

Bölüm 6

SUCUL EKOSİSTEMLERDE MAKROFİT MİKROORGANİZMA İLİŞKİLERİ VE BU İLİŞKİLERİN ÇEVRESEL ETKİLERİ

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GİRİŞ

Sulak alanlar; zengin bitkisel ve hayvansal organizmalara barınak sağlayan, üzerindeki sığ su tabakası sayesinde suyla doymuş toprağa sahip olup, karasal ve sulcul ortamlar arasında bir geçiş zonu olarak karakterize edilirler.

Tatlı su ekosistemlerindeki bitkisel organizmalar; fitoplankton, diatom ve diğer algler gibi mikrofitlerle, makrofitlerden oluşmaktadır. “Makrofit” terimi; kısmen ya da tamamen su içinde yaşayan ve çıplak gözle görülebilen bitkileri ifade etmek için kullanılır^(1,2). Makrofitler taksonomik olarak; phanerogam, pteridophyte, br-yophyte ve makroskobik algleri içeir⁽²⁻⁴⁾. Yaşam formlarına göre ise; su üstü, su altı, serbest yüzen ve yüzen yapraklı makrofitler olarak gruplandırılırlar.

Yüzey sularının temel olarak, makrofit ve mikrofitlerin büyüebilmesi için, organik karbona ek olarak azot, fosfat ve diğer makro-nutrientleri içermeleri gerekmektedir⁽⁵⁾. Bu sularda mako ve mikrofitlerin yanı sıra, genellikle mikrobiyal döküntüler, biyofilm ve planktonik makroalg-bakteri toplulukları şeklinde göz-lemelenen mikrobiyal bileşenler de vardır^(6,7). Sulcul ortamlarda bakteri gelişimi genellikle bir yüzeye bağlı (biyofilm) olarak meydana gelir⁽⁸⁾. Biyofilm oluşumu sadece cansız yüzeylerde değil, aynı zamanda sulcul bitkiler üzerinde de gerçekleşir⁽⁹⁾. Biyofilm şeklindeki mikrobiyal topluluklar çoğunlukla, su altı bitkilerinin rizosferinde (kök çevresi), özellikle de rizoplan (kökün temas yüzeyi) bölgesinde ve sedimentin katı yüzeyinde bulunur. Değişik çevresel koşullar, örneğin; ötrofikasyon⁽¹⁰⁾ ve sudaki toksik maddeler biyofilmi ve biyofilmin yapısını etkiler⁽¹¹⁾.

Tatlı su sistemlerinin su kalitesi doğal bozulma, ötrofikasyon ve insan aktivitelerinin etkisine göre değerlendirilmektedir. Akuatik kirlilikle ilgili çok sayıda çalışma vardır⁽¹²⁻¹⁴⁾ ve kirliliğin biyolojik giderimiyle ilgili referanslar da⁽¹⁵⁻¹⁶⁾ su kirliliği çalışmalarının içinde yer almaktadır.

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SONUÇ

Sucul ortamlarda özellikle kök yüzeyinde bitki mikroorganizma etkileşimi yoğun olarak görülür. Bitki büyümekte olan yumuşak dokularını korumak için, farklı organik maddeler (amino asit, polisakkarit, lipid, fenolik birleşikler, nükleik asit vb.) salgılar. Salgılanan bu maddeler mikroorganizmaları cezbeder. Bitki mikroorganizma etkileşiminin doğasında, pozitif ya da negatif gelişmeler, etkileşimin şekline bağlı olarak oluşabilir. Buna ilaveten, çeşitli mikroorganizmalar birbirleriyle etkileşime girerek, biyofilm olarak adlandırılan bir mikrobiyal ağ oluştururlar. Bitki türlerinin ürettiği organik karbon ve kök yüzeylerindeki oksijen seviyesine bağlı olarak, üzerlerinde gelişen mikrobiyal birliğin yapısı da değişiklik gösterebilmektedir.

Derin sulardaki köklü makrofitler; radyal oksijen kaybı olarak bilinen, gövdeden köklere doğru gerçekleştirilen oksijen salınımının devamlılığını sağlamak zorundadırlar. Çünkü; gövdeden köke gönderilerek, kök yüzeyinden sedimente salınan oksijen sayesinde mikrobiyal ve kimyasal reaksiyonlar gerçekleşmektedir. Kök yüzeyindeki radyal oksijen kaybı, bu alanı elektron taşıma bölgesi haline getirir ve burada oksijen elektron alıcı görevi yaparak aerobik yaşamı destekler. Öte yandan, oksijenin düşük olduğu sedimentte ise; CO_2 , CH_4^+ ve NO_3^- gibi elektron alıcıları bulunur ve bunlar da aneorobik yaşamı desteklerler.

Sucul ortamlarda kök yüzeyi birçok biyolojik ve fiziko-kimyasal reaksiyonun oluştuğu aktif bir zondur. Bu reaksiyonlardan bağımsız olarak; her bir makrofit türü kendine özgü bir mikrofloraya sahiptir ve bu ilişki ile her iki organizma da hayatta kalabilmek için; gerek besin maddelerinin kullanımı ve gerekse kirleticilerin uzaklaştırılması amacıyla, işbirliği yaparlar. Bir örnek vermek gerekirse; bakteri türleri poly-aromatik hidrokarbonları ayrıştırarak bitki büyümesi için gerekli olan Indol Asetik Asit (IAA) hormonunu sentezlerler ⁽¹⁶⁾.

KAYNAKLAR

1. Westlake DF. (1974). Macrophytes. In RA Vollenweider (Ed.), *A Manual on Methods for Measuring Primary Production in Aquatic Environments. I. B. P. Handbook 12*. Blackwell Scientific: Oxford, 225 p.
2. Wetzel RG. (1983). *Limnology* (Second edit). Philadelphia: Saunders College Pub, 860 p.
3. Casper SJ & Krausch HD. (1981). Pteridophyta and Anthophyta 23/24. In H. Ettl et al. (Eds), *Die Süßwasserflora von Mitteleuropa*. Stuttgart: Gustav Fischer Verlag, 512 p.
4. Frahm JP & Frey W. (1992). *Moosflora* (3th edit). Stuttgart: Ulmer, 528 p.
5. Srivatava S, Gupta A & Chandra H. Managing water quality with aquatic macrophytes. *Rev. Environ. Sci. Bio/technol.* 2008; 7: 255-266.

6. Paerl HW& Pinckley JL. A mini review of microbial consortia: their roles in aquatic production and biogeochemical cycling. *Microb. Ecol.* 1996; 31 (3): 225-247.
7. Battin TJ, Kaplan LA, Newbold JD, et al. Contributions of microbial biofilms to ecosystem processes in stream mesocosm. *Nature*, 2003; 426, 439-442.
8. Costerton JW, Lewandowski Z, Caldwell DE, et al. Microbial Biofilms. *Annual. Rev. Microbiol.* 1995; 49: 711-745.
9. Hempel M, Blueme M, Blindow I, et al. Epiphytic bacteria community composition on two common submersed macrophytes in brackish water and freshwater. *BMC Microbiol.* 2008; 8: 58. Doi:10.1186/1471-2180-8-58.
10. Giaramida L, Manage PM, Edwards C, et al. Bacterial communities' response to microcystins exposure and nutrient availability-linking degradation capacity to community structure. *Int. Biodeterior. Biodegrad.* 2013; 84: 111-117.
11. Calherios CSC, Duque AE, Moura A, et al. Changes in the bacterial community structure in two-stage constructed wetlands with different plants for industrial wastewater treatment. *Biores. Technol.* 2009, 100(13): 3228-3235.
12. Nagai T, Imai A, Matsushige K, et al. Dissolved iron and its speciation in shallow eutrophic lake and its inflowing rivers. *Water Res.* 2007; 41: 775-784.
13. Camargo JA & Alonso A. Ecological and toxicological effects of inorganic nitrogen pollution in aquatic ecosystems: a global assessment. *Environ Int.* 2006; 32: 831-849.
14. Khan SA & Srivastava J. Interaction of ions in water affects water quality of freshwater lake: a case study of of major lakes of North India. *Environmentalist*, 2008; 28: 217-221.
15. Sooknah RD & Wilkie AC. Nutrient removal by floating aquatic macrophytes cultured in anaerobically digested flushed dairy manure wastewater. *Ecol. Eng.* 2004; 22 (1): 27-42.
16. Srivastava J, Naraian R, Kalra SJS, et al. Advances in microbial bioremediation and the factors influencing the process. *Int. J. Environ. Sci. Technol.* 2014; 11 (6): 1787-1800.
17. Stout LM & Nüsslein K. Biochemical potential of aquatic plant-microbe interactions. *Curr. Opin. Biotechnol.* 2010; 21: 339-345.
18. Sharma R, Sharma K, Singh N, et al. Rhizosphere biology of aquatic microbes in order to Access their bioremediation potential along with different aquatic macrophytes. *Rec. Res. Sci. Technol.* 2013; 5 (1): 29-32.
19. Lu Y, Nakai S, Hosomi M, et al. Stimulation of nitrogen removal in the rhizosphere of aquatic duckweed by root exudate components. *Planta.* 2014; 239 (3): 591-603.
20. Stottmeister U, Wiebner A, Kusch P, et al. Effects of plants and microorganisms in constructed wetlands for wastewater treatment. *Biotechnol. Adv.*, 2003; 22: 93-117.
21. Toyama T, Furukawa T, Maeda N, et al. Accelerated biodegradation of pyrene and benzo[a]pyrene in the *Phragmites australis* rhizosphere by bacteria-root exudates interactions. *Water Res.* 2011; 45: 1629-1638.
22. Chakraborty R, Mitra AK & Mukherjee S. Synergistic chromium bioremediation by Water Lettuce (*Pistia*) and bacteria (*Bacillus cereus* GXBC-1) interaction. *J. Biol. Chem. Res.* 2013; 30(2): 421-431.
23. Hahn MW. The microbial diversity of inland waters. *Curr. Opin. Biotechnol.*, 2006; 17: 256-261.
24. Zehr JP. Microbes in earth's aqueous environments. *Front Microbiol.* 2010; 1(4): 1-2.
25. Wang Y, Ke X, Wu L, et al. Community composition of ammonia-oxidising bacteria and archaea in rice field soil affected by nitrogen fertilization. *System Appl. Microbiol.* 2009; 32(1): 27-36.
26. Wei B, Yu X, Zhang S, et al. Comparison of the community structures of ammonium-oxidizing bacteria and archaea in rhizoplanes of floating aquatic macrophytes. *Microbiol. Res.* 2011; 166: 468-474.

27. Gagnon V, Chazarenc F, Comeau Y, et al. Influence of macrophyte species on microbial density and activity in constructed wetlands. *Water Sci. Technol.* 2007; 56(3), 249-254.
28. Branda S, Vik A, Friedman L, et al. Biofilms: the matrix revisited. *Trends Microbiol.* 2005; 13: 20-26.
29. Yannarell AC & Triplett EW. Within and between lake variability in the composition of bacterioplankton communities: investigations using multiple spatial scales. *Appl. Environ. Microbiol.* 2004; 70: 214-223.
30. Kierek-Pearson K & Karatan E. Biofilm development in bacteria. *Adv. Appl. Microbiol.*, 2005; 57: 79-111.
31. Crump BC & Koch EW. Attached bacterial populations shared by four species of aquatic angiosperms. *Appl. Environ. Microbiol.* 2008; 74(19): 5948-5957.
32. Truu M, Juhanson J, & Truu J. Microbial biomass activity and community composition in constructed wetlands. *Sci. Total Environ.* 2009; 407: 3958-3971.
33. Srivasta JK, Chandra H, Kalra JS, et al. Plant-microbe interaction in aquatic system and their role in the management of water quality: a review. *Appl. Water Sci.* 2017; 7: 1079-1090.
34. Buosi PRB, Pauleto GM, Lansac-Toha FA, et al. Ciliate community associated with aquatic macrophytes roots: effects of nutrient enrichment on the community composition and species richness. *Eur. J. Protistol.* 2011; 47(2): 86-102.
35. Calherios CSC, Teixeira A, Pires C, et al. Bacterial community dynamics in horizontal flow constructed wetlands with different plants for high salinity industrial waste water polishing. *Water Res.* 2010; 44(17), 5032-5038.
36. Bronmark C. Interactions between epiphytes, macrophytes and freshwater snails: A review. *Journal of Molluscan studies*, 1989; 55: 299.
37. Weyens N, van-der Lelie D, Taghavi S, et al. Phytoremediation: plant-endophyte partnerships take the challenge. *Curr. Opin. Biotechnol.* 2009; 20: 248-254.
38. Neilsen LB, Finstek K, Welsh DT, et al. Sulphate reduction and nitrogen fixation rates associated with roots, rhizomes and sediments from *Zostera noltii* and *Spartina maritima* meadows. *Environ. Microbiol.* 2001; 3: 63-71.
39. Özbay H. Kırmızı Eğrelti *Azolla*: Biyolojisi ve Tarımda Kullanımı. *Türk Sucul Yaşam Dergisi*, 2004; 2: 590-601.
40. Bronmark C. Interaction between macrophytes, epiphytes and herbivores: an experimental approach. *Oikos*, 1985; 45: 26-30.
41. Hutchinson GE. (1975). *A Treatise on Limnology, vol III, Limnological Botany*. New York, John Wiley and Sons, 672 p.
42. Sand-Jensen K. Effects of epiphytes on eelgrass photosynthesis, *Aquatic Botany*, 1977; 3: 55-63.
43. Phillips GL, Eminson DF, Moss B. A mechanism to account for macrophytes decline in progressively eutrophicated freshwaters. *Aquatic Botany*, 1978; 4, 103-126.
44. Bulthuis DA & Woelkerling WmJ. Biomass accumulation and shading effects of epiphytes of seagrass, *Heterozostera tasmanica*, in Victoria, Australia. *Aquatic Botany*, 1983; 16: 137-148.
45. Baker JH. (1981). Direct observation and enumeration of microbes on plant surfaces by light microscope. In JP. Blakeman (Ed.) *Microbial Ecology of phylloplane* (pp.3-14). London: Academic Press.
46. Baker, JH. & Orr, DR. Distribution of epiphytic bacteria on freshwater plants, *Journal of Ecology*, 1986, 75: 155-165.
47. Jacques MA, Kinkel LL & Morris CE. Population sizes, immigration, and growth of epiphytic bacteria on leaves of different age and positions of field-grown endive (*Cichorium endivia* var. *latifolia*). *Applied and Environmental Microbiology*, 1995; 61 (3): 899-906.

48. Bowker DW & Denny, P. The seasonal succession and distribution of epifititk algae in phyllosphere of *Lemna minor* L. *Arch. Hydrobiol.*, 1980; 90: 39-55.
49. Round FE. (1973). *The Biology of the algae*. London: Edward Alnold Publication, 278 p.
50. Özbay H. Epiphytic Bacteria population on *Polygonum amphibium* –from two different lakes in Northeastern Turkey. *Journal of Freshwater Ecology*, 2008; 23 (3): 487-489.
51. Godward MBE. An investigation of the causal distribution of algal epiphytes. *Bech. Bot. Zbl.*, 1934; 52: 506-539.
52. Stangenburg M. Bacteriostatic effects of some algae and *Lemna minor* eextracts. *Hydrobiologia*, 1968; 32: 88-96.
53. Fitzgerald GP. Some factors in the competition on antoganism among bacteria, algae and aquatic weeds. *Journal of Phycology*, 1969; 5: 351-359.
54. Hossell JC & Baker JH. A note on the enumeration of epiphytic bacteria by microscopic methods with particular reference to two freshwater plants. *Journal of Applied Bacteriology*, 1979;46: 87-92.
55. Özbay H. Bacterial community on submersed plants in running water, *Desalination and Water Treatment*, 1018; 106: 267-272.
56. Christensen PB, Revsbech NP & Sand-Jensen K. Microsensor analysis of oxygen in the rhizosphere of the aquatic macrophyte *litorella uniflora* (L.) Ascherson. *Plant Physiol.* 1994; 105: 847-852.
57. Sand-Jensen K, Pederson :oetid *Lobelia dortmanna* and the marine seagrass *Zostera marina*. *Ann. Bot.* 2005; 96: 613-623.
58. Soda S, Ike M, Ogasawara Y, et al. Effects of light intensity and water teperature on oxygen release from roots into water lettuce rhizosphere. *Water Res.* 2007; 41: 487-491.
59. Brix H. Do macrophytes play a role in constructed treatment wetlands? *Water Sci. Technol.* 1997, 35: 11-17.
60. Weissner A, Kusch P, Kastner M, et al. Abilities of helophyte species to release oxygen into rhizospheres with varying redox conditions inlaboratory-scale hydroponic systems. *Int. J. Phytotorem.* 2002; 4: 1-15.
61. Bodelier PLE. (2003). Interactions between oxygen-releasing roots and microbial processes in flooded soils ans sediments. In H de Kroon, EJW Visser (Eds.), *Root ecology, series ecological studies vol 168* (pp. 331-362). Berlin: Springer.
62. Laanbroek HJ. Methane emission from natural wetlands: interplay between emergent macrophytes and soil microbial processes. A mini-review. *Ann. Bot.* 2010; 105: 141-153.
63. Germ M & Simcic T. Vitality of aquatic plants and microbial activity of sediment in an oligotrophic lake (Lake Bohinj, Slovenia). *J. Limnol.* 2011; 70(2): 305-312.
64. Conrad R. (2004). Methanogenic microbial communities associated with aquatic plants. In A Varma, L Abbott, D Werner, R Hampp (Eds.), *Plant surface microbiology* (pp. 35-50). Berlinn, Heidelberg: Springer.
65. Pilson-Smits EAH & Freeman JL. Environmental cleanup using plants: biotechnological advances and ecological considerations. *Front Ecol. Environ.* 2006; 4(4): 203-210.
66. Fennell DE, Du S, Liu H, et al. (2011). Dehalogenation of polychlorinated dibenzo-p-dioxins and dibenzofurans, polychlorinated biphenyls and brominated flame retardants and potential as a bioremediation strategy. In M Moo-Young, M Butler, C Webb et al (Eds.), *Comprehensive biotechnology (second edition, pp, 136-149)*. Oxford: Pergamon press.
67. Mori K, Toyama T & Sei K. Surfactants degradind activities in the rhizosphere of giant duckweed (*Spirodela polyrhiza*). *Jpn. J. Water Traet. Biol.* 2005; 41: 129-140.

68. Gasol JM & Duarte CM. Comparative analysis in aquatic microbial ecology: how far do they go? *FEMS Microbiol. Ecol.* 2000; 31: 99-106.
69. Okabe S, Kokazi T & Watanabe Y. Biofilm formation potentials in drinking waters treated by different advance treatment processes. *Water Sci. Technol.* 2002; 2(4): 97-104.
70. Simpson DR. Biofilm processes in biologically active carbon water purification. *Water Res.* 2009; 42, 2839-2848.
71. Tranvik LJ. (1998). Degradation of dissolved organic matter in humic waters by bacteria. In DO Hensen, LJ Tranvik (Eds.), *Aquatic humic substances-book series, ecological studies* (pp. 259-283). Berlin Heidelberg: Springer.
72. Semrau JD, DiSpirito AA & Yoon S. Methanotrops and copper. *FEMS Microbiol. Rev.* 2010; 34: 496-531.
73. Kraemer U. Phytoremediation. Novel approaches to cleaning up polluted soils. *Curr. Opin. Biotechnol.* 2005; 16: 1-9.
74. Rai PK. Heavy metal phytoremediation from aquatic ecosystems with special reference to aquatic macrophytes. *Crit. Rev. Env. Sci. Technol.* 2009; 39: 697-753.
75. Munoz R, Alvarez MT, Munoz A. et al. Sequential removal of heavy metals and organic pollutants using an algal-bacterium consortium. *Chemosphere*, 2006; 63(6): 903-911.
76. Zhou Q, Zhang J, Fu J, et al. Biomonitoring: an appealing tool for assessment of metal pollution in the aquatic ecosystem. *Analyt. Chim. Acta*, 2008; 606: 135-150.
77. Hansel CM, Fendorf S, Sutton S, et al. Characterization of Fe plaque and associated metals on the roots of mine-waste impacted aquatic plants. *Environ. Sci. Technol.* 2001; 35(19): 3863-3868.
78. King GM & Garey MA. Ferric iron reduction by bacteria associated with the roots of freshwater and marine macrophytes. *Appl. Environ. Microbiol.* 1999; 65(10): 4393-4398.
79. Tripathi M, Munot HP, Shouche Y, et al. Isolation and functional characterization of siderophore-producing lead- and cadmium-resistant *Pseudomonas putida* KNP9. *Curr. Microbiol.* 2005; 50: 233-237.
80. Burd GI, Dixon DG & Glick BR. Plant growth-promoting bacteria that decrease heavy metal toxicity in plants. *Can. J. Microbiol.* 2000; 46, 237-245.
81. Machemer SD & Wildeman TR. Adsorption compared with sulphide precipitation as metal removal processes from acid mine drainage in a constructed wetland. *Contam. Hydrol.* 1992; 9: 115-131.
82. Han X & Gu J. (2010). Sorption and transformation of toxic metals by microorganisms. In R Mitchell, J Gu (Eds.), *Environmental microbiology* (pp.153-176). Hoboken: Wiley Blackwell.
83. Sraj-Krzic N, Pongrac P, Klemenc M, et al. Mycorrhizal colonization in plants from intermittent aquatic habitats. *Aquat. Bot.* 2006; 85: 331-336.
84. Srivastava J, Shuckla D, Chand V, et al. Mycorrhizal colonization affects the survival of *Vetiveria zizanioides* (L.) Nash grown in water containing As(III). *Clean-Soil Air Water.* 2010, 38(8): 771-774.