

## SANTRAL SINİR SİSTEMİ TÜMÖRLERİNDE İMMUNOTERAPİNİN YERİ

Utku OFLAZOĞLU<sup>1</sup>

### GİRİŞ

Glioma en sık görülen primer malign beyin tümörüdür, yetişkinlerde vakaların yaklaşık % 80'ini oluşturmaktadır. Glial kaynaklı tümörler, glial fibril asidik protein pozitif (GFAPC) astrositik tümörler, oligodendrogliomalar, ependimomlar ve alt tiplerin bir karışımını içeren histolojik alt tipe göre sınıflandırılır<sup>1</sup>. Bunlardan, GBM olarak adlandırılan astrositik glioma derece IV, tanı sonrası ortalama 14,6 ay sağkalma sahip olup ortalama 5 yıllık hayatı kalma oranı % 5'in altında olan en yaygın ve en ölümcül alt tiptir<sup>2,3</sup>. Rezeksiyon, radyoterapi ve kemoterapiyi birleştiren mevcut tedaviler istilacı sınırlardan / inoperabl cerrahi yataktan kaynaklanan rezidüel hastalığa dayalı tümör nüksünü önleyememektedir. Gen tedavisi gibi yeni yaklaşımlara, hedefleyici kemoterapötiklere ve / veya radyoterapötik yöntemlere rağmen yeni tanı konulan GBM hastalarının tedavisinde daha iyi tedavi seçeneklerine ihtiyaç son 10 yılda değişmeden kalmıştır. Ayrıca, tekrarlayan GBM'li hastalar için standart bir tedavi yoktur. SSS'de metastatik tümörlerin prevalansı GBM vakalarının sayısını fazlasıyla aşmaktadır, ancak genel sağkalım (GS) benzer şekilde düşüktür. Bu makalede, SSS'deki agresif tümörleri kalıcı olarak kontrol etmek için aşılama, immun kontrol noktası blokajı, adoptive T hücre transfer gibi yeni ve / veya genişletilmiş yaklaşımlar ve bunların etkinliği inceleneciktir.

### SSS TÜMÖRLERİ VE İMMÜNOSUPRESYON

Santral sinir sistemi'nin (SSS) başlangıçta, subkutan veya intramusküler büyümeye kıyasla beyine intrakranial olarak enjekte edilen sıçan osteosarkom hücreleri-

<sup>1</sup> İzmir Katip Çelebi Üniversitesi Atatürk Eğitim ve Araştırma Hastanesi, Tıbbi Onkoloji Kliniği, İzmir/Türkiye u.oflaz35@gmail.com

gu olarak, ikili PD-L1 ve CTLA-4 blokajına karşı dirençli olan uzun süreli klinik öncesi melanomlar, farelerin yaklaşık %33’ünde aşılama ortadan kalkmıştır; ancak aşılama anti-PD-L1 ile kombine edildiğinde bu oran farelerde %80’e çıkmaktadır.<sup>21</sup> Subkütanöz tümörlere yönelik bir klinik öncesi bağımsız çalışmada, PD-(L)1 ve CTLA-4 inhibisyonuyla kombine edilen aşılama, ikili ve monoterapötik tedaviyle kıyaslandığında farelerde tümör rejeksyonu ve sağkalımı iyileştirmiştir.<sup>94</sup> Geçmişteki çalışmaların çoğunluğu SSS dışı tümör modelleri üzerinde gerçekleştirildiğinden dolayı, gelecekteki klinik öncesi ve klinik çalışmaların bu tedavi yaklaşımını GBM hastalarında değerlendirmesine ihtiyaç duyulmaktadır.

## SONUÇ

GBM anatomik lokalizasyonu nedeniyle tedavisi zor olan aynı zamanda geleneksel tedavilere dirençli olan oldukça fazla immunsüoresif bir tümördür. Beyinin metastatik tümörleri GBM’ e kıyasla 20 kat daha sık görülür. Beyin tümörleri için geçmiş immünoterapötik çabalar ağırlıklı olarak ümit verici bağışıklık aktivitesi ve klinik tepkiler elde etmiş olan terapötik aşılamaya odaklanmıştır. Gen tedavisi de dahil olmak üzere mevcut umut eden yaklaşımları daha da fazla test etme ihtiyacı yeni nesil terapötikler (yani, IDO inhibitörleri / STING agonistleri), ve yeni immünoterapötik kombinasyonları test etmeyi geliştirir. Antitümör immün yanıtları inflamasyon bağlamında gerçekleştiğinden, tümör ve tedaviye bağlı inflamasyonun ilave etkisi/sinerjisi beyin ödeme ve nörolojik tehlikeye neden olma olasılığı tanınmalıdır. Deksametazon rutin olarak beyin ödemi önlemek için kullanılmasına rağmen, immünoterapötik çalışmalarında kullanımı da aşırı derecede immünsüpresif olduğu için düşük dozlarla sınırlıdır. Yeni nesil CNS immünoterapileri, eğer daha etkiliyse, beyin ödemi ve nörolojik tehlike için daha da yüksek bir risk taşıyabilir, bu nedenle immünsüpresif olmayan anti-enflamatuar yaklaşımların belirlenmesi önemlidir. VEGF inhibitörü bir antikor olan bevacizumab’ı kullanmak sekonder enflamasyonu azaltan, şu anda GBM immünoterapisi ile kombinasyon halinde araştırılan bu yaklaşımlardan biridir. (NCT02336165, NCT01814813). Gelecekteki çalışmaları, hastalara sürekli gelişen seçenekleri sunmaya odaklanmalı, ayrıca CNS malignitelerinin özel olarak hedeflenmesi gereken benzersiz immünsüpresif fenotiplere sahip olduğunu kabul etmelidir.

**Anahtar Kelimeler:** Gliom, immünoterapi, aşılama, SSS,

## KAYNAKLAR

- Ostrom QT, Bauchet L, Davis FG, Deltour I, Fisher JL, Langer CE, Pekmezci M, Schwartzbaum JA, Turner MC, Walsh KM et al. The epidemiology of glioma in adults: a “state of the science” review. Neuro-Oncol 2014; 16:896-913.
- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, Belanger K, Brandes

- AA, Marosi C, Bogdahn U et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *New Eng J Med* 2005; 352:987-96.
3. Krex D, Klink B, Hartmann C, von Deimling A, Pietsch T, Simon M, Sabel M, Steinbach JP, Heese O, Reifenberger G et al. Long-term survival with glioblastoma multiforme. *Brain* 2007; 130:2596-606.
  4. Shirai Y. On the transplantation of the rat sarcoma in adult heterogenous animals. *Jap Med World* 1921; 1:14-5.
  5. Carson MJ, Doose JM, Melchior B, Schmid CD, Ploix CC. CNS immune privilege: hiding in plain sight. *Immunological Rev* 2006; 213:48-65.
  6. Rivest S. Regulation of innate immune responses in the brain. *Nature Rev Immunol* 2009; 9:429-39.
  7. Farin A, Suzuki SO, Weiker M, Goldman JE, Bruce JN, Canoll P. Transplanted glioma cells migrate and proliferate on host brain vasculature: a dynamic analysis. *Glia* 2006; 53:799-808.
  8. Watkins S, Robel S, Kimbrough IF, Robert SM, Ellis-Davies G, Sontheimer H. Disruption of astrocyte-vascular coupling and the bloodbrain barrier by invading glioma cells. *Nature Communications* 2014; 5:4196.
  9. Prins RM, Soto H, Konkankit V, Odesa SK, Eskin A, Yong WH, Nelson SF, Liau LM. Gene expression profile correlates with Tcell infiltration and relative survival in glioblastoma patients vaccinated with dendritic cell immunotherapy. *Clin Cancer Res* 2011; 17:1603-15.
  10. Gill BJ, Pisapia DJ, Malone HR, Goldstein H, Lei L, Sonabend A, Yun J, Samanamud J, Sims JS, BanuMet al. MRI-localized biopsies reveal subtype- specific differences in molecular and cellular composition at the margins of glioblastoma. *Proc Natl Acad Sci U S A* 2014; 111:12550-5.
  11. Mitsuka K, Kawataki T, Satoh E, Asahara T, Horikoshi T, Kinouchi H. Expression of indoleamine 2,3-dioxygenase and correlation with pathological malignancy in gliomas. *Neurosurgery* 2013; 72:1031-8.
  12. Wainwright DA, Balyasnikova IV, Chang AL, Ahmed AU, Moon KS, Auffinger B, Tobias AL, Han Y, Lesniak MS. IDO expression in brain tumors increases the recruitment of regulatory T cells and negatively impacts survival. *Clin Cancer Res* 2012; 18:6110-21.
  13. Zhai L, Lauing KL, Chang AL, Dey M, Qian J, Cheng Y, Lesniak MS, Wainwright DA. The role of IDO in brain tumor immunotherapy. *J Neurooncol* 2014; 123(3):395-403.
  14. Heimberger AB, Abou-Ghazal M, Reina-Ortiz C, Yang DS, Sun W, Qiao W, Hiraoka N, Fuller GN. Incidence and prognostic impact of FoxP3C regulatory T cells in human gliomas. *Clin Cancer Res* 2008; 14:5166-72.
  15. Parsa AT, Waldron JS, Panner A, Crane CA, Parney IF, Barry JJ, Cachola KE, Murray JC, Tihan T, Jensen MC et al. Loss of tumor suppressor PTEN function increases B7-H1 expression and immunoresistance in glioma. *Nat Med* 2007; 13:84-8.
  16. Bloch O, Crane CA, Kaur R, Safaei M, Rutkowski MJ, Parsa AT. Gliomas promote immunosuppression through induction of B7-H1 expression in tumor-associated macrophages. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2013; 19:3165-75.
  17. Berghoff AS, Kiesel B, Widhalm G, Rajky O, Ricken G, Wohrer A, Dieckmann K, Filipits M, Brandstetter A, Weller M et al. Programmed death ligand 1 expression and tumor-infiltrating lymphocytes in glioblastoma. *Neuro-oncology* 2014; 17(8):1064-75.
  18. Fecchi PE, Ochiai H, Mitchell DA, Grossi PM, Sweeney AE, Archer GE, Cummings T, Allison JP, Bigner DD, Sampson JH. Systemic CTLA-4 blockade ameliorates glioma-induced changes to the CD4C T cell compartment without affecting regulatory T-cell function. *Clin Cancer Res* 2007; 13:2158-67.
  19. Grohmann U, Orabona C, Fallarino F, Vacca C, Calcinaro F, Falorni A, Candeloro P, Belladonna ML, Bianchi R, Fioretti MC et al. CTLA-4-Ig regulates tryptophan catabolism in vivo. *Nat Immunol* 2002; 3:1097-101.
  20. Thomas DL, Kim M, Bowerman NA, Narayanan S, Kranz DM, Schreiber H, Roy EJ. Recurrence of intracranial tumors following adoptive T cell therapy can be prevented by direct and indirect

- killing aided by high levels of tumor antigen cross-presented on stromal cells. *J Immunol* 2009; 183:1828-37.
21. Binder DC, Engels B, Arina A, YuP, Slauch JM, Fu YX, Garrison T, Burnette B, Idel C, Zhao Met al. Antigen-specific bacterial vaccine combined with anti-PD-L1 rescues dysfunctional endogenous T cells to reject long established cancer. *Cancer Immunol Res* 2013; 1:123-33;
  22. Brooks CL, Schietinger A, Borisova SN, Kufer P, Okon M, Hirama T, Mackenzie CR, Wang LX, Schreiber H, Evans SV. Antibody recognition of a unique tumor-specific glycopeptide antigen. *Proc Natl Acad Sci U S A* 2010; 107:10056-61.
  23. Schietinger A, Philip M, Schreiber H. Specificity in cancer immunotherapy. *Semin Immunol* 2008; 20:276-85.
  24. Carrel S, de Tribolet N, Mach JP. Expression of neuroectodermal antigens common to melanomas, gliomas, and neuroblastomas. I. Identification by monoclonal anti-melanoma and anti-glioma antibodies. *Acta Neuropathol* 1982; 57:158-64.
  25. Bloch O, Crane CA, Fuks Y, Kaur R, Aghi MK, Berger MS, Butowski NA, Chang SM, Clarke JL, McDermott MW et al. Heat-shock protein peptide complex-96 vaccination for recurrent glioblastoma: a phase II, single-arm trial. *Neuro-oncology* 2014; 16:274-9.
  26. Phuphanich S, Wheeler CJ, Rudnick JD, Mazer M, Wang H, Nuno MA, Richardson JE, Fan X, Ji J, Chu RM et al. Phase I trial of a multi-epitope-pulsed dendritic cell vaccine for patients with newly diagnosed glioblastoma. *Cancer Immunol Immunother* 2013; 62:125-35.
  27. Okada H, Kalinski P, Ueda R, Hoji A, Kohanbash G, Donegan TE, Mintz AH, Engh JA, Bartlett DL, Brown CK et al. Induction of CD8+T-cell responses against novel glioma-associated antigen peptides and clinical activity by vaccinations with  $\{\alpha\}$ -type 1 polarized dendritic cells and polyinosinic-polycytidylic acid stabilized by lysine and carboxymethylcellulose in patients with recurrent malignant glioma. *J Clin Oncol* 2011; 29:330-6.
  28. Fadul CE, Fisher JL, Hampton TH, Lallana EC, Li Z, Gui J, Szczepiorkowski ZM, Tosteson TD, Rhodes CH, Wishart HA et al. Immune response in patients with newly diagnosed glioblastoma multiforme treated with intranodal autologous tumor lysate-dendritic cell vaccination after radiation chemotherapy. *J Immunother* 2011; 34:382-9.
  29. Ardon H, Van Gool S, Lopes IS, Maes W, Sciot R, Wilms G, Demaezel P, Bijntebier P, Claes L, Goffin J et al. Integration of autologous dendritic cell-based immunotherapy in the primary treatment for patients with newly diagnosed glioblastoma multiforme: a pilot study. *J Neuro-oncol* 2010; 99:261-72.
  30. Valle RD, de Cerio AL, Inoges S, Tejada S, Pastor F, Villanueva H, Gallego J, Espinos J, Aristu J, Idoate MA et al. Dendritic cell vaccination in glioblastoma after fluorescence-guided resection. *World J Clin Oncol* 2012; 3:142-9.
  31. Chang CN, Huang YC, Yang DM, Kikuta K, Wei KJ, Kubota T, Yang WK. A phase I/II clinical trial investigating the adverse and therapeutic effects of a postoperative autologous dendritic cell tumor vaccine in patients with malignant glioma. *J Clin Neurosci* 2011; 18: 1048-54.
  32. Monach PA, Meredith SC, Siegel CT, Schreiber H. A unique tumor antigen produced by a single amino acid substitution. *Immunity* 1995; 2:45-59.
  33. Peloski CE, Ballman KV, Furth AF, Zhang L, Lin E, Sulman EP, Bhat K, McDonald JM, Yung WK, Colman H et al. Epidermal growth factor receptor variant III status defines clinically distinct subtypes of glioblastoma. *J Clin Oncol* 2007; 25:2288-94.
  34. Heimberger AB, Hlatky R, Suki D, Yang D, Weinberg J, Gilbert M, Sawaya R, Aldape K. Prognostic effect of epidermal growth factor receptor and EGFRvIII in glioblastoma multiforme patients. *Clin Cancer Res* 2005; 11:1462-6.
  35. Humphrey PA, Wong AJ, Vogelstein B, Zalutsky MR, Fuller GN, Archer GE, Friedman HS, Kwatra MM, Bigner SH, Bigner DD. Anti-synthetic peptide antibody reacting at the fusion junction of deletion-mutant epidermal growth factor receptors in human glioblastoma. *Proc Natl Acad Sci U S A* 1990; 87:4207-11.
  36. Sampson JH, Heimberger AB, Archer GE, Aldape KD, Friedman AH, Friedman HS, Gilbert MR, Herndon JE 2nd, McLendon RE, Mitchell DA et al. Immunologic escape after prolonged

- progressionfree survival with epidermal growth factor receptor variant III peptide vaccination in patients with newly diagnosed glioblastoma. *J Clin Oncol* 2010; 28:4722-9.
- 37. Schuster J, Lai RK, Recht LD, Reardon DA, Paleologos NA, Groves MD, Mrugala MM, Jensen R, Baehring JM, Sloan A et al. A phase II, multicenter trial of rindopepimut (CDX-110) in newly diagnosed glioblastoma: the ACT III study. *Neuro-oncology* 2015; 17(6):854- 61.
  - 38. Sampson JH, Aldape KD, Archer GE, Coan A, Desjardins A, Friedman AH, Friedman HS, Gilbert MR, Herndon JE, McLendon RE et al. Greater chemotherapy-induced lymphopenia enhances tumorspecific immune responses that eliminate EGFRvIII-expressing tumor cells in patients with glioblastoma. *Neuro-Oncol* 2011; 13:324-33.
  - 39. Lesniak MS. Immunotherapy for glioblastoma: the devil is in the details. *J Clin Oncol* 2011; 29:3105.
  - 40. Weller M, Butowski N, Tran DD, Recht LD, Lim M et al. Rindopepimut with temozolomide for patients with newly diagnosed, EGFRvIII-expressing glioblastoma (ACT IV): a randomised, double-blind, international phase 3 trial. *Lancet Oncol*. 2017 Oct;18(10):1373-1385
  - 41. Yan H, Parsons DW, Jin G, McLendon R, Rasheed BA, Yuan W, Kos I, Batinic-Haberle I, Jones S, Riggins GJ et al. IDH1 and IDH2 mutations in gliomas. *N Eng J Med* 2009; 360:765-73.
  - 42. Schumacher T, Bunse L, Pusch S, Sahm F, Wiestler B, Quandt J, Menn O, Osswald M, Oezen I, Ott M et al. A vaccine targeting mutant IDH1 induces antitumour immunity. *Nature* 2014; 512:324-7.
  - 43. Basu S, Binder RJ, Ramalingam T, Srivastava PK. CD91 is a common receptor for heat shock proteins gp96, hsp90, hsp70, and calreticulin. *Immunity* 2001; 14:303-13.
  - 44. Crane CA, Han SJ, Ahn B, Oehlke J, Kivett V, Fedoroff A, Butowski N, Chang SM, Clarke J, Berger MS et al. Individual patient-specific immunity against high-grade glioma after vaccination with autologous tumor derived peptides bound to the 96 KD chaperone protein. *Clin Cancer Res* 2013; 19:205-14.
  - 45. Engels B, Engelhard VH, Sidney J, Sette A, Binder DC, Liu RB, Kranz DM, Meredith SC, Rowley DA, Schreiber H et al. Relapse or eradication of cancer is predicted by peptide-major histocompatibility complex affinity. *Cancer Cell* 2013; 23:516-26.
  - 46. Robbins PF, Lu YC, El-Gamil M, Li YF, Gross C, Gartner J, Lin JC, Teer JK, Cliften P, Tycksen E et al. Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nat Med* 2013; 19:747-52.
  - 47. van Rooij N, van Buuren MM, Philips D, Velds A, Toebees M, Heemskerk B, van Dijk LJ, Behjati S, Hilkmann H, El Atmioui D et al. Tumor exome analysis reveals neoantigen-specific T-cell reactivity in an ipilimumab-responsive melanoma. *J Clin Oncol* 2013; 31:e439-42.
  - 48. Binder DC, Schreiber H. High-affinity peptide-based anticancer vaccination to overcome resistance to immunostimulatory antibodies. *Oncioimmunology* 2013; 2:e26704.
  - 49. Binder DC, Schreiber H. Dual Blockade of PD-1 and CTLA-4 Combined with Tumor Vaccine Effectively Restores T-Cell Rejection Function in Tumors-Letter. *Cancer Res* 2014; 74:632.
  - 50. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* 1996; 271:1734-6.
  - 51. Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. *J Exp Med* 1995; 182:459- 65.
  - 52. Wing K, Onishi Y, Prieto-Martin P, Yamaguchi T, Miyara M, Fehervari Z, Nomura T, Sakaguchi S. CTLA-4 control over Foxp3C regulatory T cell function. *Science* 2008; 322:271-5.
  - 53. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC et al. Improved survival with ipilimumab in patients with metastatic melanoma. *New Eng J Med* 2010; 363:711-23.
  - 54. Margolin K, Ernsthoff MS, Hamid O, Lawrence D, McDermott D, Puzanov I, Wolchok JD, Clark JI, Sznol M, Logan TF et al. Ipilimumab in patients with melanoma and brain metastases: an open-label, phase 2 trial. *Lancet Oncol* 2012; 13:459-65.
  - 55. Barber DL, Wherry EJ, Masopust D, Zhu B, Allison JP, Sharpe AH, Freeman GJ, Ahmed R. Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature* 2006; 439:682-7.

56. Taube JM, Anders RA, Young GD, Xu H, Sharma R, McMiller TL, Chen S, Klein AP, Pardoll DM, Topalian SL et al. Colocalization of inflammatory response with B7-h1 expression in human melanocytic lesions supports an adaptive resistance mechanism of immune escape. *Sci Transl Med* 2012; 4:127ra37.
57. Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Eng J Med* 2012; 366:2455-65.
58. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD, Carvajal RD, Sosman JA, Atkins MB et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Eng J Med* 2012; 366:2443-54.
59. Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, Chmielowski B, Spasic M, Henry G, Ciobanu V et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* 2014; 515:568-71.
60. Curran MA, Montalvo W, Yagita H, Allison JP. PD-1 and CTLA-4 combination blockade expands infiltrating T cells and reduces regulatory T and myeloid cells within B16 melanoma tumors. *Proc Natl Acad Sci U S A* 2010; 107:4275-80.
61. Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, Linette GP, Meyer N, Giguere JK, Agarwala SS et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N Eng J Med* 2015; 372:2006-17.
62. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, Schadendorf D, Dummer R, Smylie M, Rutkowski P et al. Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. *N Eng J Med* 2015; 373(1):23-34.
63. Wainwright DA, Chang AL, Dey M, Balyasnikova IV, Kim CK, Tobias A, Cheng Y, Kim JW, Qiao J, Zhang L et al. Durable therapeutic efficacy utilizing combinatorial blockade against IDO, CTLA-4, and PD-L1 in mice with brain tumors. *Clin Cancer Res* 2014; 20:5290-301.
64. Blumenthal DT, Yalon M, Vainer GW, et al. Pembrolizumab: first experience with recurrent primary central nervous system (CNS) tumors. *J Neurooncol* 2016; 129:453.
65. Chamberlain MC, Kim BT. Nivolumab for patients with recurrent glioblastoma progressing on bevacizumab: a retrospective case series. *J Neurooncol* 2017; 133:561.
66. Omuro A, Vlahovic G, Lim M, et al. Nivolumab with or without ipilimumab in patients with recurrent glioblastoma: results from exploratory phase I cohorts of CheckMate 143. *Neuro Oncol* 2018; 20:674.
67. Mantica M, Pritchard A, Lieberman F, Drappatz J. Retrospective study of nivolumab for patients with recurrent high grade gliomas. *J Neurooncol* 2018; 139:625.
68. Kurz SC, Cabrera LP, Hastie D, et al. PD-1 inhibition has only limited clinical benefit in patients with recurrent high-grade glioma. *Neurology* 2018; 91:e1355.
69. Reardon D, Omuro A, Brandes A, et al. Randomized phase 3 study evaluating the efficacy and safety of nivolumab vs bevacizumab in patients with recurrent glioblastoma: CheckMate 143. *Neuro Oncol* 2017; 19 (suppl 3):iii21.
70. Cloughesy TF, Mochizuki AY, Orpilla JR, et al. Neoadjuvant anti-PD-1 immunotherapy promotes a survival benefit with intratumoral and systemic immune responses in recurrent glioblastoma. *Nat Med* 2019; 25:477.
71. David A. Reardon, Lakshmi Nayak, Katherine B. Peters, Jennifer Leigh Clarke, Justin T Jordan et al. Phase II study of pembrolizumab or pembrolizumab plus bevacizumab for recurrent glioblastoma (rGBM) patients. DOI: 10.1200/JCO.2018.36.15\_suppl.2006 *Journal of Clinical Oncology* 36, no. 15\_suppl (May 20 2018))
72. Rosenberg SA, Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. *Science* 2015; 348:62-8.
73. Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, Citrin DE, Restifo NP, Robbins PF, Wunderlich JR et al. Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy. *Clin Cancer Res* 2011; 17:4550-7.
74. Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, Chew A, Gonzalez VE, Zheng Z,

- Lacey SF et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N Eng J Med* 2014; 371:1507-17.
75. Cobbs CS, Harkins L, Samanta M, Gillespie GY, Bharara S, King PH, Nabors LB, Cobbs CG, Britt WJ. Human cytomegalovirus infection and expression in human malignant glioma. *Cancer Res* 2002; 62:3347-50.
76. Schuessler A, Smith C, Beagley L, Boyle GM, Rehan S, Matthews K, Jones L, Crough T, Dasari V, Klein K et al. Autologous T-cell therapy for cytomegalovirus as a consolidative treatment for recurrent glioblastoma. *Cancer Res* 2014; 74:3466-76.
77. Nair SK, De Leon G, Boczkowski D, Schmittling R, Xie W, Staats J, Liu R, Johnson LA, Weinhold K, Archer GE et al. Recognition and killing of autologous, primary glioblastoma tumor cells by human cytomegalovirus pp65-specific cytotoxic T cells. *Clin Cancer Res* 2014; 20:2684-94.
78. Ahmed N, Salsman VS, Kew Y, Shaffer D, Powell S, Zhang YJ, Grossman RG, Heslop HE, Gottschalk S. HER2-specific T cells target primary glioblastoma stem cells and induce regression of autologous experimental tumors. *Clin Cancer Res* 2010; 16:474- 85.
79. Johnson LA, Scholler J, Ohkuri T, Kosaka A, Patel PR, McGettigan SE, Nace AK, Dentchev T, Thekkat P, Loew A et al. Rational development and characterization of humanized anti-EGFR variant III chimeric antigen receptor T cells for glioblastoma. *Sci Transl Med* 2015; 7:275ra22.
80. Ahmed N., Brawley V., Hegde M., Bielamowicz K., Kalra M. et al. HER2-Specific Chimeric Antigen Receptor-Modified Virus-Specific T Cells for Progressive Glioblastoma. *JAMA Oncol.* 2017 Aug 1; 3(8): 1094-1101.
81. Lee Y, Auh SL, Wang Y, Burnette B, Wang Y, Meng Y, Beckett M, Sharma R, Chin R, Tu T et al. Therapeutic effects of ablative radiation on local tumor require CD8C T cells: changing strategies for cancer treatment. *Blood* 2009; 114:589-95.
82. Deng L, Liang H, Xu M, Yang X, Burnette B, Arina A, Li XD, Mauceri H, Beckett M, Darga T et al. STING-Dependent Cytosolic DNA Sensing Promotes Radiation-Induced Type I Interferon-Dependent Antitumor Immunity in Immunogenic Tumors. *Immunity* 2014; 41:843- 52.
83. Fujita M, Scheurer ME, Decker SA, McDonald HA, Kohanbash G, Kastenhuber ER, Kato H, Bondy ML, Ohlfest JR, Okada H. Role of type 1 IFNs in antglioma immunosurveillance—using mouse studies to guide examination of novel prognostic markers in humans. *Clin Cancer Res* 2010; 16:3409-19.
84. 74. Ohkuri T, Ghosh A, Kosaka A, Zhu J, Ikeura M, David M, Watkins SC, Sarkar SN, Okada H. STING contributes to antglioma immunity via triggering type I IFN signals in the tumor microenvironment. *Cancer Immunol Res* 2014; 2:1199-208.
85. Okada H, Scheurer ME, Sarkar SN, Bondy ML. Integration of epidemiology, immunobiology, and translational research for brain tumors. *Ann N Y Acad Sci* 2013; 1284:17-23.
86. Zeng J, See AP, Phallen J, Jackson CM, Belcaid Z, Ruzevick J, Durham N, Meyer C, Harris TJ, Albesiano E et al. Anti-PD-1 blockade and stereotactic radiation produce long-term survival in mice with intracranial gliomas. *Int J Radiat Oncol Biol Phys* 2013; 86:343-9.
87. Deng L, Liang H, Burnette B, Beckett M, Darga T, Weichselbaum RR, Fu YX. Irradiation and anti-PD-L1 treatment synergistically promote antitumor immunity in mice. *J Clin Invest* 2014; 124:687-95.
88. Demaria S, Kawashima N, Yang AM, Devitt ML, Babb JS, Allison JP, Formenti SC. Immune-mediated inhibition of metastases after treatment with local radiation and CTLA-4 blockade in a mouse model of breast cancer. *Clin Cancer Res* 2005; 11:728-34.
89. Twyman-Saint Victor C, Rech AJ, Maity A, Rengan R, Pauken KE, Stelekati E, Bencic JL, Xu B, Dada H, Odorizzi PM et al. Radiation and dual checkpoint blockade activate non-redundant immune mechanisms in cancer. *Nature* 2015; 520:373-7.
90. Belcaid Z, Phallen JA, Zeng J, See AP, Mathios D, Gottschalk C, Nicholas S, Kellett M, Ruzevick J, Jackson C et al. Focal radiation therapy combined with 4-1BB activation and CTLA-4 blockade yields long-term survival and a protective antigen-specific memory response in a murine glioma model. *PLoS One* 2014; 9:e101764.

91. Grimaldi AM, Simeone E, Giannarelli D, Muto P, Falivene S, Borzillo V, Giugliano FM, Sandomenico F, Petrillo A, Curvietto M et al. Abscopal effects of radiotherapy on advanced melanoma patients who progressed after ipilimumab immunotherapy. *Oncimmunology* 2014; 3:e28780.
92. Bai A, Higham E, Eisen HN, Wittrup KD, Chen J. Rapid tolerization of virus-activated tumor-specific CD8C T cells in prostate tumors of TRAMP mice. *Proc Natl Acad Sci U S A* 2008; 105:13003-8.
93. Lu H. TLR Agonists for Cancer Immunotherapy: Tipping the Balance between the Immune Stimulatory and Inhibitory Effects. *Front Immunol* 2014; 5:83.
94. Duraiswamy J, Kaluza KM, Freeman GJ, Coukos G. Dual blockade of PD-1 and CTLA-4 combined with tumor vaccine effectively restores T-cell rejection function in tumors. *Cancer Res* 2013; 73:3591-603.