

KARACİĞER MOLEKÜLER BİYOLOJİSİ İMMÜNOLOJİSİ, HEPATİT VE HEPATOSELLÜLER KARSINOMA

Sevgi KARABULUT UZUNÇAKMAK³

GİRİŞ

Karaciğer sindirim, detoksifikasyon, sıvı elektrolit dengesi, homeostazi gibi çok sayıda metabolik sürecin bir parçasıdır. Karaciğer görevleri sentezleme, safra üretme, detoksifikasyon şeklinde sıralanabilir. Karaciğerin fonksiyonsuz kaldığı durumlarda toksin birikimi, koma ve ölüm gerçekleşir(1,2). İnsan karaciğerinden portal ven ve hepatik arter aracılığıyla her dakikada 1.5 lt kan geçmektedir. Bu kan da hepatik immün sistemin tolere ettiği diet ile gelen antijenik yük ve genel ürünler bulunmaktadır.

Karaciğer, parenkimal hücrelerden (hepatosit) ve parenkimal olmayan (endotel hücreler, Kupffer hücreleri, lenfositler, stellat hücreler) hücrelerden oluşan kendini yenileyebilme kabiliyetine sahip nadir bir organdır. Hepatositler normal koşullar altında bölünmez hücre döngüsünün G0 fazında beklerler(3). Fakat karaciğer bütünlüğünü etkileyen ve karaciğerin normal ebatlarının küçüldüğüne dair bir uyarı almaları durumunda içerisinde bulundukları G0 fazından çıkış hızla kaybolan hücrelerin yerine yenisini koymak üzere prolifere olurlar. Hepatositler, karaciğer kitlesinin %80'ını oluşturur. Çok miktarda mitokondri ve endoplazmik retikulum sahip olan bu hücreler pihtlaşma faktörleri, safra asidi, serum proteinleri gibi çok sayıda molekül üretmektedirler. Hepatositler içerdikleri detoksifikasyon enziminleri (p450) ile vücut metabolizması sonucu oluşan ya da vücuta alınan toksinlerin ayırtmasını gerçekleştirir.

Dokular homeostaziyi koruyabilmek için ya var olan hücrelerin replikasyonunu sağlarlar ya da kök hücrelerin farklılaşmasını uyarırlar. Karaciğer küçüldüğüne dair bir uyarı alındığında homeostaziyi korumak için hem parakrin hem de otokrin yolla hücre sayısını arttırır. Hepatositler fonksiyonel heterojeniteye sahiptir. Farklı bölgelerdeki hepatositler farklı alt tip belirteçler eksprese eden en az üç tipe (periportal, midlobüler ve perisantral) ayrılmaktadır. Prolifere ve kendini yenileyebilen lobun merkez venine yakın hepatositler eksprese ettikleri *Tbx3*, *Axin2* Wnt/β-katenin hedef geni, gibi genler proliferasyon kapasitesinin diğer hepatositlerden fazla olduğunu gösterir(4). Hasarlanmamış karaciğerde azalan Wnt sinyalizasyonu azalan proliferasyon ve azalan organ kitlesiyle ilişkilidir(5). Periportal hepatositler SRY-box (*Sox*)9 gibi progenitor belirteçlere sahip olmalarının yanısıra safra kanalı spesifik genlerce ve hepatosit nükleer faktör (*Hnf*)4α gibi hepatosit spesifik transkripsiyon faktörlerince zengindirler. Bu hepatositler kronik hasar durumunda karaciğer kitlesini yeniden oluştururlar(6). Karaciğer loblündeki hepatositler farklı genomik içeriğe de sahip olabilirler. Bu onların proliferasyon kapasitesini etkileyebilmektedir(4).

İmmün olgunlaşma ve farklılaşmaya bağlı olarak karaciğer bazı immünite ilişkili değişimlere de gitmektedir(7). Alınan antjenik uyarılara binaen karaciğer bünyesinde bulunan ya da oluşan sinyallere bağlı olarak karaciğere göç eden hücreler karaciğerin savunmasını sağlamaktadır.

³ Doktor Öğretim Üyesi Bayburt Üniversitesi,
skarabulut@bayburt.edu.tr

P53 yolağı, aflatoksin ilişkili HCC gelişimine birçok aşamada etki etmektedir. Aflatoksin ilişkili vakaların %50’nde p53 geni mutasyonları tespit edilmiştir. p14ARF mikrodelesyonu p53 mutant HCC vakalarında %15-20 oranında görülmüşdür(185). Artan MDM2 ekspresyonu, gankyrin aşırı ekspresyonu HCC’li hastalarda sıkılıkla görülen değişimlerdir(186).

Sonuç

Karaciğer, vücutta çok farklı görevleri yapan birden fazla fonksiyona sahip hayatı bir organdır. Homeostazi için bu kadar önemli olması onun kendini koruması ve devam ettirebilmesi için ona hem fiziki hem de hücresel düzeyde avantajlar kazandırmıştır. Barındırdığı farklı türdeki hücrelerin yanısıra aynı hücrelerin farklı fonksiyonları yerine getirip multifonksiyonlu olması da kendi devamlılığının garantisidir. Çok sayıda sahip olduğu avantajlara rağmen genetik ve çevresel etkenler karaciğerde bazı hastalıklara sebep olmaktadır. Tüm hastalıklarda olduğu gibi karaciğerde oluşan hastalıkların temelinde moleküler mekanizmalardaki değişimler yatmaktadır. Gen ekspresyon değişimleri, yapışal bozulmalar, polimorfizmler, gen insersiyon ya da delesyonları birçok hastalıkta olduğu gibi karaciğerde de görülmektedir.

KAYNAKLAR

1. Karl Mm, Howell Ra, Hutchinson Jh, Catanzaro Fj. Liver coma, with particular reference to management. AMA Arch Intern Med. 1953;91(2):159–76.
2. Tan AKY, Loh KM, Ang LT. Evaluating the regenerative potential and functionality of human liver cells in mice. Differentiation. 98:25–34.
3. Michalopoulos GK, DeFrances MC. Liver regeneration. Science. 1997;276(5309):60–6.
4. Wang B, Zhao L, Fish M, Logan CY, Nusse R. Self-renewing diploid Axin2(+) cells fuel homeostatic renewal of the liver. Nature. 2015;524(7564):180–5.
5. Planas-Paz L, Orsini V, Boulter L, Calabrese D, Pikiolek M, Nigsch F, et al. The RSPO-LGR4/5-ZNRF3/RNF43 module controls liver zonation and size. Nat Cell Biol. 2016;18(5):467–79.
6. Font-Burgada J, Shalapour S, Ramaswamy S, Hsueh B, Rossell D, Umemura A, et al. Hybrid Periportal Hepatocytes Regenerate the Injured Liver without Giving Rise to Cancer. Cell. 2015;162(4):766–79.
7. Abo T. Extrathymic pathways of T-cell differentiation and immunomodulation. Int Immunopharmacol. 2001;1(7):1261–73.
8. Hu S-J, Jiang S-S, Zhang J, Luo D, Yu B, Yang L-Y, et al. Effects of apoptosis on liver aging. World J Clin Cases. 2019;7(6):691–704.
9. Doherty DG. Immunity, tolerance and autoimmunity in the liver: A comprehensive review. J Autoimmun. 2016;66:60–75.
10. Robinson MW, Harmon C, O’Farrelly C. Liver immunology and its role in inflammation and homeostasis. Cell Mol Immunol. 2016;13(3):267–76.
11. Jenne CN, Kubes P. Immune surveillance by the liver. Nat Immunol. 2013;14(10):996–1006.
12. Wisse E, Braet F, Luo D, De Zanger R, Jans D, Crabbé E, et al. Structure and function of sinusoidal lining cells in the liver. Toxicol Pathol. 24(1):100–11.
13. Robinson MW, Harmon C, O’Farrelly C. Liver immunology and its role in inflammation and homeostasis. Cell Mol Immunol. 2016;13(3):267–76.
14. Kelly A, Fahey R, Fletcher JM, Keogh C, Carroll AG, Siddachari R, et al. CD141⁺ myeloid dendritic cells are enriched in healthy human liver. J Hepatol. 2014;60(1):135–42.
15. Doherty DG, Norris S, Madrigal-Estebas L, McEntee G, Traynor O, Hegarty JE, et al. The human liver contains multiple populations of NK cells, T cells, and CD3+CD56+ natural T cells with distinct cytotoxic activities and Th1, Th2, and Th0 cytokine secretion patterns. J Immunol. 1999;163(4):2314–21.
16. Norris S, Collins C, Doherty DG, Smith F, McEntee G, Traynor O, et al. Resident human hepatic lymphocytes are phenotypically different from circulating lymphocytes. J Hepatol. 1998;28(1):84–90.
17. Wu J, Meng Z, Jiang M, Zhang E, Trippler M, Broering R, et al. Toll-like receptor-induced innate immune responses in non-parenchymal liver cells are cell type-specific. Immunology. 2010;129(3):363–74.
18. Knolle PA, Limmer A. Control of immune responses by savenger liver endothelial cells. Swiss Med Wkly. 2003;133(37–38):501–6.
19. von Oppen N, Schurich A, Hegenbarth S, Stabenow D, Tolba R, Weiskirchen R, et al. Systemic antigen cross-presented by liver sinusoidal endothelial cells induces liver-specific CD8 T-cell retention and tolerization. Hepatology. 2009;49(5):1664–72.
20. Schurich A, Berg M, Stabenow D, Böttcher J, Kern M, Schild H-J, et al. Dynamic regulation of CD8 T cell tolerance induction by liver sinusoidal endothelial cells. J Immunol. 2010;184(8):4107–14.
21. Diehl L, Schurich A, Grochtmann R, Hegenbarth S, Chen L, Knolle PA. Tolerogenic maturation of liver sinusoidal endothelial cells promotes B7-homolog 1-dependent CD8+ T cell tolerance. Hepatology. 2008;47(1):296–305.
22. Knolle PA, Schmitt E, Jin S, Germann T, Duchmann R, Hegenbarth S, et al. Induction of cytokine production in naive CD4(+) T cells by antigen-presenting murine

- liver sinusoidal endothelial cells but failure to induce differentiation toward Th1 cells. *Gastroenterology*. 1999;116(6):1428–40.
23. Klugewitz K, Blumenthal-Barby F, Schrage A, Knolle PA, Hamann A, Crispe IN. Immunomodulatory effects of the liver: deletion of activated CD4+ effector cells and suppression of IFN-gamma-producing cells after intravenous protein immunization. *J Immunol*. 2002;169(5):2407–13.
 24. Knolle PA, Germann T, Treichel U, Uhrig A, Schmitt E, Hegenbarth S, et al. Endotoxin down-regulates T cell activation by antigen-presenting liver sinusoidal endothelial cells. *J Immunol*. 1999;162(3):1401–7.
 25. Grant CR, Liberal R. Liver immunology: How to reconcile tolerance with autoimmunity. *Clin Res Hepatol Gastroenterol*. 2017;41(1):6–16.
 26. Zhang X, Meng Z, Qiu S, Xu Y, Yang D, Schlaak JE, et al. Lipopolysaccharide-induced innate immune responses in primary hepatocytes downregulates woodchuck hepatitis virus replication via interferon-independent pathways. *Cell Microbiol*. 2009;11(11):1624–37.
 27. Volanakis JE. Transcriptional regulation of complement genes. *Annu Rev Immunol*. 1995;13:277–305.
 28. Crispe IN. Hepatocytes as Immunological Agents. *J Immunol*. 2016;196(1):17–21.
 29. Roychowdhury S, McMullen MR, Pisano SG, Liu X, Nagy LE. Absence of receptor interacting protein kinase 3 prevents ethanol-induced liver injury. *Hepatology*. 2013;57(5):1773–83.
 30. Takemoto K, Hatano E, Iwaisako K, Takeiri M, Noma N, Ohmae S, et al. Necrostatin-1 protects against reactive oxygen species (ROS)-induced hepatotoxicity in acetaminophen-induced acute liver failure. *FEBS Open Bio*. 2014;4:777–87.
 31. You Q, Cheng L, Kedl RM, Ju C. Mechanism of T cell tolerance induction by murine hepatic Kupffer cells. *Hepatology [Internet]*. 2008;48(3):978–90.
 32. Seki E, Tsutsui H, Nakano H, Tsuji N, Hoshino K, Adachi O, et al. Lipopolysaccharide-induced IL-18 secretion from murine Kupffer cells independently of myeloid differentiation factor 88 that is critically involved in induction of production of IL-12 and IL-1beta. *J Immunol*. 2001;166(4):2651–7.
 33. Seki E, Tsutsui H, Tsuji NM, Hayashi N, Adachi K, Nakano H, et al. Critical roles of myeloid differentiation factor 88-dependent proinflammatory cytokine release in early phase clearance of *Listeria monocytogenes* in mice. *J Immunol*. 2002;169(7):3863–8.
 34. Kopydlowski KM, Salkowski CA, Cody MJ, van Rooijen N, Major J, Hamilton TA, et al. Regulation of macrophage chemokine expression by lipopolysaccharide in vitro and in vivo. *J Immunol*. 1999;163(3):1537–44.
 35. Schwabe RF, Seki E, Brenner DA. Toll-like receptor signaling in the liver. *Gastroenterology*. 2006;130(6):1886–900.
 36. Weiskirchen R, Tacke F. Cellular and molecular functions of hepatic stellate cells in inflammatory responses and liver immunology. *Hepatobiliary Surg Nutr*. 2014;3(6):344–63.
 37. Bomble M, Tacke F, Rink L, Kovalenko E, Weiskirchen R. Analysis of antigen-presenting functionality of cultured rat hepatic stellate cells and transdifferentiated myofibroblasts. *Biochem Biophys Res Commun*. 2010;396(2):342–7.
 38. Winau F, Hegasy G, Weiskirchen R, Weber S, Cassan C, Sieling PA, et al. Ito cells are liver-resident antigen-presenting cells for activating T cell responses. *Immunity*. 2007;26(1):117–29.
 39. Viñas O, Bataller R, Sancho-Bru P, Ginès P, Berenguer C, Enrich C, et al. Human hepatic stellate cells show features of antigen-presenting cells and stimulate lymphocyte proliferation. *Hepatology*. 2003;38(4):919–29.
 40. Gressner AM, Weiskirchen R. Modern pathogenetic concepts of liver fibrosis suggest stellate cells and TGF-beta as major players and therapeutic targets. *J Cell Mol Med*. 2010;14(1):76–99.
 41. Tacke F, Weiskirchen R. Update on hepatic stellate cells: pathogenic role in liver fibrosis and novel isolation techniques. *Expert Rev Gastroenterol Hepatol*. 2012;6(1):67–80.
 42. Wasmuth HE, Weiskirchen R. [Pathogenesis of liver fibrosis: modulation of stellate cells by chemokines]. *Z Gastroenterol*. 2010;48(1):38–45.
 43. David BA, Rezende RM, Antunes MM, Santos MM, Freitas Lopes MA, Diniz AB, et al. Combination of Mass Cytometry and Imaging Analysis Reveals Origin, Location, and Functional Repopulation of Liver Myeloid Cells in Mice. *Gastroenterology*. 2016;151(6):1176–91.
 44. Kubes P, Mehal WZ. Sterile inflammation in the liver. *Gastroenterology*. 2012;143(5):1158–72.
 45. Mafra K, Nakagaki BN, Castro Oliveira HM, Rezende RM, Antunes MM, Menezes GB. The liver as a nursery for leukocytes. *J Leukoc Biol*. 2019.
 46. Freitas-Lopes MA, Mafra K, David BA, Carvalho-Gontijo R, Menezes GB. Differential Location and Distribution of Hepatic Immune Cells. *Cells*. 2017;6(4).
 47. Shang N, Figini M, Shangguan J, Wang B, Sun C, Pan L, et al. Dendritic cells based immunotherapy. *Am J Cancer Res*. 2017;7(10):2091–102.
 48. Pierre P, Turley SJ, Gatti E, Hull M, Meltzer J, Mirza A, et al. Developmental regulation of MHC class II transport in mouse dendritic cells. *Nature*. 1997;388(6644):787–92.
 49. Kahraman A, Barreyro FJ, Bronk SF, Werneburg NW, Mott JL, Akazawa Y, et al. TRAIL mediates liver injury by the innate immune system in the bile duct-ligated mouse. *Hepatology*. 2008;47(4):1317–30.
 50. Radaeva S, Sun R, Jaruga B, Nguyen VT, Tian Z, Gao B. Natural killer cells ameliorate liver fibrosis by killing activated stellate cells in NKG2D-dependent and tumor necrosis factor-related apoptosis-inducing ligand-dependent manners. *Gastroenterology*. 2006;130(2):435–52.
 51. Huber S, Shi C, Budd RC. Gammadelta T cells promote a Th1 response during coxsackievirus B3 infection in vivo: role of Fas and Fas ligand. *J Virol*. 2002;76(13):6487–94.

52. Farouk SE, Mincheva-Nilsson L, Krensky AM, Dieli F, Troye-Blomberg M. Gamma delta T cells inhibit in vitro growth of the asexual blood stages of Plasmodium falciparum by a granule exocytosis-dependent cytotoxic pathway that requires granulysin. *Eur J Immunol.* 2004;34(8):2248–56.
53. Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol.* 2010;10(7):467–78.
54. Hammerich L, Tacke F. Role of gamma-delta T cells in liver inflammation and fibrosis. *World J Gastrointest Pathophysiol.* 2014;5(2):107–13.
55. Murakami J, Shimizu Y, Kashii Y, Kato T, Minemura M, Okada K, et al. Functional B-cell response in intrahepatic lymphoid follicles in chronic hepatitis C. *Hepatology.* 1999;30(1):143–50.
56. Zhang H, Stoltz DB, Chalasani G, Thomson AW. Hepatic B cells are readily activated by Toll-like receptor-4 ligation and secrete less interleukin-10 than lymphoid tissue B cells. *Clin Exp Immunol.* 2013;173(3):473–9.
57. Coban Z, Barton MC. Integrative genomics: liver regeneration and hepatocellular carcinoma. *J Cell Biochem.* 2012;113(7):2179–84.
58. Michalopoulos GK. Hepatostat: Liver regeneration and normal liver tissue maintenance. *Hepatology.* 2017;65(4):1384–92.
59. Pahlavan PS, Feldmann RE, Zavos C, Kountouras J. Prometheus' challenge: molecular, cellular and systemic aspects of liver regeneration. *J Surg Res.* 2006;134(2):238–51.
60. Mohn KL, Laz TM, Hsu JC, Melby AE, Bravo R, Taub R. The immediate-early growth response in regenerating liver and insulin-stimulated H-35 cells: comparison with serum-stimulated 3T3 cells and identification of 41 novel immediate-early genes. *Mol Cell Biol.* 1991;11(1):381–90.
61. Michalopoulos GK. Principles of liver regeneration and growth homeostasis. *Compr Physiol.* 2013;3(1):485–513.
62. Michalopoulos GK. Liver regeneration. *J Cell Physiol.* 2007;213(2):286–300.
63. Taub R. Liver regeneration 4: transcriptional control of liver regeneration. *FASEB J.* 1996;10(4):413–27.
64. Mao SA, Glorioso JM, Nyberg SL. Liver regeneration. *Transl Res.* 2014;163(4):352–62.
65. Hou JZ, Kan MK, McKeehan K, McBride G, Adams P, McKeehan WL. Fibroblast growth factor receptors from liver vary in three structural domains. *Science.* 1991;251(4994):665–8.
66. Padriassa-Altés S, Bachofner M, Bogorad RL, Pohlmeier L, Rossolini T, Böhm F, et al. Control of hepatocyte proliferation and survival by Fgf receptors is essential for liver regeneration in mice. *Gut.* 2015;64(9):1444–53.
67. Block GD, Locker J, Bowen WC, Petersen BE, Katyal S, Strom SC, et al. Population expansion, clonal growth, and specific differentiation patterns in primary cultures of hepatocytes induced by HGF/SF, EGF and TGF alpha in a chemically defined (HGM) medium. *J Cell Biol.* 1996;132(6):1133–49.
68. Tao Y, Wang M, Chen E, Tang H. Liver Regeneration: Analysis of the Main Relevant Signaling Molecules. *Mediators Inflamm.* 2017;2017:4256352.
69. Ding B-S, Nolan DJ, Butler JM, James D, Babazadeh AO, Rosenwaks Z, et al. Inductive angiocrine signals from sinusoidal endothelium are required for liver regeneration. *Nature.* 2010;468(7321):310–5.
70. Monga SPS. Role of Wnt/β-catenin signaling in liver metabolism and cancer. *Int J Biochem Cell Biol.* 2011;43(7):1021–9.
71. Tan CY, Lai RC, Wong W, Dan YY, Lim S-K, Ho HK. Mesenchymal stem cell-derived exosomes promote hepatic regeneration in drug-induced liver injury models. *Stem Cell Res Ther.* 2014;5(3):76.
72. Salehi S, Brereton HC, Arno MJ, Darling D, Quaglia A, O'Grady J, et al. Human liver regeneration is characterized by the coordinated expression of distinct microRNA governing cell cycle fate. *Am J Transplant.* 2013;13(5):1282–95.
73. Apte U, Gkretsi V, Bowen WC, Mars WM, Luo J-H, Donthamsetty S, et al. Enhanced liver regeneration following changes induced by hepatocyte-specific genetic ablation of integrin-linked kinase. *Hepatology.* 2009;50(3):844–51.
74. Campbell JS, Prichard L, Schaper F, Schmitz J, Stephen-Son-Famy A, Rosenfeld ME, et al. Expression of suppressors of cytokine signaling during liver regeneration. *J Clin Invest.* 2001;107(10):1285–92.
75. Deryck R, Zhang YE. Smad-dependent and Smad-independent pathways in TGF-beta family signalling. *Nature.* 2003;425(6958):577–84.
76. Michalopoulos GK, Bowen WC, Mulè K, Stoltz DB. Histological organization in hepatocyte organoid cultures. *Am J Pathol.* 2001;159(5):1877–87.
77. Chari RS, Price DT, Sue SR, Meyers WC, Jirtle RL. Down-regulation of transforming growth factor beta receptor type I, II, and III during liver regeneration. *Am J Surg.* 1995;169(1):126–31; discussion 131–2.
78. Wuestefeld T, Pesic M, Rudalska R, Dauch D, Longrich T, Kang T-W, et al. A Direct *in vivo* RNAi screen identifies MKK4 as a key regulator of liver regeneration. *Cell.* 2013;153(2):389–401.
79. Yilmalai D, Christodoulou C, Galli GG, Yanger K, Pepe-Mooney B, Gurung B, et al. Hippo pathway activity influences liver cell fate. *Cell.* 2014;157(6):1324–38.
80. Zhao B, Tumaneng K, Guan K-L. The Hippo pathway in organ size control, tissue regeneration and stem cell self-renewal. *Nat Cell Biol.* 2011;13(8):877–83.
81. Wang C, Zhang L, He Q, Feng X, Zhu J, Xu Z, et al. Differences in Yes-associated protein and mRNA levels in regenerating liver and hepatocellular carcinoma. *Mol Med Rep.* 2012;5(2):410–4.
82. Bataller R, Brenner DA. Liver fibrosis. *J Clin Invest.* 2005;115(2):209–18.

83. Omenetti A, Porrello A, Jung Y, Yang L, Popov Y, Choi SS, et al. Hedgehog signaling regulates epithelial-mesenchymal transition during biliary fibrosis in rodents and humans. *J Clin Invest.* 2008;118(10):3331–42.
84. Schuppan D. Liver fibrosis: Common mechanisms and antifibrotic therapies. *Clin Res Hepatol Gastroenterol.* 2015;39 Suppl 1:S51–9.
85. de Oliveira da Silva B, Ramos LF, Moraes KCM. Molecular interplays in hepatic stellate cells: apoptosis, senescence, and phenotype reversion as cellular connections that modulate liver fibrosis. *Cell Biol Int.* 2017;41(9):946–59.
86. Milani S, Herbst H, Schuppan D, Stein H, Surrenti C. Transforming growth factors beta 1 and beta 2 are differentially expressed in fibrotic liver disease. *Am J Pathol.* 1991;139(6):1221–9.
87. Pinzani M, Milani S, Herbst H, DeFranco R, Grappone C, Gentilini A, et al. Expression of platelet-derived growth factor and its receptors in normal human liver and during active hepatic fibrogenesis. *Am J Pathol.* 1996;148(3):785–800.
88. Omenetti A, Porrello A, Jung Y, Yang L, Popov Y, Choi SS, et al. Hedgehog signaling regulates epithelial-mesenchymal transition during biliary fibrosis in rodents and humans. *J Clin Invest.* 2008;118(10):3331–42.
89. Meyer DH, Bachem MG, Gressner AM. Modulation of hepatic lipocyte proteoglycan synthesis and proliferation by Kupffer cell-derived transforming growth factors type beta 1 and type alpha. *Biochem Biophys Res Commun.* 1990;171(3):1122–9.
90. Win KM, Charlotte F, Mallat A, Cherqui D, Martin N, Mavier P, et al. Mitogenic effect of transforming growth factor-beta 1 on human Ito cells in culture: evidence for mediation by endogenous platelet-derived growth factor. *Hepatology.* 1993;18(1):137–45.
91. Schuppan D, Ruehl M, Somasundaram R, Hahn EG. Matrix as a modulator of hepatic fibrogenesis. *Semin Liver Dis.* 2001;21(3):351–72.
92. Brown B, Lindberg K, Reing J, Stoltz DB, Badylak SF. The basement membrane component of biologic scaffolds derived from extracellular matrix. *Tissue Eng.* 2006;12(3):519–26.
93. Breitkopf K, Godoy P, Ciucan L, Singer M V, Dooley S. TGF-beta/Smad signaling in the injured liver. *Z Gastroenterol.* 2006;44(1):57–66.
94. Sahin H, Trautwein C, Wasmuth HE. Functional role of chemokines in liver disease models. *Nat Rev Gastroenterol Hepatol.* 2010;7(12):682–90.
95. Schwabe RF, Bataller R, Brenner DA. Human hepatic stellate cells express CCR5 and RANTES to induce proliferation and migration. *Am J Physiol Gastrointest Liver Physiol.* 2003;285(5):G949–58.
96. Lee UE, Friedman SL. Mechanisms of hepatic fibrogenesis. *Best Pract Res Clin Gastroenterol.* 2011;25(2):195–206.
97. Kubes P, Mehal WZ. Sterile inflammation in the liver. *Gastroenterology.* 2012;143(5):1158–72.
98. Duffield JS, Forbes SJ, Constandinou CM, Clay S, Partolina M, Vuthoori S, et al. Selective depletion of macrophages reveals distinct, opposing roles during liver injury and repair. *J Clin Invest.* 2005;115(1):56–65.
99. Tacke F, Zimmermann HW. Macrophage heterogeneity in liver injury and fibrosis. *J Hepatol.* 2014;60(5):1090–6.
100. Pellicoro A, Ramachandran P, Iredale JP, Fallowfield JA. Liver fibrosis and repair: immune regulation of wound healing in a solid organ. *Nat Rev Immunol.* 2014;14(3):181–94.
101. Karlmark KR, Weiskirchen R, Zimmermann HW, Gassler N, Ginhoux F, Weber C, et al. Hepatic recruitment of the inflammatory Gr1+ monocyte subset upon liver injury promotes hepatic fibrosis. *Hepatology.* 2009;50(1):261–74.
102. Ramachandran P, Pellicoro A, Vernon MA, Boulter L, Aucott RL, Ali A, et al. Differential Ly-6C expression identifies the recruited macrophage phenotype, which orchestrates the regression of murine liver fibrosis. *Proc Natl Acad Sci U S A.* 2012;109(46):E3186–95.
103. Tacke F, Zimmermann HW. Macrophage heterogeneity in liver injury and fibrosis. *J Hepatol.* 2014;60(5):1090–6.
104. Schuppan D. Liver fibrosis: Common mechanisms and antifibrotic therapies. Vol. 39, *Clinics and Research in Hepatology and Gastroenterology.* 2015. p. S51–9.
105. Ikejima K, Okumura K, Kon K, Takei Y, Sato N. Role of adipocytokines in hepatic fibrogenesis. *J Gastroenterol Hepatol.* 2007;22 Suppl 1:S87–92.
106. Yamaguchi K, Yang L, McCall S, Huang J, Yu XX, Pandey SK, et al. Inhibiting triglyceride synthesis improves hepatic steatosis but exacerbates liver damage and fibrosis in obese mice with nonalcoholic steatohepatitis. *Hepatology.* 2007;45(6):1366–74.
107. Jeong W, Osei-Hyiaman D, Park O, Liu J, Bátkai S, Mukhopadhyay P, et al. Paracrine activation of hepatic CB1 receptors by stellate cell-derived endocannabinoids mediates alcoholic fatty liver. *Cell Metab.* 2008;7(3):227–35.
108. Zvibel I, Atias D, Phillips A, Halpern Z, Oren R. Thyroid hormones induce activation of rat hepatic stellate cells through increased expression of p75 neurotrophin receptor and direct activation of Rho. *Lab Invest.* 2010;90(5):674–84.
109. Ruddell RG, Oakley F, Hussain Z, Yeung I, Bryan-Lluka LJ, Ramm GA, et al. A role for serotonin (5-HT) in hepatic stellate cell function and liver fibrosis. *Am J Pathol.* 2006;169(3):861–76.
110. Guo J, Loke J, Zheng F, Hong F, Yea S, Fukata M, et al. Functional linkage of cirrhosis-predictive single nucleotide polymorphisms of Toll-like receptor 4 to hepatic stellate cell responses. *Hepatology.* 2009;49(3):960–8.
111. Pradere J-P, Troeger JS, Dapito DH, Mencin AA, Schwabe RF. Toll-like receptor 4 and hepatic fibrogenesis. *Semin Liver Dis.* 2010;30(3):232–44.

112. Christen U, Hintermann E. Pathogens and autoimmune hepatitis. *Clin Exp Immunol.* 2019;195(1):35–51.
113. Mieli-Vergani G, Vergani D, Czaja AJ, Manns MP, Krawitt EL, Vierling JM, et al. Autoimmune hepatitis. *Nat Rev Dis Prim.* 2018;4:18017.
114. Floreani A, Restrepo-Jiménez P, Secchi MF, De Martin S, Leung PSC, Krawitt E, et al. Etiopathogenesis of autoimmune hepatitis. *J Autoimmun.* 2018;95:133–43.
115. Arndtz K, Hirschfield GM. The Pathogenesis of Autoimmune Liver Disease. *Dig Dis.* 34(4):327–33.
116. Liberal R, Vergani D, Mieli-Vergani G. Update on Autoimmune Hepatitis. *J Clin Transl Hepatol.* 2015;3(1):42–52.
117. Moy L, Levine J. Autoimmune hepatitis: a classic autoimmune liver disease. *Curr Probl Pediatr Adolesc Health Care.* 2014;44(11):341–6.
118. Gatselis NK, Zachou K, Koukoulis GK, Dalekos GN. Autoimmune hepatitis, one disease with many faces: etiopathogenetic, clinico-laboratory and histological characteristics. *World J Gastroenterol.* 2015;21(1):60–83.
119. Liberal R, Grant CR, Holder BS, Cardone J, Martínez-Llordella M, Ma Y, et al. In autoimmune hepatitis type 1 or the autoimmune hepatitis-sclerosing cholangitis variant defective regulatory T-cell responsiveness to IL-2 results in low IL-10 production and impaired suppression. *Hepatology.* 2015;62(3):863–75.
120. Liberal R, Longhi MS, Mieli-Vergani G, Vergani D. Pathogenesis of autoimmune hepatitis. *Best Pract Res Clin Gastroenterol.* 2011;25(6):653–64.
121. Seldin MF. The genetics of human autoimmune disease: A perspective on progress in the field and future directions. *J Autoimmun.* 2015;64:1–12.
122. Bittencourt PL, Goldberg AC, Cançado EL, Porta G, Carrilho FJ, Farias AQ, et al. Genetic heterogeneity in susceptibility to autoimmune hepatitis types 1 and 2. *Am J Gastroenterol.* 1999;94(7):1906–13.
123. de Boer YS, van Gerven NMF, Zwiers A, Verwer BJ, van Hoek B, van Erpecum KJ, et al. Genome-wide association study identifies variants associated with autoimmune hepatitis type 1. *Gastroenterology.* 2014;147(2):443–52.e5.
124. Bittencourt PL, Goldberg AC, Cançado EL, Porta G, Carrilho FJ, Farias AQ, et al. Genetic heterogeneity in susceptibility to autoimmune hepatitis types 1 and 2. *Am J Gastroenterol.* 1999;94(7):1906–13.
125. Stretell MD, Donaldson PT, Thomson LJ, Santrach PJ, Moore SB, Czaja AJ, et al. Allelic basis for HLA-encoded susceptibility to type 1 autoimmune hepatitis. *Gastroenterology.* 1997;112(6):2028–35.
126. Oka S, Furukawa H, Yasunami M, Kawasaki A, Nakamura H, Nakamura M, et al. HLA-DRB1 and DQB1 alleles in Japanese type 1 autoimmune hepatitis: The predisposing role of the DR4/DR8 heterozygous genotype. *PLoS One.* 2017;12(10):e0187325.
127. Xu E, Cao H, Lin L, Liu H. rs10499194 polymorphism in the tumor necrosis factor- α inducible protein 3 (TNFAIP3) gene is associated with type-1 autoimmune hepatitis risk in Chinese Han population. *PLoS One.* 2017;12(4):e0176471.
128. Liu M, Shah VH. New Prospects for Medical Management of Acute Alcoholic Hepatitis. *Clin liver Dis.* 2019;13(5):131–5.
129. Shipley LC, Kodali S, Singal AK. Recent updates on alcoholic hepatitis. *Dig Liver Dis.* 2019;51(6):761–8.
130. Keshavarzian A, Farhadi A, Forsyth CB, Rangan J, Jakate S, Shaikh M, et al. Evidence that chronic alcohol exposure promotes intestinal oxidative stress, intestinal hyperpermeability and endotoxemia prior to development of alcoholic steatohepatitis in rats. *J Hepatol.* 2009;50(3):538–47.
131. Mutlu EA, Gillevet PM, Rangwala H, Sikaroodi M, Naqvi A, Engen PA, et al. Colonic microbiome is altered in alcoholism. *Am J Physiol Gastrointest Liver Physiol.* 2012;302(9):G966–78.
132. Singal AK, Louvet A, Shah VH, Kamath PS. Grand Rounds: Alcoholic Hepatitis. *J Hepatol.* 2018;69(2):534–43.
133. Taïeb J, Delarche C, Paradis V, Mathurin P, Grenier A, Crestani B, et al. Polymorphonuclear neutrophils are a source of hepatocyte growth factor in patients with severe alcoholic hepatitis. *J Hepatol.* 2002;36(3):342–8.
134. Matsushita H, Takaki A. Alcohol and hepatocellular carcinoma. *BMJ open Gastroenterol.* 2019;6(1):e000260.
135. Machida K, Cheng KTH, Sung VM-H, Levine AM, Foung S, Lai MMC. Hepatitis C virus induces toll-like receptor 4 expression, leading to enhanced production of beta interferon and interleukin-6. *J Virol.* 2006;80(2):866–74.
136. Sookoian S, Flichman D, Scian R, Rohr C, Dopazo H, Gianotti TF, et al. Mitochondrial genome architecture in non-alcoholic fatty liver disease. *J Pathol.* 2016;240(4):437–49.
137. Clarke JD, Novak P, Lake AD, Shipkova P, Aranibar N, Robertson D, et al. Characterization of hepatocellular carcinoma related genes and metabolites in human nonalcoholic fatty liver disease. *Dig Dis Sci.* 2014;59(2):365–74.
138. Arendt BM, Comelli EM, Ma DWL, Lou W, Teterina A, Kim T, et al. Altered hepatic gene expression in nonalcoholic fatty liver disease is associated with lower hepatic n-3 and n-6 polyunsaturated fatty acids. *Hepatology.* 2015;61(5):1565–78.
139. Katrinli S, Ozdil K, Dinler-Doganay G, Doganay L. Non-alcoholic Fatty Liver Disease: What We Learn from Omics Studies. In: Non-Alcoholic Fatty Liver Disease - Molecular Bases, Prevention and Treatment. InTech; 2018.
140. Brown GT, Kleiner DE. Histopathology of nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. *Metabolism.* 2016;65(8):1080–6.
141. Karagozian R, Derdák Z, Baffy G. Obesity-associated mechanisms of hepatocarcinogenesis. *Metabolism.* 2014;63(5):607–17.

142. McPherson S, Hardy T, Henderson E, Burt AD, Day CP, Anstee QM. Evidence of NAFLD progression from steatosis to fibrosing-steatohepatitis using paired biopsies: implications for prognosis and clinical management. *J Hepatol.* 2015;62(5):1148–55.
143. Sookoian S, Rosselli MS, Gemma C, Burgueño AL, Fernández Gianotti T, Castaño GO, et al. Epigenetic regulation of insulin resistance in nonalcoholic fatty liver disease: impact of liver methylation of the peroxisome proliferator-activated receptor γ coactivator 1 α promoter. *Hepatology.* 2010;52(6):1992–2000.
144. Sookoian S, Pirola CJ. Genetics of Nonalcoholic Fatty Liver Disease: From Pathogenesis to Therapeutics. *Semin Liver Dis.* 2019;39(2):124–40.
145. Lefebvre P, Laloyer F, Baugé E, Pawlak M, Gheeraert C, Dehondt H, et al. Interspecies NASH disease activity whole-genome profiling identifies a fibrogenic role of PPAR α -regulated dermatopontin. *JCI insight.* 2017;2(13).
146. Cermelli S, Ruggieri A, Marrero JA, Ioannou GN, Bettella L. Circulating microRNAs in patients with chronic hepatitis C and non-alcoholic fatty liver disease. *PLoS One.* 2011;6(8):e23937.
147. Pirola CJ, Fernández Gianotti T, Castaño GO, Mallardi P, San Martino J, Mora Gonzalez Lopez Ledesma M, et al. Circulating microRNA signature in non-alcoholic fatty liver disease: from serum non-coding RNAs to liver histology and disease pathogenesis. *Gut.* 2015;64(5):800–12.
148. Stapleton JT, Foung S, Muerhoff AS, Bukh J, Simmonds P. The GB viruses: a review and proposed classification of GBV-A, GBV-C (HGV), and GBV-D in genus Pegivirus within the family Flaviviridae. *J Gen Virol.* 2011;92(Pt 2):233–46.
149. Zeisel MB, Lucifora J, Mason WS, Sureau C, Beck J, Levrero M, et al. Towards an HBV cure: state-of-the-art and unresolved questions--report of the ANRS workshop on HBV cure. *Gut.* 2015;64(8):1314–26.
150. Schulze A, Gripon P, Urban S. Hepatitis B virus infection initiates with a large surface protein-dependent binding to heparan sulfate proteoglycans. *Hepatology.* 2007;46(6):1759–68.
151. Yan H, Zhong G, Xu G, He W, Jing Z, Gao Z, et al. Sodium taurocholate cotransporting polypeptide is a functional receptor for human hepatitis B and D virus. *Elife.* 2012;3.
152. Ferrari C, Missale G, Boni C, Urbani S. Immunopathogenesis of hepatitis B. *J Hepatol.* 2003;39:36–42.
153. Hayes CN, Zhang P, Zhang Y, Chayama K. Molecular Mechanisms of Hepatocarcinogenesis Following Sustained Virological Response in Patients with Chronic Hepatitis C Virus Infection. *Viruses.* 2018;10(10).
154. Westbrook RH, Dusheiko G. Natural history of hepatitis C. *J Hepatol.* 2014;61(1 Suppl):S58–68.
155. Mentha N, Clément S, Negro F, Alfaiate D. A review on hepatitis D: From virology to new therapies. *J Adv Res.* 2019;17:3–15.
156. Ahmad I, Holla RP, Jameel S. Molecular virology of hepatitis E virus. *Virus Res.* 2011;161(1):47–58.
157. Kamar N, Marion O, Abravanel F, Izopet J, Dalton HR. Extrahepatic manifestations of hepatitis E virus. *Liver Int.* 2016;36(4):467–72.
158. Feng Z, Lemon SM. Peek-a-boo: membrane hijacking and the pathogenesis of viral hepatitis. *Trends Microbiol.* 2014;22(2):59–64.
159. Takeda H, Takai A, Inuzuka T, Marusawa H. Genetic basis of hepatitis virus-associated hepatocellular carcinoma: linkage between infection, inflammation, and tumorigenesis. *J Gastroenterol.* 2017;52(1):26–38.
160. Chiba T, Marusawa H, Ushijima T. Inflammation-associated cancer development in digestive organs: mechanisms and roles for genetic and epigenetic modulation. *Gastroenterology.* 2012;143(3):550–63.
161. Fujimoto A, Furuta M, Totoki Y, Tsunoda T, Kato M, Shiraishi Y, et al. Whole-genome mutational landscape and characterization of noncoding and structural mutations in liver cancer. *Nat Genet.* 2016;48(5):500–9.
162. Schulze K, Imbeaud S, Letouzé E, Alexandrov LB, Calderaro J, Reboulissou S, et al. Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. *Nat Genet.* 2015;47(5):505–11.
163. Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA, Kinzler KW. Cancer genome landscapes. *Science.* 2013;339(6127):1546–58.
164. Totoki Y, Tatsuno K, Covington KR, Ueda H, Creighton CJ, Kato M, et al. Trans-ancestry mutational landscape of hepatocellular carcinoma genomes. *Nat Genet.* 2014;46(12):1267–73.
165. Nault JC, Mallet M, Pilati C, Calderaro J, Bioulac-Sage P, Laurent C, et al. High frequency of telomerase reverse-transcriptase promoter somatic mutations in hepatocellular carcinoma and preneoplastic lesions. *Nat Commun.* 2013;4:2218.
166. Shibata T, Aburatani H. Exploration of liver cancer genomes. *Nat Rev Gastroenterol Hepatol.* 2014;11(6):340–9.
167. Kawai-Kitahata F, Asahina Y, Tanaka S, Kakinuma S, Murakawa M, Nitta S, et al. Comprehensive analyses of mutations and hepatitis B virus integration in hepatocellular carcinoma with clinicopathological features. *J Gastroenterol.* 2016;51(5):473–86.
168. Totoki Y, Tatsuno K, Yamamoto S, Arai Y, Hosoda F, Ishikawa S, et al. High-resolution characterization of a hepatocellular carcinoma genome. *Nat Genet.* 2011;43(5):464–9.
169. Lau C-C, Sun T, Ching AKK, He M, Li J-W, Wong AM, et al. Viral-human chimeric transcript predisposes risk to liver cancer development and progression. *Cancer Cell.* 2014;25(3):335–49.
170. Sung W-K, Zheng H, Li S, Chen R, Liu X, Li Y, et al. Genome-wide survey of recurrent HBV integration in hepatocellular carcinoma. *Nat Genet.* 2012;44(7):765–9.

171. Niu B, Hann H-W. Hepatitis B Virus–Related Hepatocellular Carcinoma: Carcinogenesis, Prevention, and Treatment. In: *Updates in Liver Cancer*. InTech; 2017.
172. Kuo T-C, Chao CC-K. Hepatitis B virus X protein prevents apoptosis of hepatocellular carcinoma cells by upregulating SATB1 and HURP expression. *Biochem Pharmacol*. 2010;80(7):1093–102.
173. Said A, Ghufran A. Epidemic of non-alcoholic fatty liver disease and hepatocellular carcinoma. *World J Clin Oncol*. 2017;8(6):429–36.
174. Kuthu O, Kaleli HN, Ozer E. Molecular Pathogenesis of Nonalcoholic Steatohepatitis- (NASH-) Related Hepatocellular Carcinoma. *Can J Gastroenterol Hepatol*. 2018;2018:8543763.
175. Sookoian S, Pirola CJ. Meta-analysis of the influence of I148M variant of patatin-like phospholipase domain containing 3 gene (PNPLA3) on the susceptibility and histological severity of nonalcoholic fatty liver disease. *Hepatology*. 2011;53(6):1883–94.
176. Dongiovanni P, Donati B, Fares R, Lombardi R, Mancina RM, Romeo S, et al. PNPLA3 I148M polymorphism and progressive liver disease. *World J Gastroenterol*. 2013;19(41):6969–78.
177. Mondul A, Mancina RM, Merlo A, Dongiovanni P, Rametta R, Montalcini T, et al. PNPLA3 I148M Variant Influences Circulating Retinol in Adults with Nonalcoholic Fatty Liver Disease or Obesity. *J Nutr*. 2015;145(8):1687–91.
178. Pirazzi C, Valenti L, Motta BM, Pingitore P, Hedfalk K, Mancina RM, et al. PNPLA3 has retinyl-palmitate lipase activity in human hepatic stellate cells. *Hum Mol Genet*. 2014;23(15):4077–85.
179. Bugianesi E, Manzini P, D'Antico S, Vanni E, Longo F, Leone N, et al. Relative contribution of iron bur-
- den, HFE mutations, and insulin resistance to fibrosis in nonalcoholic fatty liver. *Hepatology*. 2004;39(1):179–87.
180. Nelson JE, Bhattacharya R, Lindor KD, Chalasani N, Raaka S, Heathcote EJ, et al. HFE C282Y mutations are associated with advanced hepatic fibrosis in Caucasians with nonalcoholic steatohepatitis. *Hepatology*. 2007;46(3):723–9.
181. Tryndyak VP, Han T, Muskhelishvili L, Fuscoe JC, Ross SA, Beland FA, et al. Coupling global methylation and gene expression profiles reveal key pathophysiological events in liver injury induced by a methyl-deficient diet. *Mol Nutr Food Res*. 2011;55(3):411–8.
182. Liu F, Li H, Chang H, Wang J, Lu J. Identification of hepatocellular carcinoma-associated hub genes and pathways by integrated microarray analysis. *Tumori*. 101(2):206–14.
183. de Conti A, Ortega JF, Tryndyak V, Dreval K, Moreno FS, Rusyn I, et al. MicroRNA deregulation in nonalcoholic steatohepatitis-associated liver carcinogenesis. *Oncotarget*. 2017;8(51):88517–28.
184. Khalid A, Hussain T, Manzoor S, Saalim M, Khalil S. PTEN: A potential prognostic marker in virus-induced hepatocellular carcinoma. *Tumour Biol*. 2017;39(6):1010428317705754.
185. Singh AK, Kumar R, Pandey AK. Hepatocellular Carcinoma: Causes, Mechanism of Progression and Biomarkers. *Curr Chem genomics Transl Med*. 2018;12:9–26.
186. Higashitsuji H, Higashitsuji H, Itoh K, Sakurai T, Nagao T, Sumitomo Y, et al. The oncoprotein gankyrin binds to MDM2/HDM2, enhancing ubiquitylation and degradation of p53. *Cancer Cell*. 2005;8(1):75–87.