hypothesis. *Semin Cell Dev Biol* 2003;14:313-18.
89. Pettersson E, Lundeberg J, Ahmadian A. Generations of sequencing technologies. *Genomics* 2009;93:105-11.
90. Arnold JJ, Sarks JP, Killingsworth MC, et al. Adult vitelliform macular degeneration: a clinicopathological study. *Eye (Lond)* 2003;17(6):717-26.
91. Shah KH, Levinson RD, Yu F, et al. Birdshot chorioretinopathy. *Surv Ophthalmol* 2005;50(6):519-41.
92. Priem HA, Kijlstra A, Noens L, et al. HLA typing in birdshot chorioretinopathy. *Am J Ophthalmol* 1988;105(2):182-5.
93. Brucker AJ, Deglin EA, Bene C, et al. Subretinal choroidal neovascularization in birdshot retinochoroidopathy. *Am J Ophthalmol* 1985;99(1):40-4.
94. Miller SA, Bresnick GH, Chandra SR. Choroidal neovascular membrane in Best's vitelliform macular dystrophy. *Am J Ophthalmol* 1976;82(2):252-5.
95. Miller SA. Multifocal Best's vitelliform dystrophy. *Arch Ophthalmol* 1977;95(6):984-90.

96. Testa F, Rossi S, Passerini I, et al. A normal electro-oculography in a family affected by best disease with a novel spontaneous mutation of the BEST1 gene. *Br J Ophthalmol* 2008;92(11):1467-70.
97. Goebel HH. The neuronal ceroid-lipofuscinoses. *Semin Pediatr Neurol* 1996;3(4):270-8.
98. Kufs H. Über einen Fall von spätester Form der amaurotischen Idiotie mit dem Beginn im 42 und Tod im 59 Lebens-jahre in klinischer, histologischer und Vererbungs-pathologischer Beziehung. *Z ges Neurol* 1931;137:432-48.
99. Weleber RG. The dystrophic retina in multisystem disorders: the electroretinogram in neuronal ceroid lipofuscinoses. *Eye (Lond)* 1998;12(Pt 3b):580-90.
100. Rowe SE, Trobe JD, Sieving PA. Idiopathic photoreceptor dysfunction causes unexplained visual acuity loss in later adulthood. *Ophthalmology* 1990;97(12):1632-7.
101. Robson AG, Michaelides M, Luong VA, et al. Functional correlates of fundus autofluorescence abnormalities in patients with RPGR or RIMS1 mutations causing cone or cone rod dystrophy. *Br J Ophthalmol* 2008;92(1):95-102.
102. Celesia GG, Kaufman D. Pattern ERGs and visual evoked potentials in maculopathies and optic nerve diseases. *Invest Ophthalmol Vis Sci* 1985;26:726-35.
103. Zeitz C, Robson AG, Audo I. Congenital stationary night blindness: an analysis and update of genotype-phenotype correlations and pathogenic mechanism. *Prog Retin Eye Res* 2015;45:58-110.
104. Bijveld MM, Florjin RJ, Bergen AA, et al. Genotype and phenotype of 101 Dutch patients with congenital stationary night blindness. *Ophthalmology* 2013;120(10):2072-81.
105. Rosenberg T, Haim M, Piczenik Y, et al. Autosomal dominant stationary night-blindness: a large family rediscovered. *Acta Ophthalmol (Copenh)* 1991;69(6):694-702.
106. Niwa Y, Kondo M, Ueno S, et al. Cone and rod dysfunction in fundus albipunctatus with RDH5 mutation: an electrophysiological study. *Invest Ophthalmol Vis Sci* 2005;46(4):1480-5.
107. Gouras P. Electroretinography: some basic principles. *Invest Ophthalmol* 1970;9(8):557-69.
108. Franceschetti A, Francois J, Babel J. Chorioretinal heredodegenerations. Springfield, IL: Charles C Thomas;1974.
109. Lambert SR, Kriss A, Taylor D, et al. Follow-up and diagnostic reappraisal of 75 patients with Leber's congenital amaurosis. *Am J Ophthalmol* 1989;107:624-31.
110. Majander A, Robson AG, Joäc C, et al. The pattern of retinal ganglion cell dysfunction in Leber hereditary optic neuropathy. *Mitochondrion* 2017;36:138-49.
111. Blair NP, Goldberg MF, Fishman GA, Salzano T. Autosomal dominant vitreoretinochoroidopathy (ADVIRC). *Br J Ophthalmol* 1984;68(1):2-9.
112. Han DP, Lewandowski MF. Electro-oculography in autosomal dominant vitreoretinochoroidopathy. *Arch Ophthalmol* 1992;110(11):1563-7.
113. Kinnick TR, Mullins RF, Dev S, et al. Autosomal recessive vitelliform macular dystrophy in a large cohort of vitelliform macular dystrophy patients. *Retina*

- 2011;31(3):581-95.
114. Hsieh RC, Fine BS, Lyons JS. Patterned dystrophies of the retinal pigment epithelium. *Arch Ophthalmol* 1977;95(3):429-35.
 115. Brinton GS, Norton EW, Zahn JR, Knighton RW. Ocular quinine toxicity. *Am J Ophthalmol* 1980;90(3):403-10.
 116. Miller FS 3rd, Bunt-Milam AH, Kaline RE. Clinical-ultrastructural study of thioridazine retinopathy. *Ophthalmology* 1982;89(12):1478-88.
 117. Kertes PJ, Lee TK, Coupland SG. The utility of multifocal electroretinography in monitoring drug toxicity: deferoxamine retinopathy. *Can J Ophthalmol* 2004;39:656-61.
 118. Tanino TO. Studies on pigmentary retinal dystrophy. II. Recordability of electroretinogram and the mode of inheritance. *Jpn J Ophthalmol* 1976;20:482-6.
 119. Berson EL, Sandberg MA, Rosner B, et al. Natural course of retinitis pigmentosa over a three-year interval. *Am J Ophthalmol* 1985;99(3):240-51.
 120. Heckenlively JR, Yoser SL, Friedman LH, Oversier JJ. Clinical findings and common symptoms in retinitis pigmentosa. *Am J Ophthalmol* 1988;105:504-11.
 121. Berson EL, Howard J. Temporal aspects of the electroretinogram in sector retinitis pigmentosa. *Arch Ophthalmol* 1971;86:653-65.
 122. Cotterman CW, Falls HF, Neel JV. Some hereditary diseases having subclinical manifestations in carriers. *Genetics* 1948;33(6):608.
 123. Franceschetti A, Francois J, Babel J. Retinitis punctata albescens. In Franceschetti A, Francois J, Babel J, editors. *Chorioretinal heredodegenerations*. Springfield, IL: Charles C Thomas;1974. p. 222-31.
 124. Scholl HP, Besch D, Vonthein R, et al. Alterations of slow and fast rod ERG signals in patients with molecularly confirmed Städardt disease type 1. *Invest Ophthalmol Vis Sci* 2002;43(4):1248-56.
 125. Sikkink SK, Biswas S, Parry NR, et al. X-linked retinoschisis: an update. *J Med Genet* 2007;44(4):225-32.